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(54) Title: T CELL EPITOPES OF THE MAJOR ALLERGENS FROM AMBROSIA ARTEMISIIFOLIA

(57) Abstract

The present invention provides isolated peptides of the major protein allergens of Ambrosia artemisiifolia, or short ragweed pollen. Peptides within the scope of the invention comprise at least one T cell epitope, or preferably at least two T cell epitopes of a protein allergen selected from the allergens Amb a I.1, Amb a I.2, Amb a I.3, Amb a I.4 and Amb a II. Modified peptides having similar or enhanced therapeutic properties as the corresponding, naturally-occurring allergen or portion thereof, but having reduced side effects are disclosed. The invention also provides nucleic acids having sequences encoding peptides of the invention. Methods of treatment or of diagnosis of sensitivity to ragweed pollen allergens in an individual and therapeutic compositions comprising one or more peptides of the invention are also provided including multipeptide formulations for human therapeutic use.

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T CELL EPITOPES OF THE MAJOR ALLERGENS FROM AMBROSIA ARTEMISIIFOLIA

Background of the Invention

Ambrosia artemisiifolia or short ragweed pollen is the major cause of late summer hayfever in North America and Canada and considerable effort has been expended in trying to identify the major allergens produced by this species. Amb a I or Antigen E (AgE) has been reported to be the predominant allergen. (King, T.P., et al. Biochemistry, 3:458 (1964)). AgE has been characterized and reported to be a nonglycosylated protein of 38kD molecular mass (King, T.P., Adv. Immunol. 23:77 (1976); King, T.P., et al., Arch. Biochem. Biophys., 212:127 (1981)). An immunochemically related protein, Amb a II (AgK), has been reported to have similar properties (King, T.P., Adv. Immunol. 23:77 (1976); King, T.P., Biochemistry, 11:367 (1972)). Amb a I or AgE can be purified using conventional chromatographic or biochemical techniques. However, it has been reported that due to cleavage of the 38 kD single-chain precursors by the action of a trypsin-like pollen protease. purification often results in the isolation of two noncovalently associated chains of 26 and 12 kD molecular mass, designated a and B, respectively (King, T.P., et al., Arch. Biochem. Biophys., 212:127 (1981); King, T.P., et al., Immunochemistry, 11:83 (1974)). It has been reported that biochemically purified Amb a I has been used as an immunogen to produce murine monoclonal antibodies (mAb) as well as rabbit polyclonal antisera reactive with both the native (Olson, J.R., and D.G. Klapper, J. Immunol, 136:2109 (1986)) and denatured (Smith, et al., Molec. Immunol., 25;355 (1988)) protein.

Early studies using skin tests with Amb a I-depleted pollen extract led to the estimation that at least 90% of the allergenic activity in ragweed pollen can be attributed to Amb a I (King. T.P., et al., Biochemistry, 3:458 (1964); King. T.P., et al., Immunol., 23:77 (1976)). Competition experiments using murine mAb to inhibit the binding of Amb a I by human IgE in ELISA assays have confirmed that Amb a I binds a substantial proportion of human ragweed allergic IgE (Olson, J.R., et al., J. Immunol., 136:2109 (1986)). Recently, three cDNAs encoding proteins with properties of Amb a I were cloned and Amb a I was reported to be a family of homologous, but distinguishable sequences (Rafnar, T., et al., J. Biol. Chem., 266:1229 (1991)). Rafnar and co-workers reported that the individual cloned members of the Amb a I family, designated Amb a I.1, Amb a I.2, and Amb a I.3, share amino acid sequence homology exceeding 80%. The fourth family member designated Amb a I.D (Amb a I.4) is disclosed in U.S.S.N. 07/529,951, filed May 29, 1990. The deduced amino acid sequence of Amb a II has been disclosed and reported to share approximately 65% sequence identity with the Amb a I multigene family of allergens. (Rogers, B. L., et al., Journal of Immunology 147:2547-2552 (1992)).

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Summary of the Invention

The present invention provides isolated peptides of the major protein allergens of Ambrosia artemisiifolia including peptides derived from the family of related proteins, previously designated Amb a IA, Amb a IB, Amb a IC, and Amb a ID. These allergens have been renamed according to the IUIS approved nomenclature as Amb a I.1 (Amb a IA), Amb a I.2 (Amb a IB), Amb a I.3 (Amb a IC) and Amb a I.4 (Amb a ID). Peptides within the scope of the invention comprise at least one T cell epitope, preferably at least two T cell epitopes, of a protein allergen selected from the family of Amb a I allergens and Amb a II. The invention further provides peptides comprising at least two regions, each region comprising at least one T cell epitope of a ragweed pollen allergen. The regions are derived from the same or from different ragweed pollen allergens.

The invention also provides modified peptides having similar or enhanced therapeutic properties as the corresponding, naturally-occurring allergen or portion thereof, but having reduced side effects as well as modified peptides having improved properties such as increased solubility and stability. Peptides of the invention are capable of modifying, in a ragweed pollen-sensitive individual to whom they are administered, the allergic response of the individual to a ragweed pollen allergen. Methods of treatment or of diagnosis of sensitivity to a ragweed pollen allergen in an individual and therapeutic compositions comprising one or more peptides of the invention are also provided and human clinical testing is described.

Brief Description of the Drawings

Fig. 1 shows Western blot analysis of IgE binding to recombinant Amb a 1 proteins.

Fig. 2 is a graphic representation of a direct binding assay of IgE from a single ragweed allergic patient to recombinant Amb a I and Amb a II proteins.

Fig. 3 is a graphic representation of the results of a direct binding assay of IgE from pooled human sera to native Amb a I, Amb a II, recombinant Amb a I.1, recombinant Amb a II and pollen extract.

Fig. 4A and 4B are graphic representations depicting the responses of lymph node cells isolated from mice tolerized *in vivo* with either *Amb a* I.1 or PBS and CFA and challenged *in vitro* with various antigens.

Fig. 5A-5F are graphic representations depicting the responses of lymph node cells isolated from mice tolerized *in vivo* with *Amb a* I.1 or pollen extract, challenged with *Amb a* I.1, and tested with various antigens.

Fig. 6A-6F are graphic representations depicting the responses of lymph node cells isolated from mice tolerized with pollen extract challenged with pollen extract, and tested with various antigens.

Fig. 7 shows various peptides of desired length derived from the Amb a I.1, Amb a I.2 and Amb a I.3 protein allergens.

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Fig. 8 is a graphic representation depicting the responses of T cell lines from 39 patients primed *in vitro* to recombinant *Amb a* I.1 protein and analyzed for response to various overlapping *Amb a* I.1 peptides and selected *Amb a* I.2 and *Amb a* I.3 peptides by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

Fig. 9 shows selected peptides of desired lengths derived from the Amb a 1.1 protein allergen.

Fig. 10 is a graphic representation depicting the responses of T cell lines from 48 patients primed *in vitro* to recombinant *Amb a* I.1 protein and analyzed for response to selected peptides derived from Region 1 of the *Amb a* I.1 protein, by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

Fig. 11 is a graphic representation depicting the responses of T cell lines from 48 patients primed *in vitro* to recombinant *Amb a* I.1 protein and analyzed for response to selected peptides derived from Region 2 of the *Amb a* I.1 protein, by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

Fig. 12 is a graphic representation depicting the responses of T cell lines from 48 patients primed *in vitro* to recombinant *Amb a* I.1 protein and analyzed for response to selected peptides derived from Region 3 of the *Amb a* I.1 protein, by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

Fig. 13 is a graphic representation depicting the responses of T cell lines from 48 patients primed *in vitro* to recombinant *Amb a* I.1 protein and analyzed for response to selected peptides derived from Region 4 of the *Amb a* I.1 protein, by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

Fig. 14 shows selected peptides of desired lengths derived from the Amb a I.1 protein allergen and the Amb a I.3 protein allergen.

Fig. 15 is a graphic representation depicting the responses of T cell lines from 23 patients primed *in vitro* to recombinant *Amb a* 1.1 protein and analyzed for response to selected peptides derived from Region 1 of the *Amb a* 1.1 protein, by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

Fig. 16 is a graphic representation depicting the responses of T cell lines from 23 patients primed *in vitro* to recombinant Amb a I.1 protein and analyzed for response to selected peptides derived from Region 2 of the Amb a I.1 protein, by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

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Fig. 17 is a graphic representation depicting the responses of T cell lines from 23 patients primed *in vitro* to recombinant *Amb a* I.1 protein and analyzed for response to selected peptides derived from Region 3 of the *Amb a* I.1 protein, by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

Fig. 18 is a graphic representation depicting the responses of T cell lines from 23 patients primed *in vitro* to recombinant *Amb a* I.1 protein and analyzed for response to selected peptides derived from Region 4 of the *Amb a* I.1 protein, by percent of positive responses within the individuals tested, the mean stimulation index of positive responses for the peptide and the ranked sum of peptide responses.

Fig. 19 is a graphic representation depicting the responses of T cell lines of 9 patients primed in vitro to recombinant Amb a I.1 protein or recombinant Amb a I.3 protein and analyzed for response to selected peptides derived from Amb a I.1, by percent of positive responses within the individuals tested and the mean stimulation index of positive responses for the peptide.

Fig. 20 is a graphic representation depicting the responses of T cell lines of 9 patients primed in vitro to recombinant Amb a I.1 protein or recombinant Amb a I.3 protein and analyzed for response to selected peptides derived from Amb a I.3, by percent of positive responses within the individuals tested and the mean stimulation index of positive responses for the peptide.

Fig. 21 is a graphic representation of a direct binding assay of IgE from a single ragweed allergic patient to peptides derived from Amb a I.

Fig. 22 is a graphic representation depicting the responses of T cell lines of 28 patients primed in vitro to recombinant Amb a 1.1 protein and analyzed for response to selected peptides derived from Amb a 1.1 by percent of positive responses within the individuals tested, the mean stimulation index of positive responses to the peptide and the ranked sum of peptide responses.

Fig. 23 is a graphic representation depicting the responses of T cell lines of 28 patients primed in vitro to recombinant Amb a 1.1 protein and analyzed for response to selected peptides derived from Region 4 of Amb a 1.1 by percent of positive responses within the individuals tested, the mean stimulation index of positive responses to the peptide and the ranked sum of peptide responses.

Fig. 24 is a graphic representation depicting the responses of T cell lines of 32 patients primed in vitro to recombinant Amb a 1.1 protein and analyzed for response to six selected peptides derived from Amb a 1.1 by percent of positive responses within the individuals tested, the mean stimulation index of positive responses to the peptide and the ranked sum of peptide responses.

Fig. 25 shows various peptides derived from peptide RAE 70.1 which include modifications designed to increase the solubility of the peptide.

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Fig. 26 is a graphic representation depicting the response of a T cell line from patient 956.2 primed in vitro to Fel d I and analyzed for response to various peptides derived from the Amb a I.1 protein.

Fig. 27 is a graphic representation depicting the response of a T cell line from patient 119 primed in vitro with recombinant Amb a I.1 and analyzed for response to various modified peptides derived from Region 2 of the Amb a I.1 protein by tritiated thymidine incorporation.

Fig. 28 is a graphic representation depicting the response of a T cell line from patient 1199_primed in vitro with recombinant Amb a I.1 and analyzed for response to various modified peptides derived from Region 2 of the Amb a I.1 protein by tritiated thymidine incorporation.

Fig. 29 is a graphic representation depicting the response of a T cell clone generated by limiting dilution from an Amb a I.1 specific T cell line stimulated with the AMB 2-10.1 peptide, primed in vitro w/recombinant Amb a I.1 and analyzed for response to various modified peptides derived from Region 2 of the Amb a I.1 protein by tritiated thymidine incorporation.

Fig. 30 is a graphic representation depicting the percent of total histamine release in blood samples from 8 ragweed-allergic patients in response to selected peptides derived from the Amb a I.1 protein.

Fig. 31 is a graphic representation depicting the responses of T cell lines from 39 patients primed in <u>vitro</u> to purified native *Amb a* I.1 protein and analyzed for response to various overlapping *Amb a* I.1 peptides by percent of positive responses within the individuals tested (above the bar), the mean stimulation index of positive responses for the peptide (in parenthesis above the bar) and the ranked sum of peptide responses (X axis).

Detailed Description of the Invention

The present invention provides isolated peptides derived from the major protein allergens of Ambrosia artemisiifolia. As used herein, a peptide refers to an amino acid sequence having fewer amino acids than the entire amino acid sequence of a protein from which the peptide is derived. Peptides of the invention include peptides derived from Amb a I.1. Amb a I.2. Amb a I.3. Amb a I.4 and Amb a II which comprise at least one T cell epitope of the allergen.

Peptides comprising at least two regions, each region comprising at least one T cell epitope of a protein allergen of Ambrosia artemisiifolia are also within the scope of the invention. Each region of such peptides is derived from the same or from different ragweed pollen allergens. Isolated peptides or regions of isolated peptides, each comprising at least two T cell epitopes of a ragweed pollen allergen are particularly desirable for increased therapeutic effectiveness. Peptides which are immunologically related (e.g., by antibody or T cell cross-reactivity) to peptides of the present invention are also within the scope of the

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invention. Peptides immunologically related by antibody cross-reactivity are bound by antibodies specific for a peptide of a protein allergen of *Ambrosia artemisiifolia*. Peptides immunologically related by T cell cross-reactivity are capable of reacting with the same T cells as a peptide of the invention.

The present invention also pertains to a ragweed pollen allergen encoded by a nucleic acid sequence of clone IPC1/5. The full-length nucleic acid sequence of clone IPC1/5 has been determined and the encoded protein has been produced recombinantly in both the pSEM vector (as a fusion protein with \(\beta\)-galactosidase) and the pET11d vector. The recombinant \bar{p} rotein was determined to bind approximately 10-20% of allergic serum IgE on a Western blot. The protein encoded by clone IPC1/5 was found to have a high degree of amino acid sequence homology with cysteine proteinase inhibitors in man and rice. The protein has 66.6% homology with the rice protein oryzacystatin-I. The nucleic acid sequence and deduced amino acid sequence of the allergen encoded by clone IPC1/5 is represented in SEQ ID NO. 11 and 12.

Isolated proteins and isolated peptides of the invention can be produced by recombinant DNA techniques in a host cell transformed with a nucleic acid having a sequence encoding such protein or peptide. The isolated proteins and isolated peptides of the invention can also be produced by chemical synthesis. In certain limited situations, isolated peptides can be produced by chemical cleavage of a protein allergen. When a protein or peptide is produced by recombinant techniques, host cells transformed with a nucleic acid having a sequence encoding the protein or peptide or the functional equivalent of the nucleic acid sequence are cultured in a medium suitable for the cells and protein or peptides can be purified from cell culture medium, host cells, or both using techniques known in the art for purifying proteins and peptides including ion-exchange chromatography, gel filtration chromatography, ultrafiltration, electrophoresis or immunopurification with antibodies specific for the protein or peptide, the protein allergen of Ambrosia artemisiifolia from which the peptide is derived, or a portion thereof. By isolated is meant that protein and peptides of the present invention are substantially free of cellular material or culture medium when produced by recombinant DNA techniques, or substantially free of chemical precursors or other chemicals when synthesized chemically. Recombinant ragweed pollen proteins including recombinant Amb a I.1, Amb a I.2, Amb a I.3, Amb a I.4, and Amb a II have been produced.

Suitable expression vectors for producing recombinant protein and recombinant peptides of the invention include pTRC, pGEX, pMAL, pRIT5, pET11d and pCA. The use of pTRC, pET11d and pGEX as expression vectors will result in expression of ragweed pollen protein as an unfused protein. The use of pMAL, pRIT5, pCA and pSEM as expression vectors will result in expression of ragweed pollen protein fused to maltose E binding protein (pMAL), protein A (pRIT5), truncated protein A (pCA), or \(\beta\)-galactosidase (pSEM). Suitable expression vectors are commercially available. When produced as a

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fusion protein, recombinant ragweed pollen protein can be recovered from the fusion protein through enzymatic or chemical (e.g., cyanogen bromide or dilute acid) cleavage and biochemical purification. For example, enzymatic cleavage sites for Factor Xa or thrombin can be introduced at the fusion junction between the carrier protein (e.g., Protein A) and the ragweed pollen protein. Suitable host cells for expression of recombinant ragweed pollen protein include bacteria, yeast and insect or mammalian cells. Appropriate vectors for expression in yeast include YepSec, pMFa and JRY88. These vectors are also commercially available.

_ To obtain isolated peptides of the present invention, a ragweed pollen allergen is divided into non-overlapping peptides of desired lengths or overlapping peptides of desired lengths as discussed in Example V which may be produced recombinantly, synthetically or in certain limited situations by chemical cleavage of the allergen. Peptides comprising at least one T cell epitope are capable of reducing T cell responsiveness or inducing T cell nonresponsiveness. To determine peptides comprising at least one T cell epitope, isolated peptides are tested by, for example, T cell biology techniques to determine whether the peptides elicit a T cell response or induce T cell non responsiveness. Those peptides found to elicit a T cell response or induce T cell nonresponsiveness are defined as having T cell stimulating activity. As discussed in the Examples, human T cell stimulating activity can be tested by culturing T cells obtained from an individual sensitive to a ragweed pollen allergen, (i.e., an individual who has an IgE mediated immune response to a ragweed pollen allergen) with a peptide derived from the allergen and determining whether proliferation of T cells occurs in response to the peptide as measured, e.g., by cellular uptake of tritiated thymidine. As described in detail in the Examples, stimulation indices for responses by T cells to peptides can be calculated as the maximum CPM in response to a peptide divided by the medium control CPM. As used throughout this application, a peptide comprising at least one T cell epitope, when determined by T cell stimulation requires a stimulation index of at least 2.0. A peptide having a T cell stimulation index of 2.0 is considered useful as a therapeutic agent. Preferred peptides have a stimulation index of at least 2.5, more preferably at least 3.5, and most preferably at least 5.0.

In order to determine precise T cell epitopes by, for example, fine mapping techniques, a peptide having T cell stimulating activity and thus comprising at least one T cell epitope as determined by T cell biology techniques is modified by addition or deletion of amino acid residues at either the amino or carboxy terminus of the peptide and tested to determine a change in T cell reactivity to the modified peptide. If two or more peptides which share an area of overlap in the native protein sequence are found to have human T cell stimulating activity, as determined by T cell biology techniques, additional peptides can be produced comprising all or a portion of such peptides and these additional peptides can be tested by a similar procedure. Following this technique, peptides are selected and produced recombinantly or synthetically. Peptides are selected based on various factors, including the

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strength of the T cell response to the peptide (e.g., stimulation index), the frequency of the T cell response to the peptide in a population of individuals sensitive to ragweed pollen, and the potential cross-reactivity of the peptide with Amb a I family members and Amb a II. The physical and chemical properties of these selected peptides (e.g., solubility, stability) are examined to determine whether the peptides are suitable for use in therapeutic compositions or whether the peptides require modification as described herein. The ability of the selected peptides or selected modified peptides to stimulate human T cells (e.g., induce proliferation, lymphokine secretion) is determined.

In addition, preferred peptides of the invention do not bind immunoglobulin E (IgE) or bind IgE to a substantially lesser extent (i.e. preferably at least 100 fold and more preferably at least 1000 fold less) than the protein allergen from which the peptide is derived binds IgE. Recombinant ragweed pollen allergens including recombinant Amb a 1.1, Amb a I.2, Amb a I.3, Amb a I.4, and Amb a II have been produced and shown to have reduced IgE binding activity as compared to the corresponding native protein allergen (See Fig. 3). The major complications of standard immunotherapy are IgE-mediated responses such as anaphylaxis. Immunoglobulin E is a mediator of anaphylactic reactions which result from the binding and cross-linking of antigen to IgE on mast cells or basophils and the release of mediators (e.g., histamine, serotonin, eosinophil chemotacic factors). Thus, anaphylaxis in a substantial percentage of a population of individuals sensitive to ragweed pollen allergen could be avoided by the use in immunotherapy of a recombinant protein, a peptide or peptides which do not bind IgE in a substantial percentage (e.g., at least about 75%) of a population of individuals sensitive to ragweed pollen allergen, or if the protein or peptide(s) binds IgE, such binding does not result in the release of mediators from mast cells or basophils. Additionally, the risk of anaphylaxis can be reduced by the use in immunotherapy of a recombinant protein, a peptide or peptides which have reduced IgE binding. Moreover, recombinant protein or peptides which have minimal IgE stimulating activity are desirable for therapeutic effectiveness. Minimal IgE stimulating activity refers to IgE production that is less than the amount of IgE production and/or IL-4 production stimulated by the native protein allergen (e.g., Amb a I.1).

A peptide or recombinant protein of the invention, when administered to a ragweed pollen-sensitive individual, is capable of modifying the allergic response of the individual to the allergen. Particularly, peptides of the invention comprising at least one T cell epitope of a ragweed pollen allergen or at least two regions derived from a ragweed pollen allergen each comprising at least one T cell epitope, when administered to a ragweed pollen-sensitive individual are capable of modifying the T cell response of the individual to the allergen. As used herein, modification of the allergic response of a ragweed pollen-sensitive individual to a ragweed pollen allergen can be defined as non-responsiveness or diminution in symptoms to a ragweed pollen allergen, as determined by standard clinical procedures (see e.g., Varney et al., British Medical Journal 302: 265-269 (1990)).

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As a result of the work described herein, peptides derived from ragweed pollen allergens comprising at least one T cell epitope have been produced. T cell epitopes are believed to be involved in initiation and perpetuation of the immune response to ragweed pollen allergen(s) which are responsible for the clinical symptoms of ragweed pollen allergy. These T cell epitopes are thought to trigger early events at the level of the T helper cell by binding to an appropriate HLA molecule on the surface of an antigen presenting cell and - stimulating the relevant T cell subpopulation. These events lead to T cell proliferation, lymphokine secretion, local inflammatory reactions, the recruitment of additional immune cells to the site, and activation of the B cell cascade leading to production of antibodies. One isotype of these antibodies, IgE, is fundamentally important in the development of allergic symptoms and its production is influenced early in the cascade of events, at the level of the T helper cell, by the nature of the lymphokines secreted. A T cell epitope is the basic element or smallest unit of recognition by a T cell receptor, where the epitope comprises amino acid residues essential to receptor recognition which may be contiguous and/or non-contiguous in the amino acid sequence of the protein. Amino acid sequences which mimic those of T cell epitopes and which modify the allergic response to protein allergens of Ambrosia artemisiifolia are within the scope of this invention.

Exposure of ragweed pollen allergic patients to peptides of the present invention, in a non-immunogenic form, may induce T cell non-responsiveness of appropriate T cell subpopulations such that they become non-responsive to ragweed pollen allergen(s) and do not participate in mounting an immune response upon such exposure.

While not intending to be limited to any theory, it is believed that T cell nonresponsiveness (which includes reduced T cell responsiveness) is induced as a result of not providing an appropriate costimulatory signal sometimes referred to as a "second signal" Briefly, it is believed that stimulation of T cells requires two types of signals, the first is the recognition by the T cell via the T cell receptor of appropriate MHC-associated processed antigens on antigen presenting cells (APCs) and the second type of signal is referred to as a costimulatory signal(s) or "second signal" which may be provided by certain competent APCs. When a composition of the invention is administered without adjuvant, it is believed that competent APCs which are capable of producing the second signal or costimulatory signal are not engaged in the stimulation of appropriate T cells therefore resulting in T cell nonresponsiveness or reduced T cell responsiveness. In addition, there are a number of antibodies or other reagents capable of blocking the delivery of costimulatory signals such as the "second signal" which include, but are not limited to B7 (including B7-1, B7-2, and BB-1), CD28, CTLA4, CD40 CD40L CD54 and CD11a/18 (Jenkins and Johnson, Current Opinion in Immunology, 5:361-367 (1993), and Clark and Ledbetter, Nature, 367:425-428 (1994)) Thus, a peptide of the invention may be administered in nonimmunogenic form as discussed above, in conjunction with a reagent

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capable of blocking costimulatory signals such that the level of T cell nonresponsiveness is enhanced.

In addition, administration of a peptide of the present invention may modify the lymphokine secretion profile as compared with exposure to the naturally-occurring ragweed pollen allergen or portion thereof (e.g., result in a decrease of IL-4 and/or an increase in IL-2). Furthermore, exposure to a peptide of the invention may influence T cell subpopulations which normally participate in the response to ragweed pollen allergen(s) such that these T cells are drawn away from the site(s) of normal exposure to the allergen (e.g., nasal mucosa, skin, and lung) towards the site(s) of therapeutic administration of the peptide. This redistribution of T cell subpopulations may ameliorate or reduce the ability of an individual's immune system to stimulate the immune response at the site of normal exposure to the ragweed pollen allergen(s), resulting in a diminution in allergic symptoms.

Isolated peptides of the invention comprise at least one T cell epitope of a protein allergen of Ambrosia artemisiifolia (i.e., the peptide comprises at least approximately seven amino acid residues of the protein allergen). For purposes of therapeutic effectiveness, therapeutic compositions of the invention preferably comprise at least two T cell epitopes of a ragweed pollen allergen. Accordingly, isolated peptides of the invention preferably comprise at least two T cell epitopes (i.e., the peptide comprises at least approximately eight amino acid residues, and preferably fifteen amino acid residues). Additionally, isolated peptides of the invention preferably comprise a sufficient percentage of the T cell epitopes of the entire protein allergen such that upon administration of the peptide to an individual sensitive to ragweed pollen, T cells of the individual are rendered non-responsive to the protein allergen. Isolated peptides of the invention comprising up to approximately 45 amino acid residues in length, and most preferably up to approximately 30 amino acid residues in length are particularly desirable as increases in length may result in difficulty in peptide synthesis as well as retention of an undesirable property (e.g., immunoglobulin binding or enzymatic activity) due to maintenance of conformational similarity between the peptide and the protein allergen from which it is derived. All of the peptides shown in Fig. 8 were found to have human T cell stimulating activity.

To determine whether a peptide (candidate peptide) or a combination of candidate peptides are likely contain a sufficient percentage of T cell epitopes of ragweed protein antigen to induce non-responsiveness in a substantial percentage of a population of individuals sensitive to the protein antigen, an algorithm can be used. In accordance with one such algorithm, a human T cell stimulation index (discussed above) for the peptide(s) in an *in vitro* T cell proliferation assay is calculated for each individual tested in a population of individuals sensitive to ragweed protein allergen. The remaining peptides in the *in vitro* T cell proliferation assay are overlapping peptides (overlapping by between about 5 - 10 amino acid residues) which cover the remainder of the protein not covered by the candidate peptide(s), which peptides are at least 12 amino acids long and which are preferably no longer than 30

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and more preferably no longer than 25 amino acid residues in length. A human T cell stimulation index for each such remaining peptide in the set of peptides produced in the *in vitro* T-cell proliferation assay with T-cells obtained from each individual in the population of individuals tested is calculated and added together. For each individual, the human T cell stimulation index for the candidate peptide(s) is divided by the sum of the human T cell stimulation indices of the remaining peptides in the set of peptides tested to determine a percent. This percent is obtained for at least twenty (20) and preferably at least thirty (30) individuals sensitive to the protein antigen of interest and a mean percent is determined (the percentage of positive T cell responses (S.I. greater than or equal to 2.0) in response to the candidate peptide or combination of candidate peptides). A mean percent of about 10% or greater for the candidate peptide(s) together with a percent positive of at least about 60%, preferably about 75% and more preferably about 90%, or most preferably 100%, indicates that the candidate peptide(s) selected is likely to contain a sufficient percentage of T cell epitopes to induce T cell non responsiveness in a substantial percentage of a population of individuals sensitive toTRFP.

Preferred peptides comprise all or a portion of the areas of major T cell reactivity within the Amb a I.1 protein allergen, i.e., Region 1, Region 2, Region 3 and Region 4. Each area is broadly defined as follows: Region 1 comprises amino acid residues 48-107; Region 2 comprises amino acid residues 171-216; Region 3 comprises amino acid residues 278-322; and Region 4 comprises amino acid residues 331-377. Preferred areas of major T cell reactivity within each Region comprise: amino acid residues 57-101; amino acid residues 182-216; amino acid residues 280-322; and amino acid residues 342-377. Similar areas of major T cell reactivity can be found within the other Amb a I family members (i.e., Amb a 1.2. Amb a 1.3 and Amb a 1.4), and Amb a 11. As shown in Example VIII, the Amb a 1 protein allergens and Amb a II demonstrate a high degree of T cell cross-reactivity. Given this crossreactivity, shared areas of major T cell reactivity and shared T cell epitopes are likely to be found in conserved regions between Amb a I and the remaining Amb a I family members and Amb a II. For example, Amb a I.1 stimulated T cells have been shown to recognize both Amb a I.1 derived peptides and homologous Amb a I.3 derived peptides (See Example IX). Similarly, Amb a 1.3 stimulated T cell recognize both Amb a 1.1 and Amb a 1.3 derived peptides.

Preferred ragweed pollen peptides comprise all or a portion of the following peptides: RAE 67.1 (SEQ ID NO:13); RAE 57.1 (SEQ ID NO:14); RAE 24.E (SEQ ID NO:15); RAE 24.1 (SEQ ID NO:16); RAE 22.E (SEQ ID NO:17); RAE 22.E-1 (SEQ ID NO:18); RAE 3.D (SEQ ID NO:19); RAE 3.1 (SEQ ID NO:20); RAE 22.E-2 (SEQ ID NO:21); RAE 5.D (SEQ ID NO:22); RAE 6.D (SEQ ID NO:23); RAE 6.1 (SEQ ID NO:24); RAE 7.D (SEQ ID NO:25); RAE 7.D-1 (SEQ ID NO:26); RAE 40.1-6 (SEQ ID NO:27); RAE 40.1-5 (SEQ ID NO:28); RAE 40.1-4 (SEQ ID NO:29); RAE 40.D (SEQ ID NO:30); RAE 40.1 (SEQ ID NO:31); RAE 61.1 (SEQ ID NO:32); RAE 80.1 (SEQ ID NO:33); RAE 45.1 (SEQ ID

NO:34); RAE 75.1 (SEQ ID NO:35); RAE 62.1 (SEQ ID NO:36); RAE 69.1 (SEQ ID NO:37); RAE 69.1-1 (SEQ ID NO:38); RAE 69.1-2 (SEQ ID NO:39); RAE 69.1-3 (SEQ ID NO:40); RAE 70.1-3 (SEQ ID NO:41); RAE 70.1-2 (SEQ ID NO:42); RAE 70.1-1 (SEQ ID NO:43); RAE 70.1 (SEQ ID NO:44); RAE 71.1 (SEQ ID NO:45); RAE 65.1 (SEQ ID NO:46); RAE 63.1 (SEQ ID NO:47); RAE 76.1 (SEQ ID NO:48); RAE 27.1 (SEQ ID 5 NO:49); RAE 66.1 (SEQ ID NO:50); RAE 66.1-1 (SEQ ID NO:51); RAE 66.1-2 (SEQ ID NO:52); RAE 66.1-3 (SEQ ID NO:53); RAE 64.1-3 (SEQ ID NO:54); RAE 64.1-2 (SEQ ID NO:55); RAE 64.1-1 (SEQ ID NO:56); RAE 64.1 (SEQ ID NO:57); RAE 73.1 (SEQ ID NO:58); RAE 74.1 (SEQ ID NO:59); RAE 74.1-1 (SEQ ID NO:60); RAE 29.1 (SEQ ID NO:61); RAE 29.1-1 (SEQ ID NO:62); RAE 28+29 (SEQ ID NO:63); RAE 29.1-2 (SEQ ID 10 NO:64); RAE 29.1-3 (SEQ ID NO:65); RAE 29.1-4 (SEQ ID NO:66); RAE 28.1-3 (SEQ ID NO:67); RAE 28.1-2 (SEQ ID NO:68); RAE 28.1-1 (SEQ ID NO:69); RAE 28.1 (SEQ ID NO:70); RAE 20.1 (SEQ ID NO:71); RAE 20.1-3 (SEQ ID NO:72); RAE 20.1-2 (SEQ ID NO:73); RAE 20.1-1 (SEQ ID NO:74); RAE 21.1 (SEQ ID NO:75); RAE 17.1 (SEQ ID NO:76); RAE 55.1 (SEQ ID NO:77); RAE 76.6 (SEQ ID NO:78); RAE 67.15 (SEQ ID 15 NO:79); RAE 45.15 (SEQ ID NO:80); RAE 27.15 (SEQ ID NO:81); AMB 1-1.1 (SEQ ID NO:85); AMB 1-2.1 (SEQ ID NO:86); AMB 1-3.1 (SEQ ID NO:87); AMB 1-4.1 (SEQ ID NO:84); AMB 1-5.1 (SEQ ID NO:83); AMB 1-6.1 (SEQ ID NO:82); AMB 1-4.15 (SEQ ID NO:88); AMB 1-2.15 (SEQ ID NO:89); AMB 2-4.1 (SEQ ID NO:90); AMB 2-3.1 (SEQ ID NO:91); AMB 2-5.1 (SEQ ID NO:92); AMB 2-6.1 (SEQ ID NO:93); AMB 2-2.1 (SEQ ID 20 NO:94); AMB 2-1.1 (SEQ ID NO:95); AMB 2-7.1 (SEQ ID NO:96); AMB 2-8.1 (SEQ ID NO:97); AMB 2-9.1 (SEQ ID NO:98); AMB 2-10.1 (SEQ ID NO:99); AMB 2-11.1 (SEQ ID NO:100); AMB 2-1.15 (SEQ ID NO:101); AMB 3-4.1 (SEQ ID NO:103); AMB 3-5.1 (SEQ ID NO:102); AMB 3-3.1 (SEQ ID NO:104); AMB 3-2.1 (SEQ ID NO:105); AMB 3-1.1 (SEQ ID NO:106); AMB 3-4.15 (SEQ ID NO:107); AMB 3-1.15 (SEQ ID NO:108); AMB 4-8.1 (SEQ ID NO:109); AMB 4-9.1 (SEQ ID NO:110); AMB 4-6.1 (SEQ ID NO:111); AMB 4-5.1 (SEQ ID NO:112); AMB 4-3.1 (SEQ ID NO:113); AMB 4-2.1 (SEQ ID NO:114); AMB 4-1.1 (SEQ ID NO:115); AMB 4-3.15 (SEQ ID NO:116); Amb 2-18.1 (SEQ ID NO:126); Amb 2-19.1 (SEQ ID NO:127); Amb 2-20.1 (SEQ ID NO:128); Amb 2-21.1 (SEQ ID NO:129); Amb 2-22.1 (SEQ ID NO:130); Amb 2-23.1 (SEQ ID NO:131); Amb 2-30 26.1 (SEQ ID NO:132); Amb 28.1 (SEQ ID NO:133); Amb 2-30.1 (SEQ ID NO:134); Amb 2-32.1 (SEQ ID NO:135); Amb 2-33.1 (SEQ ID NO:136); Amb 2-34.1 (SEQ ID NO:137); Amb 2-35.1 (SEQ ID NO:138); Amb 2-36.1 (SEQ ID NO:139); Amb 2-37.1 (SEQ ID NO:140); Amb 2-38.1 (SEQ ID NO:141); AMB 4-9.1EP (SEQ ID NO:142); AMB 4-9.1NP (SEQ ID NO:143); AMB 4-9.1AP (SEQ ID NO:144); AMB 4-9.1SP (SEQ ID NO:145); 35 AMB 4-9.1QP (SEQ ID NO:146); AMB 4-9.1DA (SEQ ID NO:147); amb 4-9.1DS (SEQ ID NO:148); AMB 4-9.1DG (SEQ ID NO:149); and RA-02.1 (SEQ ID NO:150), the amino acid sequences of such peptides being shown in Figs. 7, 14, 23, 24 and 25. Particularly preferred peptides comprise all or a portion of the following peptides: AMB 1-2.1 (SEQ ID NO:86);

AMB 2-6.1 (SEQ ID NO:93); AMB 2-4.1 (SEQ ID NO:90); Amb 2-36.1 (SEQ ID NO:139); Amb 2-38.1 (SEQ ID NO:141); RA-02.1 (SEQ ID NO:150); AMB 2-9.1 (SEQ ID NO:98); AMB 3-5.1 (SEQ ID NO:102); and AMB 4-9.1 (SEQ ID NO:110).

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Another embodiment of the present invention provides peptides comprising at least two regions, each region comprising at least one T cell epitope of a protein allergen of Ambrosia artemisiifolia (e.g., each region comprises at least approximately seven amino acid residues). These peptides comprising at least two regions can comprise as many amino acid residues as desired and preferably comprise at least about 7, more preferably at least about 15, even more preferably about 30 and most preferably at least about 40 amino acid residues of a ragweed pollen allergen. Each region of such peptide preferably comprises up to 45 amino acid residues in length, more preferably up to 40 residues in length and most preferably up to 30 amino acid residues in length as increases in length of a region may result in difficulty in peptide synthesis as well as retention of an undesirable property (e.g., immunoglobulin binding or enzymatic activity) due to maintenance of conformational similarity between the peptide and the protein allergen from which it is derived. If desired. the amino acid sequences of the regions can be produced and joined by a linker to increase sensitivity to processing by antigen-presenting cells. Such linker can be any non-epitope amino acid sequence or other appropriate linking or joining agent. To obtain preferred peptides comprising at least two regions, each comprising at least one T cell epitope, the regions are arranged in a configuration different from a naturally-occurring configuration of the regions in the allergen. For example, the regions containing T cell epitope(s) can be arranged in a noncontiguous configuration and can preferably be derived from the same protein allergen. Noncontiguous is defined as an arrangement of regions containing T cell epitope(s) which is different than that of an amino acid sequence present in the protein allergen from which the regions are derived. Furthermore, the noncontiguous regions containing T cell epitopes can be arranged in a nonsequential order (e.g., in an order different from the order of the amino acids of the native protein allergen from which the region containing T cell epitope(s) are derived in which amino acids are arranged from an amino terminus to a carboxy terminus). A peptide can comprise at least 15%, at-least 30%, at least 50% or up to 100% of the T cell epitopes of a ragweed pollen allergen.

The individual peptide regions can be produced and tested to determine which regions bind immunoglobulin E specific for a ragweed pollen allergen and which of such regions would cause the release of mediators (e.g., histamine) from mast cells or basophils. Those peptide regions found to bind immunoglobulin E and cause the release of mediators from mast cells or basophils in greater than approximately 10-15% of the allergic sera tested are preferably not included in the peptide regions arranged to form peptides of the invention.

Preferred peptides of the invention comprise two or more regions derived from the same or from different ragweed pollen allergens (e.g., Amb a I.1, Amb a I.2, Amb a I.3, Amb a I.4 and Amb a II). For example, one region can be derived from Amb a I.1 and one region

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can be derived from $Amb\ a\ I.2$; one region can be derived from $Amb\ a\ I.1$ and one region can be derived from $Amb\ a\ I.1$ and one region can be derived from $Amb\ a\ I.2$ and one region can be derived from $Amb\ a\ I.2$ and one region can be derived from $Amb\ a\ I.2$ and one region can be derived from $Amb\ a\ I.2$ and one region can be derived from $Amb\ a\ I.3$; one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.1$ and one region can be derived from $Amb\ a\ I.1$ and one region can be derived from $Amb\ a\ I.2$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and one region can be derived from $Amb\ a\ I.3$ and $Amb\ a\ I.3$

Regions of a peptide of the invention preferably comprise all or a portion of Region 1, Region 2, Region 3 and Region 4 of Amb a I.1, and the above discussed preferred areas of major T cell reactivity within each Region. If Region 1, 2, 3 or 4 is found to bind IgE and cause the release of mediators from mast cells or basophils, then it is preferred that more than one region of the peptide comprise such Region and that the various regions of the peptide do not bind IgE or cause release of mediators from mast cells or basophils. Examples of preferred regions include: AMB 1-1.1 (SEQ ID NO:85); AMB 1-2.1 (SEQ ID NO:86); AMB 1-3.1 (SEQ ID NO:87); AMB 1-4.1 (SEQ ID NO:84); AMB 1-5.1 (SEQ ID NO:83); AMB 1-6.1 (SEQ ID NO:82); AMB 1-4.15 (SEQ ID NO:88); AMB 1-2.15 (SEQ ID NO:89); AMB 2-4.1 (SEQ ID NO:90); AMB 2-3.1 (SEQ ID NO:91); AMB 2-5.1 (SEQ ID NO:92); AMB 2-6.1 (SEQ ID NO:93); AMB 2-2.1 (SEQ ID NO:94); AMB 2-1.1 (SEQ ID NO:95); AMB 2-7.1 (SEQ ID NO:96); AMB 2-8.1 (SEQ ID NO:97); AMB 2-9.1 (SEQ ID NO:98); AMB 2-10.1 (SEQ ID NO:99); AMB 2-11.1 (SEQ ID NO:100); AMB 2-1.15 (SEQ ID NO:101); AMB 3-4.1 (SEQ ID NO:103); AMB 3-5.1 (SEQ ID NO:102); AMB 3-3.1 (SEQ ID NO:104); AMB 3-2.1 (SEQ ID NO:105); AMB 3-1.1 (SEQ ID NO:106); AMB 3-4.15 (SEQ ID NO: 107); AMB 3-1.15 (SEQ ID NO:108); AMB 4-8.1 (SEQ ID NO:109); AMB 4-9.1 (SEQ ID NO:110); AMB 4-6.1 (SEQ ID NO:111); AMB 4-5.1 (SEQ ID NO:112); AMB 4-3.1 (SEQ ID NO:113); AMB 4-2.1 (SEQ ID NO:114); AMB 4-1.1 (SEQ ID NO:115); AMB 4-3.15 (SEQ ID NO:116); RA-02.1 (SEQ ID NO:150); Amb 2-36.1 (SEQ ID NO:139); and Amb 2-38.1 (SEQ ID NO:141), the amino acid sequences of such regions being shown in Fig. 14, Fig. 24 or Fig. 25, or portions of said regions comprising at least one T cell epitope.

Preferred peptides comprise various combinations of two or more regions, each region comprising all or a portion of Region 1, Region 2, Region 3 or Region 4 of *Amb a* 1.1. Preferred peptides comprise various combinations of two or more regions, each region having an amino acid sequence as shown in Fig. 14, such combination of regions including the following: AMB 4-6.1 and RAE 70.1 (SEQ ID NO:111 and SEQ ID NO:44); AMB 4-6.1 and AMB 2-5.1 (SEQ ID NO:111 and SEQ ID NO:92); AMB 4-9.1 and AMB 2-5.1 (SEQ ID NO:92); AMB 4-9.1 and SEQ ID NO:92); AMB 4-9.1 and SEQ ID NO:910 and SEQ ID

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NO:44); AMB 4-6.1, AMB 2-5.1 and AMB 1-2.1 (SEQ ID NO:111, SEQ ID NO:92 and SEQ ID NO:86); AMB 4-9.1, AMB 2-5.1 and AMB 1-2.1 (SEQ ID NO:110, SEQ ID NO:92 and SEQ ID NO:86); AMB 4-6.1, RAE 70.1 and AMB 1-2.1 (SEQ ID NO:111, SEQ ID NO:44 and SEQ ID NO:86); AMB 4-9.1, RAE 70.1 and AMB 1-2.1 (SEQ ID NO:110, SEQ ID NO:44 and SEQ ID NO:86); AMB 4-6.1, RAE 70.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:111, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:102); AMB 4-9.1, RAE 70.1. AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:110, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:102); AMB 4-6.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:111, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:102); AMB 4-9.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:110, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:102); AMB 10 4-6.1, RAE 70.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:111, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:103); AMB 4-9.1, RAE 70.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:110, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:103); AMB 4-6.1, AMB 2-5.1. AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:111, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:103); AMB 4-9.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:110, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:103); AMB 2-1.15 and AMB 4-3.15 (SEQ ID NO:101, and SEQ ID NO:116); AMB 1-2.15, AMB 2-1.15 and AMB 4-3.15 (SEQ ID NO:89, SEQ ID NO:101, and SEQ ID NO:116); and AMB 1-2.15, AMB 2-1.15, AMB 4-3.15 and AMB 3-4.15 (SEQ ID NO:89, SEQ ID NO:101, SEQ ID NO:116 and SEQ ID NO:107).

Peptides of protein allergens of Ambrosia artemisiifolia within the scope of the invention can be used in methods of treating and preventing allergic reactions to ragweed pollen allergens. Thus, one aspect of the present invention provides therapeutic compositions comprising a peptide of Amb a I.1, Amb a I.2, Amb a I.3, Amb a I.4 or Amb a II including at least one T cell epitope, or preferably at least two T cell epitopes, and a pharmaceutically acceptable carrier or diluent. In another aspect, the therapeutic composition comprises a pharmaceutically acceptable carrier or diluent and a peptide comprising at least two regions, each region comprising at least one T cell epitope of a ragweed pollen allergen and is derived from the same or from different ragweed pollen allergens.

Administration of the therapeutic compositions of the present invention to desensitize an individual can be carried out using known techniques. For example, a peptide derived from a ragweed pollen allergen comprising at least one T cell epitope can be administered in combination with an appropriate diluent, or carrier. Preferably, peptides are administered in soluble form. Pharmaceutically acceptable diluents include saline and aqueous buffer solutions. Pharmaceutically acceptable carriers include polyethylene glycol (Wie et al., International Archives of Allergy and Applied Immunology 64: 84-99 (1981)) and liposomes (Strejan et al., Journal of Neuroimmunology 7: 27 (1984)). For purposes of inducing T-cell non-responsiveness (or reduced T cell responsiveness) in an individual, the therapeutic composition is preferably administered in non-immunogenic form, e.g., one that does not

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include adjuvant. Such compositions will generally be administered by injection (e.g., intravenous, subcutaneous, intramuscular), oral administration, (e.g., as in the form of a capsule), inhalation, transdermal application or rectal administration. Preferably, therapeutic compositions are administed subcutaneously.

In addition, isolated and purified native ragweed pollen protien allergens (see, Example 1 for isolation and purification of ragweed pollen protein antigens) or portions thereof, can be administered orally.

The therapeutic compositions of the invention are administered to ragweed pollensensitive individuals at dosages and for lengths of time effective to reduce sensitivity (i.e., reduce the allergic response) of an individual to a ragweed pollen allergen. A therapeutically effective amount of one or more of the same or of different therapeutic compositions can be administered simultaneously or sequentially to a ragweed pollen-sensitive individual. Effective amounts of the therapeutic compositions will vary according to factors such as the degree of sensitivity of the individual to ragweed pollen allergens, the age, sex, and weight of the individual, and the ability of the peptide to stimulate a T cell response in the individual.

For subcutaneous injection of one or more therapeutic compositions of the invention, preferably about 1 mg- 3 mg and more preferably from about 20mg-1.5 mg, and even more preferably about 50 mg- 750 mg of each active component (peptide) per dosage unit may be administered. It is especially advantageous to formulate parenteral compositions in unit dosage form for ease of administration and uniformity of dosage. Unit dosage form as used herein refers to physically discrete units suited as unitary dosages for human subjects to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the desired pharmaceutical carrier. The specification for the novel unit dosage forms of the invention are dictated by and directly dependent on (a) the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active compound for the treatment of human subjects.

Dosage regimen may be adjusted to provide the optimum therapeutic response. For example, several divided doses may be administered over the course of days, weeks, months or years, or the dose may be proportionally increased or reduced with each subsequent injection as indicated by the exigencies of the therapeutic situation. In one preferred therapeutic regimen, subcutaneous injections of therapeutic compositions are given once a week for 3-6 weeks. The dosage may remain constant for each injection or may increase or decrease with each subsequent injection. A booster injection may be administered at intervals of about three months to about one year after initial treatment and may involve only a single injection or may involve another series of injections similar to that of the initial treatment.

To administer a composition of the invention by other than parenteral administration, (i.e. oral administration) it may be necessary to coat the composition with, or co-administer

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the composition with, a material to prevent its inactivation or enhance its absorption and bioavailability. For example, a peptide formulation may be co-administered with enzyme inhibitors or in liposomes. Enzyme inhibitors include pancreatic trypsin inhibitor, diisopropylfluorophosphate (DEP) and trasylol. Liposomes include water-in-oil-in-water CGF emulsions as well as conventional liposomes (Strejan et al., (1984) J. Neuroimmunol. 7:27). When a peptide is suitably protected, the peptide may be orally administered, for example, with an inert diluent or an assimilable edible carrier. The peptide and other ingredients may also be enclosed in a hard or soft shell gelatin capsule, compressed into tablets, or incorporated directly into the individual's diet. For oral therapeutic administration, the active compound may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, solutions, gels, suspensions, syrups, wafers, and the like. Such compositions and preparations should contain at least 1% by weight of active compound. The percentage of the composition and preparations may, of course, be varied and may conveniently be between about 5 to 80% of the weight of the unit. The amount of active compound in such therapeutically useful compositions is such that a suitable dosage will be obtained. In addition, the active compound may be incorporated into sustained-release or controlled release (steady state or pulsatile release) preparations and formulations.

In yet another aspect of the present invention, a composition is provided comprising at least two peptides (e.g., a physical mixture of at least two peptides), each comprising at least one T cell epitope of a protein allergen of Ambrosia artemisiifolia. Compositions comprising several peptides or combinations of separate peptides can include as many peptides as desired (e.g., 5, 6, 7...) for therapeutic efficacy. The peptides are derived from the same or from different ragweed pollen allergens. Such compositions can be administered in the form of a therapeutic composition with a pharmaceutically acceptable carrier or diluent. Preferably, peptides are administered in soluble form. A therapeutically effective amount of one or more of such compositions can be administered simultaneously or sequentially, preferably subcutaneously, to a ragweed pollen-sensitive individual to desensitize or tolerize the individual to ragweed pollen.

Preferred compositions and preferred combinations of peptides which can be administered simultaneously or sequentially (comprising peptides having amino acid sequences shown in Fig. 14) include the following combinations: AMB 4-6.1 and RAE 70.1 (SEQ ID NO:111 and SEQ ID NO:44); AMB 4-6.1 and AMB 2-5.1 (SEQ ID NO:111 and SEQ ID NO:92); AMB 4-9.1 and AMB 2-5.1 (SEQ ID NO:110 and SEQ ID NO:92); AMB 4-9.1 and RAE 70.1 (SEQ. ID NO:110 and SEQ ID NO:44); AMB 4-6.1, AMB 2-5.1 and AMB 1-2.1 (SEQ ID NO:111, SEQ ID NO:92 and SEQ ID NO:86); AMB 4-9.1, AMB 2-5.1 and AMB 1-2.1 (SEQ ID NO:110, SEQ ID NO:92 and SEQ ID NO:86); AMB 4-6.1, RAE 70.1 and AMB 1-2.1 (SEQ ID NO:111, SEQ ID NO:44 and SEQ ID NO:86); AMB 4-9.1, RAE 70.1 and AMB 1-2.1 (SEQ ID NO:110, SEQ ID NO:44 and SEQ ID NO:86); AMB 4-9.1,

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6.1, RAE 70.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:111, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:102); AMB 4-9.1, RAE 70.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:110, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:102); AMB 4-6.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:111, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:102); AMB 4-9.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:110, SEQ ID 5 NO:92, SEQ ID NO:86 and SEQ ID NO:102); AMB 4-6.1, RAE 70.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:111, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:103); AMB 4-9.1, RAE 70.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:110, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:103); AMB 4-6.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID 10 NO:111, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:103); AMB 4-9.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:110, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:103); AMB 2-1.15 and AMB 4-3.15 (SEQ ID NO:101, and SEQ ID NO:116); AMB 1-2.15, AMB 2-1.15 and AMB 4-3.15 (SEQ ID NO:89, SEQ ID NO:101, and SEQ ID NO:116); and AMB 1-2.15, AMB 2-1.15, AMB 4-3.15 and AMB 3-4.15 (SEQ ID NO:89, SEQ ID NO:101, SEQ ID NO:116 and SEQ ID NO:107). 15

Particularly preferred compositions and preferred combinations of peptides for therapeutic administration include the following combinations: AMB 1-2.1 and AMB 4-9.1 (SEQ ID NO:86 and SEQ ID NO:110); AMB 1-2.1, Amb 2-38.1 and AMB 4-9.1 (SEQ ID NO:86, SEQ ID NO:141 and SEQ ID NO:110); AMB 1-2.1, Amb 2-38.1, AMB 4-9.1 and AMB 2-4.1 (SEQ ID NO:86, SEQ ID NO:141, SEQ ID NO:110 and SEQ ID NO:90); AMB 1-2.1, Amb 2-38.1, AMB 4-9.1, AMB 2-4.1 and AMB 3-5.1 (SEQ ID NO:86, SEQ ID NO:141, SEQ ID NO:110, SEQ ID NO:90, and SEQ ID NO:102); AMB 1-2.1, Amb 2-36.1 and AMB 4-9.1 (SEQ ID NO:86, SEQ ID NO:139 and SEQ ID NO:139, SEQ ID NO:110 and SEQ ID NO:90); and AMB 1-2.1, Amb 2-36.1, AMB 4-9.1 and AMB 1-2.1, Amb 2-36.1, AMB 4-9.1, AMB 2-4.1 and AMB 3-5.1 (SEQ ID NO:90); and AMB 1-2.1, Amb 2-36.1, AMB 4-9.1, AMB 2-4.1 and AMB 3-5.1 (SEQ ID NO:86, SEQ ID NO:102).

Another aspect of this invention pertains to a multipeptide formulation suitable for pharmaceutical administration to ragweed sensitive individuals. The multipeptide formulation includes at least two or more peptides of ragweed pollen protein allergen having human T cell stimulating activity in an *in vitro* T cell proliferation assay (i.e., comprising at least one T cell epitope). Special considerations when preparing a multipeptide formulation include maintaining the solubility and stability of all peptides in the formulation at a physiologically acceptable pH. This requires choosing one or more pharmaceutically acceptable carriers such as excipients which are compatible with all the peptides in the multipeptide formulation. For example, suitable excipients include sterile water, sodium phosphate, mannitol or both sodium phosphate and mannitol or any combination thereof. Additionally due to the potential for dimerization of the peptides in a multipeptide formulation, there may also be included an agent such as EDTA to prevent dimerization. Alternatively, any material or procedures known in the art to prevent dimerization may be

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used. A preferred multipeptide formulation includes at least one first peptide and at least one second peptide of ragweed pollen protein each having human T cell stimulating activity and soluble at a physiologically acceptable pH and selected from the group of peptides. In a preferred embodiment, the multipeptide formulation includes Peptides Amb 1-2.1, Amb 4-9.1, Amb 2-36.1, and modifications thereof, and sodium phosphate and mannitol. In this embodiment, it is preferred that Peptides Amb 1-2.1, Amb 4-9.1, and Amb 2-36.1 are in the form of a lyophilized powder which is reconstituted in a physiologically acceptable carrier, such as sterile water, prior to use. As an illustrative example, a multipeptide formulation comprising the three peptides were produced and used in Phase I human clinical trials (see Example XV). The peptides were combined during manufacturing to produce a vial containing a sterile, pyrogen free, lyophilized powder having the following composition:

Active: Peptide Amb 1-2.1, Peptide Amb 4-9.1, and Peptide Amb 2-36.1

In concentration of 7.5-1500 µg per peptide

Inactives: 0.05 M Sodium Phosphate pH 7.5

5% w/v Mannitol, U.S.P.

Diluent: Sterile Water for Injection, U.S.P. (initial reconstitution)

0.9% Sodium Chloride for Injection (dilution beyond initial reconstitution)

The multipeptide formulation of the invention can be provided in the form of a kit, including instructions for use.

The present invention also provides methods of detecting sensitivity in individuals to ragweed pollen allergens comprising combining a blood sample obtained from the individual with a peptide of the present invention, under conditions appropriate for binding of blood components with the peptide and determining the extent to which such binding occurs. The extent to which binding occurs is determined by assessing T cell function. T cell proliferation or a combination thereof. Other diagnostic methods for allergic diseases which the protein or peptides of the invention can be used include radio-allergergosorbent test (RAST), paper radioimmunosorbent test (PRIST), enzyme linked immunosorbent assay (ELISA), radioimmunoassays (RIA), immuno-radiometric assays (IRMA), luminescence immunoassays (LIA), histamine release assays and IgE immunoblots.

The presence in individuals of IgE specific for ragweed protein allergen and the ability of T cells of the individual to respond to T cell epitope(s) of the protein allergen can be determined by administering to the individuals an Immediate Type Hypersensitivity test and a Delayed Type Hypersensivity test. The individuals are administered an Immediate Type Hypersensitivity test (see e.g. Immunology (1985) Roitt, I.M., Brostoff, J., Male, D.K. (eds), C.V. Mosby Co., Gower Medical Publishing, London, NY, pp. 19.2-19.18; pp. 22.1-22.10) utilizing purified native ragweed protein allergen, a peptide of ragweed protein, allergen or a modified form of the peptide, each of which binds IgE specific for the allergen. The same individuals are administered a Delayed Type Hypersensitivity test

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prior to, simultaneously with, or subsequent to administration of the Immediate Type Hypersensitivity test. Of course, if the Immediate Type Hypersensitivity test is administered prior to the Delayed Type Hypersensitivity test, the Delayed Type Hypersensitivity test would only be given to those individuals exhibiting a specific Immediate Type Hypersensitivity reaction. The Delayed Type Hypersensitivity test utilizes a modified form of ragweed pollen protein or a portion thereof, ragweed protein allergen produced by recombinant DNA techniques, or peptide derived from ragweed pollen protein, each of which has the ability to stimulate human T cells and each of which does not bind IgE specific for the allergen in a substantial percentage of the population of individuals sensitive to the allergen (e.g., at least about 75%). After administration of the Delayed Type Hypersensitivity test, the extent to which a specific Delayed Type Hypersensitivity reaction occurs in the individual to the protein allergen or ragweed pollen protein peptide indicating presence in the individual of T cells specific to T cell epitope(s) of the protein allergen or ragweed pollen protein peptide is determined. Those individuals found to have both a specific Immediate Type Hypersensitivity reaction and a specific Delayed Type Hypersensitivity reaction are diagnosed as having sensitivity to a ragweed allergen and may, if need be, administered a therapeutically effective amount of a therapeutic composition comprising the modified form of ragweed protein allergen or portion thereof, the ragweed protein allergen produced by recombinant DNA techniques, or peptide, each as used in the Delayed Type Hypersensitivity test, and a pharmaceutically acceptable carrier or diluent.

It is also possible to modify the structure of a peptide of the invention for such purposes as increasing solubility, enhancing therapeutic or preventive efficacy, or stability (e.g., shelf life ex vivo, and resistance to proteolytic degradation in vivo.) A modified peptide can be produced in which the amino acid sequence has been altered, such as by amino acid substitution, deletion, or addition, to modify immunogenicity and/or reduce allergenicity, or to which a component has been added for the same purpose.

For example, a peptide can be modified so that it maintains the ability to induce T cell non-responsiveness and bind MHC proteins without the ability to induce a strong proliferative response or possibly, any proliferative response when administered in immunogenic form. In this instance, critical binding residues for the T cell receptor can be determined using known techniques (e.g., substitution of each residue such as, for example, with alanine and determination of the presence or absence of T cell reactivity). Those residues shown to be essential to interact with the T cell receptor can be modified by replacing the essential amino acid with another, preferably similar amino acid residue (a conservative substitution) whose presence is shown to enhance, diminish, but not eliminate, or not affect T cell reactivity. In addition, those amino acid residues which are not essential for T cell receptor interaction can be modified by being replaced by another amino acid whose incorporation may enhance, diminish or not affect T cell reactivity, but not eliminate

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binding to relevant MHC. Preferred amino acid substitutions for non-essential amino acids include, but are not limited to substitutions with alanine, gluatamic acid or a methyl amino acid.

Another example of a modification of peptides is substitution of cysteine residues preferably with serine, threonine, leucine or glutamic acid to minimize dimerization via disulfide linkages. As described in Example XI, a peptide from Region 2, RAE 70.1 (SEQ ID NO:44) was modified to minimize dimerization by substituting serine for the cysteine residue at amino acid position 212. In addition, as shown in Figure 23, peptide AMB 4-9.1 which contains an acid-sensitive aspartic acid-proline bond at amino acid residues 360-361, was modified to increase the stability of this peptide. For example, in peptide AMB 4-9.1DA, the proline at position 361 was substituted with alanine to remove the acid-sensitive bond.

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Stability may also be enhanced in those peptides shown to be susceptible to degradation by deamidation (e.g. the peptide contains a labile Asn-Gly sequence susceptile to deamidation under various conditions. In such situations it has been found that lyophilization stabilized such peptides against deamidation. Peptide Amb 4-9.1 contained a labile Asn-Gly sequence and was found to be susceptible to deamindation under accelerated conditions (e.g. increased buffer concentration, increased ionic strength and increased temperature). Lyophilization stabilized the peptide against deamidation with no significant increase in degradants following 6 months of storage at 5°C.

In order to enhance stability and/or reactivity, peptides can also be modified to incorporate one or more polymorphisms in the amino acid sequence of a protein allergen resulting from natural allelic variation. Additionally, D-amino acids, non-natural amino acids or non-amino acid analogues can be substituted or added to produce a modified peptide within the scope of this invention. Furthermore, peptides can be modified using the polyethylene glycol (PEG) method of A. Sehon and co-workers (Wie et al., supra) to produce a peptide conjugated with PEG. In addition, PEG can be added during chemical synthesis of a peptide of the invention. Modifications of peptides or portions thereof can also include reduction/alkylation (Tarr in: Methods of Protein Microcharacterization, J.E. Silver ed. Humana Press, Clifton, NJ, pp. 155-194 (1986)); acylation (Tarr, supra); esterification (Tarr, supra); chemical coupling to an appropriate carrier (Mishell and Shiigi, eds, Selected Methods in Cellular Immunology, WH Freeman, San Francisco, CA (1980); U.S. Patent 4,939,239); or mild formalin treatment (Marsh International Archives of Allergy and Applied Immunology 41: 199-215 (1971)).

In another embodiment, peptides within an allergen group (e.g., Amb a I or Amb a II) can be modified to enhance T cell reactivity. Given the cross-reactivity within the Amb a I family and Amb a II, a peptide of one group allergen which may be less reactive than a peptide of another group allergen corresponding in amino acid position can have one or more amino acids substituted with one or more amino acids from the corresponding peptide.

Additionally, peptides can be modified to incorporate a polymorphism in the amino acid sequence of a protein allergen resulting from natural allelic variation. Modification of peptides to include one or more of these polymorphisms may result in enhanced stability and/or reactivity.

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To facilitate purification and potentially increase solubility of peptides of the invention, it is possible to add reporter group(s) to the peptide backbone. For example, polyhistidine can be added to a peptide to purify the peptide on immobilized metal ion affinity chromatography (Hochuli, E. et al., Bio/Technology, 6:1321-1325 (1988)). In addition, specific endoprotease cleavage sites can be introduced, if desired, between a reporter group and amino acid sequences of a peptide to facilitate isolation of peptides free of irrelevant sequences. In order to successfully desensitize an individual to a protein antigen, it may be necessary to increase the solubility of a peptide by adding functional groups to the peptide or by not including hydrophobic T cell epitopes or regions containing hydrophobic epitopes in the peptides. For example, in Region 3 of the Amb a I.1 protein, a selected peptide AMB 3-4.1 was modified to increase its solubility by the addition of three naturally occurring sequential residues found in the Amb a 1.1 protein, "RHG", to the 5' end of the peptide. These residues are not necessary for T cell recognition and are also found in peptide AMB 3-5.1. Similarly, as described in Example XI, peptide RAE 70.1 was divided into two fragments and modified to increase solubility. As shown in Figure 25, a fragment of RAE 70.1 ("ORIGINAL" in Figure 25, amino acid residues 194-216, SEQ ID NO:125) was modified by substitution or addition of charged amino acids to increase the hydrophilicity and decrease the pl of the peptide to thereby increase the solubility. For example, in peptide Amb 2-22.1, isoleucine was substituted with glutamic acid to decrease the pl and avoid precipitation of the peptide from solution at a physiological pH. Similar substitutions and grange. additions are shown in Figure 25. In addition, charged amino acids or charged amino acid pairs or triplets when added to the carboxy or amino terminus of the peptide may be particularly useful to increase the solubility of the peptide. Examples of charged amino acids include, but are not limited to arginine (R), lysine (K), histidine (H), glutamic acid (E), and aspartic acid (D). Examples of such modifications are shown in Fig. 14, AMB 2-8.1, AMB 2-9.1, AMB 2-10.1, AMB 2-7.1 and AMB 2-11.1. 30

To potentially aid proper antigen processing of T cell epitopes within a peptide, canonical protease sensitive sites can be recombinantly or synthetically engineered between regions, each comprising at least one T cell epitope. The resulting peptide can be rendered sensitive to cathepsin and/or other trypsin-like enzymes cleavage to generate portions of the peptide containing one or more T cell epitopes. In addition, such charged amino acid residues can result in an increase in solubility of a peptide.

Site-directed mutagenesis of DNA encoding a peptide of the invention can be used to modify the structure of the peptide. Such methods may involve PCR with degenerate oligonucleotides (Ho et al., Gene, 77:51-59 (1989)) or total synthesis of mutated genes

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(Hostomsky, Z., et al., Biochem. Biophys. Res. Comm., 161:1056-1063 (1989)). To enhance bacterial expression, the aforementioned methods can be used in conjunction with other procedures to change the eucaryotic codons in DNA constructs encoding peptides of the invention to ones preferentially used in E. coli, yeast, mammalian cells or other eucaryotic cells.

Another aspect of the invention pertains to an antibody specifically reactive with Amb a I, or a fragment thereof. The antibodies of this invention can be used to standardize allergen extracts or to isolate the naturally-occurring or native form of Amb a I. For example, by using proteins or fragments thereof based on the cDNA sequence of Amb a I, antiprotein/anti-peptide antisera or monoclonal antibodies can be made using standard methods. A mammal such as a mouse, a hamster or rabbit can be immunized with an immunogenic form of such protein or an antigenic fragment which is capable of eliciting an antibody response. Techniques for conferring immunogenicity on a protein or peptide include conjugation to carriers or other techniques well known in the art. Amb a I or fragment thereof can be administered in the presence of adjuvant. The progress of immunization can be monitored by detection of antibody titers in plasma or serum. Standard ELISA or other immunoassays can be used with the immunogen as antigen to assess the levels of antibodies.

Following immunization, anti-Amb a I antisera can be obtained and, if desired, polyclonal anti-Amb a I antibodies isolated from the serum. To produce monoclonal antibodies, antibody-producing cells (lymphocytes) can be harvested from an immunized animal and fused by standard somatic cell fusion procedures with immortalizing cells such as myeloma cells to yield hybridoma cells. Such techniques are well known in the art, for example the hybridoma technique originally developed by Kohler and Milstein, (Nature (1975) 256:495-497) as well as other techniques such as the human B cell hybridoma technique (Kozbar et al.. Immunology Today (1983) 4:72) and the EBV-hybridoma technique to produce human monoclonal antibodies (Cole et al., Monoclonal Antibodies and Cancer Therapy (1985) Alan R. Liss, Inc. pp. 77-96). Hybridoma cells can be screened immunochemically for production of antibodies specifically reactive with Amb a I and the monoclonal antibodies isolated.

The term antibody as used herein is intended to include fragments thereof which are also specifically reactive with $Amb\ a$ I. Antibodies can be fragmented using conventional techniques and the fragments screened for utility in the same manner as described above for whole antibodies. For example, $F(ab')_2$ fragments can be generated by treating antibody with pepsin. The resulting $F(ab')_2$ fragment can be treated to reduce disulfide bridges to produce Fab' fragments. The antibody of the present invention is further intended to include bispecific and chimeric molecules having an anti- $Amb\ a$ I portion.

Another aspect of this invention provides T cell clones and soluble T cell receptors specifically reactive with Amb a I or a fragment thereof. Monoclonal T cell populations (i.e., T cells genetically identical to one another and expressing identical T cell receptors) can be

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derived from an individual sensitive to Amb a I, followed by repetitive in vitro stimulation with Amb a I or portion thereof in the presence of MHC-matched antigen-presenting cells. Single Amb aI MHC responsive cells can then be cloned by limiting dilution and permanent lines expanded and maintained by periodic in vitro restimulation. Alternatively, Amb a I specific T-T hybridomas can be produced by a technique similar to B cell hybridoma production. For example, a mammal, such as a mouse can be immunized with Amb a I or fragment thereof, T cells from the mammal can be purified and fused with an autonomously growing T cell tumor line. From the resulting hybridomas, cells responding to Amb a I or fragment thereof are selected and cloned. Procedures for propagating monoclonal T cell populations are described in Cellular and Molecular Immunology (Abul K. Abbas et al. ed.), W.B. Saunders Company, Philadelphia, PA (1991) page 139. Soluble T cell receptors specifically reactive with Amb a I or fragment thereof can be obtained by immunoprecipitation using an antibody against the T cell receptor as described in Immunology: A Synthesis (Second Edition), Edward S. Golub et al., ed., Sinauer Associates, Inc., Sunderland, MA (1991) pages 366-269.

T cell clones specifically reactive with Amb a I or fragment thereof can be used to isolate and molecularly clone the gene encoding the relevant T cell receptor. In addition, a soluble T cell receptor specifically reactive with Amb a I or fragment thereof can be used to interfere with or inhibit antigen-dependent activation of the relevant T cell subpopulation, for example, by administration to an individual sensitive to a cat allergen. Antibodies specifically reactive with such a T cell receptor can be produced according to the techniques described herein. Such antibodies can be used to block or interfere with the T cell interaction with peptides presented by MHC.

The present invention also provides nucleic acids having sequences encoding proteins and peptides of the invention. Nucleic acid sequences used in any embodiment of this invention can be cDNA as described herein, or alternatively, can be any oligodeoxynucleotide sequence having all or a portion of a sequence represented herein, or their functional equivalents. Such oligodeoxynucleotide sequences can be produced chemically or mechanically, using known techniques. A functional equivalent of an oligonucleotide sequence is one which is 1) a sequence capable of hybridizing to a complementary oligonucleotide to which the sequence (or corresponding sequence portions) of SEQ ID NO: 1, 3, 5, 7, 9 and 11 or fragments thereof hybridizes, or 2) the sequence (or corresponding sequence portion) complementary SEQ ID NO: 1, 3, 5, 7, 9 and 11 and/or 3) a sequence which encodes a product (e.g., a polypeptide or peptide) having the same functional characteristics of the product encoded by the sequence (or corresponding sequence portion) of SEQ ID NO: 1, 3, 5, 7, 9 and 11. Whether a functional equivalent must meet one or more criteria will depend on its use (e.g., if it is to be used only as an oligoprobe, it need meet only the first or second criteria and if it is to be used to produce a peptide of the present invention, it need only meet the third criterion).

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As described in the Examples which follow, Amb a I and Amb a II proteins have been recombinantly expressed in E. coli, purified and shown to have reduced binding to human allergic ragweed pollen IgE on Western blots. Overlapping peptides derived from the Amb a I.1 protein and various peptides derived from Amb a I.3 and Amb a I.2 were synthesized and used to identify regions of T cell reactivity within the protein. These regions of T cell reactivity were further defined by modifying selected Amb a I.1 peptides and determining T -cell reactivity to these peptides.

This invention is further illustrated by the following non-limiting examples.

10 Example I Native Ragweed Pollen Allergen Purification

What follows is a description of the work done to biochemically purify the allergens of Ambrosia artemisiifolia in their native form as primary antigens for human T-cell epitope mapping.

50 g of defatted short ragweed pollen (Greer Labs) was extracted in 500 ml .05 M Tris pH 7.95 containing protease inhibitors. The extract was then depigmented by batch absorption with Whatman DE-52 DEAE cellulose (150 g dry weight) in the presence of 0.2 M NaCl at 4°C. Unbound material was dialysed against .025 M Tris pH 7.95 with protease inhibitors. The depigmented sample was next applied to an 80 ml DEAE cellulose column (Whatman DE-52) equilibrated in .025 M Tris pH 7.95 containing protease inhibitors.

Acidic proteins were eluted with .025 M Tris, 0.2 M NaCl pH 7.95 at 4°C with inhibitors.

In order to biochemically purify Amb a I, the acidic DEAE elution sample was fractionated by ammonium sulfate precipitation into 0-45% and 45-59% saturation samples (4°C). The Amb a I-enriched 45-59% pellet was applied at 0.5 ml/min (4°C) to a 500 ml Sephacryl S200 (Pharmacia) column in .05 M ammonium bicarbonate containing inhibitors. Purified Amb a I was recovered in the 38 kD region and dialysed against .04 M Tris pH 8.0. This sample was next applied to an 8 ml Mono Q HR 10/10 (Pharmacia) column in .04 M Tris pH 8.0 at 25°C. Elution was performed with .04 M Tris pH 8.0 containing .08M NaCl and the major peaks were analyzed as discussed below for confirmation and purity of Amb a

30 To biochemically purify Amb a II, the acidic DEAE elution sample was separated into an Amb a II-enriched fraction by ammonium sulfate precipitation at 0-45% saturation (4°C). The pellet was applied at 0.5 ml/min (4°C) to a 200 ml/Sephadex G75 (Pharmacia) column in .05 M ammonium bicarbonate containing inhibitors. Purified Amb a II was recovered in the 38 kD region and dialysed against .04 M Tris pH 8.0 at 25°C to separate contaminating Amb a I from Amb a II. Elution was performed with .04 M Tris pH 8.0 containing .08M NaCl.

The major peaks were analyzed by IEF SDS-PAGE using a mixture of 4.5-5.3 and 3.5-10.0 ampholytes (Pharmacia) in a 7% acrylamide gel. Protein sequencing was also performed to confirm Amb a II.

In another embodiment Amb a 1.1 and Amb a 1.2 were biochemically purified as follows. Defatted short ragweed pollen was purchased from Greer laboratories (Lenior, North Carolina). The pollen (50g) was extracted overnight at 4°C with extraction buffer (10ml/g pollen) containing 50 mM Tris-HCl, pH8 and protease inhibitors in final concentrations: phenyl methyl sulfonyl fluoride (170 μ g/ml); soybean trypsin inhibitor (1 μ 5 g/ml); leupeptin (1 μ g/ml) and peptstatin A (1 μ g/ml). The soluble extract was clarified by sequential filtration through Whatman #1 paper (Whatman, Maidstone England) followed by an 0.8 micron filter. The soluble pollen extract (SPE) was either used for IgE binding studies or subjected to further purification as described below. For purification of Amb a 1.1 and Amb a I.2, sodium chloride was added to SPE to a final concentration of 0.3M and the material was depigmented by batch abasorption onto What DE 52 cellulose (50 g dry weight) with exraction buffer plus 0.3 M NaCl. The solution was separated from the resin by filtration (Whatman #1 paper, Whatman, Maidstone England). The sample was then fractionated by sequential (NH₄)₂SO₄ precipitation, first at 0-47.5% and then at 47.5%-60% saturation at 4°C. The latter precipitate enriched in Amb a I, was separated by 15~ centrifugation (10,000 x g, 1h) and the pellet was resuspended in 20 mM Tris-HCl, pH 8. The sample was then loaded onto an anti-Amb a I murine monoclonal antibody affinity column (mAb-4B5B7, Dr. D. Klapper, Univ. North Carolina, Chapel Hill, NC) which was determined to bind members of the Amb a I/Amb a II family. Amb a I was eluted with 0.1 M glycine, pH11 and was immediately neutralized with 1 M sodium phosphage, pH 3.9 This 20 preparation contained both Amb a I.1 and Amb a I.2 as detected by SDS-PAGE analysis and by Western blotting. The affinity purified proteins were pooled, concentrated and dialyzed against 20 mM Tris, pH 8.5, extensively. Amb a 1.1 and Amb a 1.2 were further separated by a Source Q ionic exchange column (Pharmacia, Piscataway, NJ), which was equilibrated with 20 mM Tris, pH 8.5. The flow-through material contained Amb a 1.2 whereas Amb a 1.1 was eluted with step gradients of 24 mM and 80 mM NaCl. The latter peak also contained some α,β cleaved fragments of Amb a 1.1. Amb a 1.1 and Amb a 1.2 purity was verified by NH2terminal amino acid sequencing as being greater than 95%.

Example II Recombinant Ragweed Pollen Allergen Expression

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What follows is a description of the work done to produce the allergens of Ambrosia artemisiifolia as recombinant proteins in E. coli.

Described and provided in USSN 07/529,951, filed May 29, 1990, (incorporated herein by reference) are full length cDNAs encoding Amb a I.1, Amb a I.2, Amb a I.3, Amb a I.4 and Amb a II. cDNA inserts encoding these five protein allergens were constructed in the vector pTrc99A (Amann, E., et al., Gene, 69:301 (1988)) which was kindly provided by Dr. Egon Amann (Behringwerke AG, Marburg, FRG). The nucleotide sequences and deduced amino acid sequences of the Amb a I family members are shown in the sequence listing as SEQ ID NO:1 and 2 (Amb a I.1), SEQ ID NO:3 and 4 (Amb a I.2), SEQ ID NO:5 and 6

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(Amb a I.3), SEQ ID NO:7 and 8 (Amb a I.4), and SEQ ID NO:9 and 10 (Amb a II). The cDNAs encoding each allergen were cloned in frame with a polylinker encoding six sequential histidines, (CAC)₆, that had been inserted into the 5' end of the pTrc99A vector as a Ncol/EcoRI synthetic adapter (Maniatis T., et al.

Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1982). These cDNA inserts were then cloned in-frame into the appropriate pTrc99. To further enhance expression, a retroregular stem-loop sequence was placed at the 3' end in the untranslated region (Skoglund, C. M., et al. Gene 88:1 (1990)). The H₆ leader sequence allowed purification using QIAGEN NTA-Agarose (Diagen GmbH, Dusseldorf, FRG), a Ni²⁺ chelating support (Hochuli, E. et al., Biotechnology, 6:1321 (1988)). The vectors were transformed into the XL-1 Blue host bacteria. Expression of individual recombinant proteins was induced when cultures reached OD₆₀₀=0.6 to 0.7 by the addition of isopropyl-D-thiogalactopyranoside (IPTG) to 1 mM concentration to the culture medium. After 2 hours of further growth at 37°, the cells were pelleted. Recombinant proteins were then obtained by either of the following procedures. In one embodiment, the cells were resuspended in lysozyme containing phosphate buffer (0.4 mg/ml) and incubated for 30 minutes on ice. The cell suspension was frozen and quick thawed followed by sonication

recovered by a low speed centrifugation and solubulized in 10 ml (per 1 liter growth) of buffer containing 8 M urea, 50 mM Tris, pH 8.0, 2 mg/ml leupeptin, 2 mg/ml pepstatin and 1 mg/ml soybean trypsin inhibitor. The urea solubilized preparation was subjected to a low speed centrifugation and the recombinant proteins in the supernatant isolated by metal ion chromatography (Hochui supra).

(Bond, J. G. et al., J. Immunol., 146:3380 (1991)). Insoluble recombinant protein was

In another embodiment, the pelleted bacteria were resuspended in 6 M guanidine HC1, 100 mM 2-mercaptoethanol, 100 mM NaPO4, 10 mM Tris pH 8.0. This suspension was subjected to centrifugation at 15,000 X g, and the supernatant removed, adjusted to pH 8.0 with 10 N NaOH, and applied to an NTA agarose column that had been equilibrated in 6 M guanidine HC1, 100 mM NaPO4, 10 mM Tris pH 8.0. The column was washed in 6 M guanidine HC1, 100 mM NaPO4, 10 mM Tris pH 8.0 until the OD280 of the effluent reached background. The column buffer was then switched to 8 M urea, 100 mM NaPO4, 10 mM Tris pH 8.0. After equilibration, a more stringent wash was performed in 8 M urea, 100 mM NaOAc, 10 mM Tris pH 6.3 until the OD280 of the effluent reached background. Recombinant protein (as an H6 fusion) was then eluted in 8 M urea, 100 mM NaOAc, 10 mM Tris pH 4.5 and collected in aliquots whose OD280 profile was monitored. The protein peak was dialyzed 3 times into 500 volumes of PBS for human T-cells analysis. Yield ranged from 1-3 mg of recombinant protein per litre with purity of approximately 55% (as determined by densitometric scanning).

In another embodiment, high yields (e.g. 50-100 mg purified r $Amb\ a$ I.1 protein/L of fermentation broth) were recovered. This embodiment involves the use of a construct which

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is different from the one described above in 3 ways: 1)deletion of sequence coding for 36 non-Amb a I related amino acids, 2) attachment of a 6 histidine linker to the NH2-terminus, and 3) replacement of 2 existing arginine codons with E. coli-preferred arginine codon. The resulting construct showed significantly higher expression levels than did the original construct. This phenomenon was true in both shaker flask and 10L fermentation cultures. The purification also differed significantly from the procedure described above. Cell paste from fermentors was homogenized and inclusion bodies were purified prior to solubilization in guanidine-HCL. The solubilized inclusion bodies were then purified using metal chelate chromatography and buffer exchanged into acetate buffer. The resulting protein is greater than 90% full length r Amb a I.1.

Example III IgE Analysis With Purified Recombinant Ragweed Pollen Allergens

Both Western blotting and ELISA techniques were used to analyze the binding of IgE to purified recombinant Amb a I and Amb a II proteins. Additionally, ELISA was used for analysis of IgE reactivity to peptides derived from the Amb a 1.1 protein.

A. Western Blot Analysis

The antigens were loaded on the gels as follows: lane 1 SPE (Soluble Pollen Extract), 15 mg/lane; lane 2 rAmb a I.1 (recombinant Amb a I.1), 3 mg/lane; lane 3 rAmb a I.2, 4 mg/lane; lane 4 rAmb a I.3, 3 mg/lane; lane 5 rAmb a I.4, 3 mg/lane; and lane 6 rAmb a II.1, 4 mg/lane.

The gel electrophoresis and Western blot transfer procedures used were essentially as described elsewhere (Towbin, H., et al., Proc. Natl. Acad. Sci., USA, 76:4350 (1979)). Briefly, SDS-PAGE was performed in 10% acrylamide gels under reducing conditions (10mM dithiothreitol, at constant current). The transfer to nitrocellulose (0.1 mM, Schleicher & Schuell, Keene, NH) was performed in a Hoeffer apparatus according to the protocol of Towbin et al., supra. After transfer, the blots were rinsed in blot solution (25 mM Tris-HCl, pH 7.5, 0.17 M NaCl, and 0.05% Tween 20) and stained for 1 hour with 0.1% India ink. All subsequent incubations with antibodies and washes were performed in blot solution at room temperature. The first antibody incubations were performed overnight then rinsed and incubated with the appropriate biotinylated second antibody (Kirkegaard Perry Laboratories. Gaithersburg, MD). The final incubation was performed using 125-I streptavidin (1 mCi/25 ml blot solution) for 1 hour followed by removal of unbound labeled material and autoradiography at -80°C with an intensifying screen.

Fig. 1 shows the Western blot IgE binding pattern of two ragweed allergic patient plasma samples (#475 and #143). These patterns are representative of the typical IgE reactivity to these proteins on a Western blot. Both patients show binding to the Amb a I.1 and Amb a 1.3 gene products in the soluble pollen extract (lane 1, Fig. 1). The results

demonstrate clear IgE binding reactivity in these patients to Amb a I.1, Amb a I.3, Amb a I.4. In contrast, Amb a I.2 and Amb a II.1 showed markedly reduced IgE binding.

B. ELISA

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Patient #143 IgE was also tested for binding to purified recombinant Amb a I and Amb a II proteins by ELISA. Fig. 2 shows the results of this analysis performed according to the following method.

Corning assay plates (#25882-96) were coated with each antigen listed in Fig. 2 at the following concentrations: Soluble Pollen Extract (SPE) 15 mg/ml; rAmb a 1.1, 5 mg/ml; rAmb a I.2, 20 mg/ml; rAmb a I.3, 5 mg/ml; rAmb a I.4, 15 mg/ml; and rAmb a II, 20 mg/ml. 10 50 mls/well of the above antigens were added and coating was carried out overnight at 4° C. The coating antigens were removed and the wells were blocked with 0.5% gelatin in PBS, 200ml/well for 2 hours at room temperature. Patient #143 plasma was serially diluted with PBS-Tween 20 (PBS with 0.05% nonionic detergent Tween-20 Sigma, St. Louis MO) and 100ml/well was added and incubated overnight at 4°C (plasma dilutions are tested in 15 duplicate). The second antibody (biotinylated goat anti-Human IgE, 1:1000, Kirkegaard & Perry Laboratories Inc. Gaithersburg, MD), was added at 100 ml/well for one hour at room temperature. This solution was removed and streptavidin-HRPO, 1:10000, (Southern Biotechnology Associates, Inc., Birmingham, AL) was then added at 100ml/well for one hour at room temperature (all wells are washed three times with PBS-Tween between each incubation step). TMB Membrane Peroxidates Substrate system (Kirkegaard & Perry Laboratories) was freshly mixed, and added at 100ml/well. The color was allowed to develop for 2-5 minutes. The reaction was stopped by the addition of 100ml/well of 1 M phosphoric acid. Plates were read on a Microplate EL 310 Autoreader (Biotek Instruments, Winooski, VT) with a 450nm filter. The absorbance levels of duplicate wells was averaged. 25 The graphed results (log of the dilution vs absorbance) of the ELISA assays are shown in Fig. 2. The order of coating antigens listed vertically in these figures corresponds in order from left to right to the coating antigens listed for each histogram.

The results of the ELISA assay, Fig. 2, demonstrate a similar reactivity pattern as seen in the Western blot analysis (Fig. 1). The observation of much stronger binding to SPE is due to the fact that this antigen preparation is non-denatured. However, the recombinant Amb a proteins, because of their non-native bacterial origin behave as denatured antigens. The hierarchy of IgE binding in this patient to rAmb a proteins by ELISA shows that Amb a 1.1 binds IgE more strongly than Amb a 1.3 and Amb a 1.4. The results also show that Amb a 1.2 and Amb a II demonstrate markedly reduced binding to IgE.

As shown in Fig. 3, another ELISA assay was performed using a pool of human allergic sera. This assay demonstrates that biochemically purified (native) Amb a I and Amb a II proteins bind significantly more IgE than the respective recombinant proteins (rAmb a I.1 and rAmb a II). (The control in this experiment was plates coated without antigen and PBS).

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C. Modified ELISA for Greater Sensitivity

In another embodiment, an ELISA based assay system for serum IgE binding to peptides was developed which had greater sensitivity than direct ELISA, and creating tandem Amb a I peptide copies in the form of a recombinant trimer. One strategy for constructing a recombinant trimer based on Amb a I peptide Amb 4-9.1 is as follows.

DNA sequences encoding Amb 4-9.1 are amplified by PCR from a plasmid containing Amb a I.1 sequence. Three Amb 4-9.1 copies are made each containing unique restriction enzyme cleavage sites. The restriction sites are designed (inserted during the amplification reaction) to ensure peptide ligation to each other or ligation to plasmid. Amb 4-9.1 copies are ligated together and then the fused sequences are ligated into a sequencing vector, pUC19. After DNA sequence confirmation, the trimer sequences are excised from pUC19 and inserted into an expression vector pEt-11d containing a 6 histidine tag.

After DNA sequence confirmation, the expression plasmid containing Amb 4-9.1 trimer sequence was expressed in *E.coli* BL21. Expressed protein was purified over a Ni²⁺ column, protein was analyzed by SDS-Page, Western, ELISA and protein sequencing. The Amb 4-9.1 trimer was greater than 95% pure and was soluble in PBS. The sequence of the Amb 4-9.1 trimer was confirmed as:

MGHHHHHHEFELGTKDVLENGAIFVASGVDPVLTPEQSAGSRGTK DVLENGAIFVASGVDPVLTPEQSAGLQGTKDVLENGAIFVASGVDPVLTPEQSAG.

Side by side direct ELISAs were carried out in order to compare the sensitivity of assays using unmodified Amb 4-9.1 peptide versus recombinant Amb 4-9.1 trimer constructed as discussed above. These assays were carried out using a panel of plasma samples from individuals with high skin test reactivity to Amb a I. Plasma previously shown to have IgE binding to unmodified Amb 4-9.1 peptide served as a positive control. When IgE binding the the Amb 4-9.1 recombinant trimer was detected, competition ELISA was used to verify that the binding was relevant to the unmodified Amb 4-9.1 peptide (rather than to a novel epitope created by the recombinant Amb 4-9.1 structure).

There was a significant enhancement of assay sensitivity (both the number of positives and assay signal/noise ratio) using the Amb 4-9.1 recombinant trimer. Thus, a very sensitive assay has been developed which is useful for screening IgE binding to any Amb a pollen protein allergen peptide. This assay is particularly useful for screening Amb a 1.1 peptides which are to be formulated into a human therapeutic. This modified assay is also useful for screeninf for IgE binding to any peptide derived from any antigen by first constructing a recombinant trimer of the peptide of interest as discussed above and using the recombinant trimer in the assay for detecting IgE binding.

Example IV Tolerization of Mice with Recombinant Amb a I Proteins

Balb/c mice (H-2 d) were immunized in the hind foot pads and at the base of the tail with an emulsion of Complete Freunds Adjuvant (CFA) and 50mg/mouse Amb a I.1. Seven days later, the popliteal lymph nodes, superficial inguinal lymph nodes, and the periaortic lymph nodes were harvested and cultured (1 x 10⁵ lymph node cells + 2 x 10⁵ irradiated spleen feeders) with various challenge antigens in vitro. The in vitro antigens consisted of various doses of Amb a I.1, Amb a I.2, Amb a I.3, Amb a I.4, Amb a II, and a media control. Cultures were incubated for 3 days at 37°C in a CO2 incubator. On day 3, each culture was pulsed with 1mCi ³H-thymidine and on day 4 the cultures were harvested and proliferation was monitored by incorporation of ³H into DNA.

As seen in Fig. 4A, the mice that were immunized with $50 \text{mg } Amb \ a \ 1.1 + CFA$ have a good response to Amb a I.1, Amb a I.3, and Amb a I.4. The response to Amb a I.2 and Amb a II is also good but less than other antigens. It appears as though Amb a I.1 is very immunogenic in Balb/c mice and results in a response to each Amb a family member. When mice were immunized with PBS + CFA, there was no significant response to any of the Amb a family members (Fig. 4B).

An experiment was conducted to determine whether mice could be tolerized with Amb a I.1. The mice were tolerized with Amb a I.1 and pollen extract and then challenged with Amb a I.1 or pollen extract. The outline of this experiment #5 is shown below:

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Day 1: Group (1): 6 Balb/c mice tolerized with 300mg Amb a I.1 + Incomplete Freunds Adjuvant (IFA) intraperitoneally (i.p.) Group (2): 6 Balb/c mice tolerized with 300mg Pollen Extract + IFA i.p. Group (3): 6 Balb/c mice exposed to PBS + IFA i.p.

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Day 15 Challenge 3 mice from each group with 50mg Amb a 1.1 + CFA subcutaneously (s.c.) Challenge the remaining 3 mice from each group with 50mg Pollen extract + CFA s.c.

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Day 23 Harvest Lymph Nodes (Analyze each mouse separately) Test Lymph Node Response on Amb a I.1, I.2, I.3, I.4, II.1, Pollen Extract, Concanavalin A (Con A), and media.

The animals were sacrificed by cervical dislocation on day 23 and the popliteal lymph 35

nodes, superficial inguinal lymph nodes, and the periaortic lymph nodes were removed and placed in rinsing buffer (cold RPMI 1640 containing 1% FCS). The nodes were rinsed with rinsing buffer and forced through a fine stainless steel mesh, using a glass pestal to suspend them in rinsing buffer. The suspended cells were rinsed two times by centrifugation at 1000

rpm for 10 minutes and resuspended in rinsing buffer. An aliquot from each sample was taken in order to do a cell count. The cells (4 x 10⁶/ml) were incubated in culture media (RPMI 1640 media containing 10% FCS, 2mM L-glutamine, 50U/ml penicillin, 50 mg/ml streptomycin and 5 x 10⁻⁵ M 2-mercaptoethanol) and test antigens at various concentrations. The triplicate 0.1 ml cultures (U-bottom 96 well plates (Costar)) were incubated at 37°C and 5 at 5% CO₂. After 24 hours, 50ml of media from each well was placed in separate flat bottom 96 well plates (Costar) and frozen overnight at -20°C to eliminate carryover of live cells. The supernatants were tested after thawing for their ability to support the growth of CTLL3, an IL-2 dependent T cell clone. CTLL3 in log phase growth were rinsed 3 times by centrifugation at 1000 rpm for 10 minutes. CTLL3's were added to the warmed culture 10 supernatants (5 x 10^3 cells/well) and the IL-2 assay was incubated at 37°C and 5% CO₂. jarna In riva After 24 hours, 1 mCi/well ³H thymidine was added in 50 ml/well and the CTLL3 cells were incubated for an additional 4-6 hours. Following the pulse with ³H, the plates were harvested on a Tom Tek 96 well cell harvester. The ³H incorporation in each well was counted by a Betaplate Model 1205 scintillation counter. Background counts were not 15 subtracted.

Fig. 5 shows the lymph node responses to Amb a I.1, Amb a I.2, Amb a I.3, Amb a I.4. and Amb a II in the mice that were tolerized with Amb a I.1 or Phosphate Buffered Saline (PBS) and challenged with Amb a I.1. Fig. 5A, shows that the mice that were tolerized with Amb a I.1 have a lower response to Amb a I.1 than the mice that were tolerized with PBS. The same appears to be true in the response to Amb a I.2, Amb a I.3, and Amb a I.4 (Figs. 5B. 5C, 5D). In Figs. 5E and 5F, it appears as though there is no significant difference in the response to Amb a II or pollen extract between the tolerized group and the non-tolerized group. Figs. 6A-6F show the lymph node responses to the various proteins in the mice that were exposed to pollen extract or PBS and challenged with pollen extract. Table 1 summarizes the results of these experiments.

TABLE 1

30		Amba I.1	Amba I.2	Amba I.3	Amba I.4	Amba II.1	Pollen
	Tolerize/Challenge Control/ <u>Amba</u> I.1	+	+	+	+	-	- -
• • • •	Amba I.1/Amba I.1	-	-	/-	-	-	
	Control/Pollen	+	+	+/-	+/-	-	+
	Pollen/Pollen	-	-	-	-	-	-

(-) indicates a diminished response as compared to control

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Example V Synthesis of Overlapping Peptides

Amb a 1.1 overlapping peptides as shown in Fig. 7 were synthesized using standard Fmoc/tBoc synthetic chemistry and purified by dialysis or Reverse Phase HPLC. In addition, various peptides derived from Amb a 1.2 and Amb a 1.3 were synthesized. The amino acid residues of the synthesized peptides are in brackets by the peptide name and the amino acid sequence (in single letter code) is next to the peptide name. The peptide names are consistent throughout the Figures. In the design of the overlapping peptides, the relationship of the Amb a I family members at the level of T cell cross-reactivity as determined in Example VIII was considered. As shown in Table IV, the Amb a I protein allergens share a high degree of cross-reactivity. In addition, Amb a 1.1 and Amb a II were found to have 55.2% cross-reactivity. Given the high degree of cross-reactivity it was expected that "shared" T cell epitopes exist among Groups I and II. Thus, the amino acid sequences of the Amb a I family members and Amb a II were examined to identify conserved and variable regions. It was expected that conserved regions within the Amb a I family members and Amb a II would contain "shared" T cell epitopes.

Example VI T Cell Responses to Ragweed Peptides

Peripheral blood mononuclear cells (PBMC) were purified by Sepracell-MN or lymphocyte separation medium (LSM) centrifugation of 60 ml of heparinized blood from ragweed-allergic patients who exhibited clinical symptoms of seasonal rhinitis and were skin prick test positive for ragweed. T cell lines were established from these cells by stimulation of 1-2 x 10⁶ PBMC/ml in RPMI-1640 containing 5% human AB serum (complete medium) with recombinant Amb a I.1 at 20-30 mg/ml for 5-6 days at 37°C in a humidified CO₂ incubator. Viable cells were purified by LSM centrifugation and cultured in complete medium supplemented with 5 units recombinant human IL-2/ml and 5 units recombinant human IL-4/ml for up to three weeks until the cells no longer responded to lymphokines and were considered "rested". The ability of the T cells to proliferate to Amb a I.1, Amb a I.2 and Amb a I.3 sequence-derived synthetic peptides was then assessed.

For assay, 2 x 10⁴ rested cells were restimulated in the presence of 2 x 10⁴ autologous Epstein-Barr virus (EBV)-transformed B cells (gamma-irradiated with 25,000 RADS) or 5 x 10⁴ autologous PBMC (3,500 RADS) with various concentrations of *Amb a* I.1 synthetic peptides in a volume of 200 ml complete medium in duplicate or triplicate wells in 96-well round bottom plates for 3 days. Each well then received 1 mCi tritiated thymidine for 16-20 hours. The counts incorporated were collected onto glass fiber filter mats and processed for liquid scintillation counting. Table II shows the results of a representative assay. The maximum response in a titration of each peptide is expressed as the S.I. or stimulation index. The S.I. is the CPM incorporated by cells in response to peptide divided by the CPM incorporated by cells in medium only. An S.I. value greater than the background level is considered "positive" and indicates that the peptide contains a T cell epitope.

Same.

However, only individual S.I. values above 2.0 (a response two-fold or greater over background) were used in calculating mean stimulation indices for each peptide for the group of patients tested. The results shown in Table II demonstrate that this patient (#466) responds very well to peptides RAE 7.D, RAE 69.1, RAE 64.1, and RAE 29.1. This indicates that these peptides contain Amb a I.1 T cell epitopes recognized by T cells from this particular allergic patient.

TABLE II

10	Antigen	Concentration (mg/ml)	CPM	S.L.
	Medium		1677	••
·¥*	Amb a I.1	2	12113	• •
		10	25047	
15		50	45710	27.3
	RAE 22.E	5	3198	
	KAL ZZ.L	50	3581	2.1
	RAE 7.D	5	3948	
20	RAE 1.D	50	8572	5.1
	RAE 45.15	5	1160	
15 mg 1		50	1370	0.8
25 ⁶		5	6800	
	RAE 69.1	50	11464	6.8
		5 .	4116	2.5
30	RAE 70.1	50	3888	
			1236	
	RAE 65.1	5 50	1784	1.1
			12828	7.7
35	RAE 64.1	5 50	2259	
			6998	
	RAE 29.1	5	13078	7.8
		50	13076	7.0

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The above procedure was followed with a number of other patients. Individual patient results were used in calculating the mean S.I. for each peptide if the patient responded to the Amb a I.1 protein at an S.I. of 2.0 or greater and the patient responded to at least one peptide derived from Amb a I.1 at an S.I. of 2.0 or greater. A summary of positive experiments from 39 patients is shown in Fig. 8. The bar represents the cumulative rank of the peptide response in the 39 patients. To determine the cumulative rank, the 5 peptides with the highest S.I. in each patient were determined and assigned a numerical rank in descending order with 5 representing the strongest response. The ranks for each peptide were then summed in the 39 patients to determine the cumulative rank for the peptide. The number above each bar is the mean S.I. of the positive responses (S.I. of 2.0 or greater) from the group of patients to that peptide. In parentheses above each bar is the positivity index (P.I). The P.I. for each peptide is determined by multiplying the mean S.I. by the percent of patients who responded to that peptide. The P.I. therefore represents both the strength of the response (S.I.) and the frequency of a response to a peptide in the group of patients tested. For example, peptide RAE 69.1 had the highest cumulative rank so it was the best peptide response in the overall population of 39 even through it did not have the highest mean S.I. Similarly, RAE 70.1 had the highest mean S.I but not the best cumulative rank or P.I. Thus, the response to RAE 70.1 was strong when it occurred but it did not occur as frequently in the population as the response to other peptides. The peptide with the highest P.I., RAE 29.1, also had a strong S.I. and the second highest cumulative rank. The response to this peptide was therefore generally strong and relatively frequent in this population.

Example VII T Cell Epitope Fine Map Studies with Amb a I.1

Based on the above analysis 4 major areas of T cell reactivity within Amb a I.1 were identified. (Regions 1, 2, 3 and 4). All 39 patients responded to Amb a I.1 and a peptide from at least one of these regions: Region 1, amino acids residues 48-107, Region 2, amino acid residues 171-216; Region 3, amino acid residues 278-322; and Region 4, amino acid residues 331-377. Based in part on the T cell reactivity shown in Fig. 8, Amb a I.1 peptides were selected and modified by addition or deletion of amino acid residues at either the 5' or 3' end of the peptide. This set of peptides is shown in Fig. 9. T cell studies similar to those of Example VI were performed using these selected peptides to more precisely define the major areas of T cell reactivity within Regions 1-4 of the Amb a I.1 protein. For example, PBMC from a single ragweed-allergic patient were isolated as described in Example VI and were stimulated with 20 mg/ml of recombinant Amb a I.1 as described above. The results of proliferation assays with this one patient to selected peptides using irradiated (24,000 RADS) autologous EBV B cells as antigen presenting cells is shown in Table III. The data indicates that T cells from this patient respond well to RAE 7.D, RAE 70.1, RAE 40.1-4, and RAE 28.1-2.

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TABLE III

	Antigen	Concentration (mg/ml)	<u>CPM</u>	S.I.
5	Medium		2762	
. •	РНА	1	97864	35.4
 :·	Amb a I.1	2	28025	•
10		10	56172	*,
TEF		50	86598	31.4
•	RAE 22.E	5	3318	1.2
٠.		50	784	
15			•	
	RAE 7.D	5	53292	
		50	60943	22.1
	RAE 40.1-4	. 5	4504	
20		50	13549	4.9
	RAE 69.1	5	5213	
	ICAL O7.1	50	5981	2.2
	£			
25		5	14223	5.1
*		50	4729	
		_		•
	RAE 64.1	5	3418	1.2
	10 10 0 111	50	2538	•
30		-		
JU	RAE 28.1-2	5	7542	
	10 LL 20.1-2	50	24208	8.8
			<i>!</i>	

The above procedure was followed with a number of other patients and yielded 47 positive experiments. A summary of the results for Region 1 peptides of the Amb a I.1 protein is shown in Fig. 10, Region 2 peptides in Fig. 11, Region 3 peptides in Fig. 12, and Region 4 peptides in Fig. 13. As described in Example VI, the bar shows the cumulative rank. In these experiments the 3 best peptide responses were ranked. The number above each bar represents the mean S.I. for the peptide and the number in parentheses the P.I. for the peptide. In these experiments an individual S.I. of 2.5 or greater was used in calculating the mean S.I.

Fig. 10 indicates that the major area of T cell reactivity within Region 1 of the Amb a I.1 protein is represented by peptides RAE 6.D, RAE 7.D, RAE 40.1, RAE 40.1-4, and RAE 40.1-5. Thus, a preferred area of major T cell reactivity within Region 1 comprises amino acid residues 57-101. Fig. 11 indicates that there is a broad area of weak T cell reactivity in Region 2 of the Amb a I.1 protein relative to Region 1. A preferred area of major T cell reactivity within Region 2 thus comprises amino acid residues 182-216. Fig. 12 shows that the most frequent and dominant response within Region 3 of the Amb a I.3 protein is to peptide RAE 64.1. However, another area of T cell reactivity is represented by peptide RAE 66.1. A preferred area of major T cell reactivity within Region 3 comprises amino acid residues 280-322. Fig. 13 indicates that the major area of T cell reactivity in Region 4 is represented by peptides RAE 28.1-2, RAE 28.1-1, and RAE 28.1. From this analysis, a preferred area of major T cell reactivity within Region 4 comprises amino acid residues 342-377.

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To further validate and define peptides derived from the Amb a I.1 protein comprising T cell epitopes, selected peptides from Regions 1-4 were further modified by addition or deletion of amino acid residues as described above. These selected modified peptides are shown in Fig. 14. To determine the T cell reactivity of these peptides. PBMC were stimulated with recombinant Amb a I.1 and assayed with the selected peptides as described in Example VI, except in some cases autologous PBMC (irradiated 3.500 RADS) were used as antigen presenting cells. The assay was followed with a number of patients and resulted in 23 positive experiments. In these assays an individual S.I. of 2.0 was used in calculating the mean stimulation index. In Figs. 15-18, the bar represents the cumulative rank of each peptide. The best 3 peptide responses for each patient were ranked as described above. The mean S.I. for each peptide and percent positive are found above the bar.

Example VIII Amb a I.1 Specific T Cell Cross-Reactivity To Other Amb a I Family Members and Amb a II

To determine the relationship of the $Amb\ a$ I family members (i.e., $Amb\ a$ I.1, I.2, I.3, and I.4) at the level of T cell reactivity, PBMC were stimulated with recombinant $Amb\ a$ I.1 (r $Amb\ a$ I.1) as described in Example VI. For assay, 2×10^4 rested cells were restimulated in the presence of an equal number of autologous EBV-transformed B cells (irradiated with

25,000 RADS) with various concentrations (0-100 mg/ml) of recombinant Amb a I.1, I.2, I.3, I.4, Amb a II, or ragweed pollen extract in a volume of 200 ml complete medium in duplicate or triplicate wells in 96-well round bottom plates for 3 days. Each well then received 1 mCi tritiated thymidine for 16-20 hours. The counts incorporated were assessed and analyzed as described in Example VI. Table IV shows the S.I., P.I., percent of cultures positive for the assay antigen, and number of cultures analyzed. For these assays an individual S.I. greater or equal to 2.0 was used in calculating the mean S.I. Amb a I.1 stimulated cells respond less well and less frequently to the least homologous family members I.3 and I.2. However, this level of cross-reactivity, 68.8% and 60% respectively, is still considered high. The lowest level of Amb a I.1 stimulated T cell cross-reactivity is noted to the least homologous allergen, Amb a II. In addition, Amb a 1.1 reactive T cells are found at a high percentage from ragweed pollen extract stimulated cultures, again demonstrating the importance of the Amb a I.1 allergen.

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TABLE IV

	Assay Antigen			nt Amb a l.l	-
20		S.I.	% positive	n	P.I.
	Pollen Extract	13.5	87.8%	41	1185
721	r Amb a I.1	27.5	95.1%	41	2616
25	r Amb a I.2	5.0	60%	5	300
2	r Amb a I.3	9.3	68.8%	16	639
30	r Amb a I.4	8.5	100%	11	850
	r Amb a II	12.8	55.2%	29	70

Example IX T Cell Cross-Reactivity of Amb a 1.1 or Amb a 1.3 Stimulated Cells to Amb a 1.1 or Amb a 1.3 Peptides

Since a high degree of T cell cross-reactivity could be demonstrated between Amb a I.1 and the most highly sequence divergent family member Amb a I.3, the ability of Amb a I.1 stimulated cells to recognize both Amb a I.1 peptides and homologous Amb a I.3 peptides was assessed. In a similar fashion, the ability of Amb a I.3 stimulated cells to recognize both Amb a I.3 peptides and homologous Amb a I.1 peptides was determined. The peptides used in this study are listed below. The sequences for the Amb a I.1 homologous Amb a I.3 peptides were based on an alignment of the Amb a I.3 sequence to the Amb a I.1 sequence.

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Amb a 1.1 peptides

AMB 1-2.1 KGTVGGKDGDIYTVTSELDDDVAN

AMB 1-4.1 AENRKALADCAQGFGKGTVGGKDGD

AMB 2-1.1 GPAAPRAGSDGDAISISGSSQ

AMB 3-1.1 GSYAIGGSASPTILSQGNRFCAPDERSK

AMB 3-4.1 FFQVVNNNYDKWGSYAIGGSASPT

AMB 4-3.1 VLENGAIFVASGVDPVLTPEQSAGMIP

20 Amb a I.3 peptides

AMB 1-2.15 KGTYGGKWGDVYTVTSNLDDDVAN

AMB 1-4.15 ENNRQALADCAQGFAKGTYGGKWGD

AMB 2-1.15 GPPILRQASDGDTINVAGSSO

AMB 3-1.15 GTYAIGGSSAPTILCQGNRFLAPDDQIK

AMB 3-4.15 FFQVVNNNYDRWGTYAIGGSSAPT

AMB 4-3.15 LLENGAIFVTSGSDPVLTPVQSAGMIP

PBMC from ragweed allergic patients were stimulated with 20 mg/ml recombinant Amb a I.1 protein as described in Example VI with the addition of cultures of PBMC which were stimulated with 20 mg/ml recombinant Amb a I.3 protein. Assays were performed as described in Example VI except the homologous Amb a I.3 peptides were also tested at various doses. In these experiments an individual S.I of 2.0 or greater was used in calculating the mean S.I. Fig. 19 shows the results from a set of 9 matched patients stimulated with either recombinant Amb a I.1 protein or recombinant Amb a I.3 protein and tested on the set of Amb a I.1 peptides described above. Each bar represents the P.I. The dark bar represents cells stimulated with recombinant Amb a I.1 protein and tested with Amb a I.1 peptides, whereas, the stippled bar represents cells stimulated with recombinant Amb a I.3 protein and tested with Amb a I.3 protein and tested with Amb a I.1 peptides. With two exceptions, the results directly

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parallel each other, indicating that cells stimulated with recombinant Amb a I.3 proteins recognize Amb a I.1 derived peptides comprising at least one T cell epitope. One exception is that cells primed with Amb a I.3 protein recognize the RAE 7.D peptide poorly compared to recombinant Amb a I.1 stimulated cells. In addition, cells stimulated with recombinant Amb a I.3 protein gave stronger responses overall to Amb a I.1 derived peptides from Region 4 than the recombinant Amb a I.1 stimulated cells, particularly peptide RAE 28.1-2.

Cells were also stimulated with either recombinant Amb a I.1 or recombinant Amb a I.3 and tested for reactivity with Amb a I.3 derived peptides (Fig. 20). The results indicate that cells stimulated with recombinant Amb a I.1 protein recognize the Amb a I.3 derived peptides in a similar pattern to that of cells stimulated with recombinant Amb a I.3 protein. In Fig. 20 the bars represent the P.I. and the number above the bar the mean S.I. in the nine patients. The ".15" designation following the peptide name indicates that the peptide is derived from the Amb a I.3 protein. The data suggests a high degree of cross-reactivity at the T cell level between the Amb a I.1 protein and the Amb a I.3 protein.

Example X IgE Analysis With Peptides Derived From Amb a L1

To analyze IgE reactivity of peptides derived from the Amb a I.1 protein, a direct binding ELISA was performed according to the procedure described in Example III. The source of IgE for this analysis was a pool of ragweed allergic patient plasma from 38 ragweed skin test positive patients. The ELISA protocol was the same as Example III except that the antigen coating with peptides and proteins was a concentration of 10 mg/ml at 100 mls/well. Fig. 21 shows the graphed results of this assay demonstrating strong reactivity to both SPE and biochemically purified Amb a I.1. By this assay there is no detectable binding to any of the peptides. A set of these assays was also run with rabbit and mouse antisera to demonstrate that the coating of the peptides onto these plates was successful.

Example XI Further T Cell Epitope Fine Map Studies With Amb a L1

To further define the regions of Amb a I.1 specific T cell reactivity, selected peptides from Figure 14 were analyzed. Amb a I.1 specific T cells lines were derived from 28 ragweed allergic patients as described in Example VI. These lines were assessed for their ability to proliferate in response to the peptide in the presence of autologous EBV transformed antigen presenting cells by the uptake of tritiated thymidine as described previously. Several lysine substituted peptides derived from the peptide RAE 70.1 sequence were designed to increase the solubility of the peptide from Region 2. These peptides shown in Figure 22 were tested against non-substituted controls to determine if the modification resulted in a change in T cell reactivity. In addition, truncations of peptide AMB 4-6.1 (amino acid residues 347-377) were tested to further refine the relative T cell reactivity in that region.

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Figure 22 shows the responses of the 28 T cell lines to these peptides as analyzed by the positivity index, the mean stimulation index and the percent of positive responses. The positivity index, as defined previously, is the mean stimulation index multiplied by the percent of patients responding to a peptide. Responses to peptides were considered positive if they were greater or equal to 2 fold over background. Figure 22 demonstrates that certain lysine substituted peptides in the 182-216 sequence resulted in greater T cell responses, indicating that not all substitutions are recognized equally by a given T cell line. In addition, the T cell responses to the lysine modified peptides having a substitution of cysteine at position 212 with serine reflect the responses of T cell lines tested with peptides without the lysine substitutions. Thus, the decrease in response to AMB 2-10.1, with a substitution of leucine for cysteine at 212, reflects the decrease in the percentage of patients responding to this peptide relative to peptides AMB 2-9.1 and AMB 2-11.1 which contain serine and glutamic acid substitutions, respectively, at position 212.

In addition, the truncation peptides derived from peptide AMB 4-6.1 showed indistinguishable responses based on this analysis. Thus, the shortest sequence, 345-370 was chosen for further analysis. Substitutions at positions 360 and 361 were made to increase the stability of the AMB 4-9.1 peptide which contained an acid-sensitive aspartic acid-proline bond. As shown in Figure 23, three peptides were synthesized in which the proline at position 361 was substituted with either alanine, serine or glycine (AMB 4-9.1DA (SEQ ID NO:147), AMB 4-9.1DS (SEQ ID NO:148), and AMB 4-9.1DG (SEQ ID NO:149), 20 respectively). Similarly, five peptides were synthesized in which the aspartic acid at position 360 was substituted with the following: glutamic acid (AMB 4-9.1EP, SEQ ID NO:142); asparagine (AMB 4-9.1NP, SEQ ID NO:143); alanine (AMB 4-9.1AP, SEQ ID NO:144); serine (AMB 4-9.1SP, SEQ ID NO:145); and glutamine (AMB 4-9.1QP, SEQ ID NO:146). This group of modified peptides were tested to determine whether a change in T cell 25 reactivity occurred. 28 Amb a I.1 specific T cells lines were analyzed by the relative strength of the response to the modified peptides in a given T cell line compared to the overall response to all peptides from that line. Figure 23 shows the ranked sum of the strongest three peptide responses in the 28 T cell lines. The ranked sum was determined as described previously with the strongest response to a peptide given a value of 3, the second strongest a 30 2, and the third strongest a value of 1. The values from all 28 lines were then added to obtain the ranked sum. Figure 23 indicates that the modified peptide AMB 4-9.1DA elicits T cell responses similar to those of the native sequence AMB 4-9.1. In contrast, other substitutions, while eliciting strong responses do not rank as highly in this analysis. 35

Based in part on the above described analyses, five peptides were selected for further study (AMB 1-2.1, AMB 2-6.1, AMB 2-9.1, AMB 3-5.1 and AMB 4-9.1). An additional peptide, RA-02.1 (SEQ ID NO:150) was also synthesized and tested to confirm that the serine substitution at position 212 was recognized by Amb a I.1 specific T cell lines. The T cell reactivity of these peptides in 32 different Amb a I.1 specific T cell lines was determined

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as described previously. Figure 24 demonstrates the relative positivity indices, percent of positive responses and mean stimulation indices of these six peptides compared to the overall response to Amb a I.1. This figure shows that peptide AMB 4-9.1 elicits the strongest response in a high percentage of patients.

In contrast to previous data, the response to the 182-216 peptide (AMB 2-6.1) and substituted variants within this sequence (RA-02.1) gave weaker stimulation indices. This could reflect a potential toxic effect of the peptides in the T cell assays. To investigate this possibility, an assay was conducted to determine the ability of the peptides to inhibit T cell proliferation. Inhibition was defined as proliferation of a T cell line in response to the peptide which was less than half the proliferation of media control T cell lines plus autologous EBV transformed antigen presenting cells. As seen in Table V, the percent of patients in which inhibition of T cell proliferation is seen is far greater in cultures where the AMB 2-6.1, RA-02.1, and AMB 2-9.1 peptides are present as compared to AMB 1-2.1 or AMB 4-9.1. This data suggests that AMB 2-6.1, RA-02.1 and AMB 2-9.1 may be inhibitory due to toxicity in in vitro T cell line culture. This is further supported by the finding that these peptides also inhibit a T cell line from patient 956.2 which is specific for an irrelevant antigen, native Feld I, as shown in Figure 26.

ASSESSMENT OF PEPTIDE TOXICITY IN Amb a I.1
PRIMED SECONDARY T CELL CULTURE

25	peptide	#/(%)patients	low dose	high dose only	both
	AMB 1-2.1	3/46 (7)	2/3	1/3	. 0
	AMB 2-6.1	14/44 (32)	0	11/14	3/14
30 .	RA-02.1	19/45 (41)	0	15/19	4/19
,30	AMB 2-9.1	23/38 (61)	0	14/23	9/23
	AMB 4-9.1	1/46 (2)	0	0	1/1
35	AMB 3-5.1	19/46(41)	0	17/19	2/19

Legend: Toxicity defined as proliferation, as assessed by incorporation of tritiated thymidine, which is less than half the proliferation of media control of an Amb a I.1

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stimulated T cell line plus APC. Low dose = 5 mg/ml peptide in secondary assay, high dose = 50 mg/ml peptide.

A peptide from Region 2, RAE 70.1 (SEQ ID NO:44) was modified to minimize dimerization via disulfide linkages by substituting serine for cysteine at position 212. In addition, peptide RAE 70.1 was divided into two fragments and modified to increase solubility and to further define the residues necessary for T cell reactivity. As shown in Figure 25, a fragment of RAE 70.1 (amino acid residues 194-216) was modified by substitution or addition of amino acids which would increase the hydrophilicity and decrease the pI of the peptide to thereby increase solubility.

Peptides Amb 2-18.1 (SEQ ID NO:126), Amb2-19.1 (SEQ ID NO:127), Amb2-20.1 (SEQ ID NO:128) and Amb2-21.1 (SEQ ID NO:129) correspond to different lengths of the original peptide which were synthesized (including the serine substitution at position 212) and analyzed to determine solubility. It was found that peptide Amb2-19.1 (residues 200-217) was most soluble. Thus, substitutions and/or amino acid additions to this peptide were made in order to further increase the solubility while maintaining T cell reactivity. For example, in peptides Amb2-22.1 (SEQ ID NO:130) and Amb2-23.1 (SEQ ID NO:131) isoleucine at position 201 was substituted with glutamic acid (Amb2-22.1) or lysine (Amb2-23.1) to lower the pI of the resulting peptide and avoid precipitation of the peptide at physiological pH. Similarly, the following peptides were also synthesized with various substitutions or additions designed to decrease the pI and increase the hydrophilicity of the peptide, each is shown in Figure 25: Amb2-26.1 (SEQ ID NO:132); Amb2-28.1 (SEQ ID NO:133); Amb2-30.1 (SEQ ID NO:134); Amb2-32.1 (SEQ ID NO:135); Amb2-33.1 (SEQ ID NO:136); Amb2-34.1 (SEQ ID NO:137); Amb2-35.1 (SEQ ID NO:138); Amb2-36.1 (SEQ ID NO:139); Amb2-37.1 (SEQ ID NO:140); and Amb2-38.1 (SEQ ID NO:141).

Figures 27 and 28 show representative examples of proliferation of two individual Amb a I.1 specific T cell lines to peptides selected from those shown in Figure 25. Assays were performed as described previously. Both Figure 27 and Figure 28 indicate a hierarchy of responses to these peptides. Patient 119 shown in Figure 27, has a hierarchy of response from strongest to weakest as follows: AMB2-23.1>AMB2-22.1>AMB2-30.1>AMB2-26.1>AMB2-33.1>AMB2-32.1>AMB2-18.1>AMB2-19.1. Peptides AMB2-34.1, AMB2-35.1, and AMB 2-9.1 did not elicit significant T cell proliferation in this patient. Patient 1199 shown in Figure 28 demonstrated strong responses to AMB 2-26.1>AMB 2-22.1>AMB 2-33.1. T cells from this patient showed less than 2 fold over background responses to all other peptides. These responses are not significantly different from the media control of the T cell line plus autologous EBV antigen presenting cells.

To more closely examine the T cell epitopes within the above set of peptides to which Amb a I.1 specific T cell lines respond, a T cell clone was generated by limiting dilution from an Amb a I.1 specific T cell line stimulated with AMB 2-10.1. Briefly, an Amb a I.1 specific

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T cell line from patient 776 was shown to respond to AMB 2-10.1 in a proliferation assay as described previously. To generate a AMB 2-10.1 specific T cell clone, Amb a I.1 specific T cells were plated at 0.3 cells/well in a V-bottom 96 well plate with 20,000 irradiated autologous EBV transformed antigen presenting cells, AMB 2-10.1 at 40 mg/ml, leukoagglutin at 1 mg/ml, and recombinant human IL-2 and IL-4 at 10 units/ml in a total volume of 100ml/well. Plating the T cells at 0.3 cells/well insures not more than 1 T cell/well can potentially proliferate to the peptide with the progeny of that T cell representing a clonal population. After five days, wells received an additional 15 units/ml of recombinant Water. human IL-2 and IL-4 to expand peptide-specific T cells. This addition of IL-2 and IL-4 was repeated again every three days for the duration of the culture period. Twelve days after the 10 initiation of culture, T cells in the wells were restimulated with 20,000 irradiated EBV antigen presenting cells and 40 mg/ml AMB 2-10.1. This was repeated again 20 days after the initial stimulation. T cells from wells which showed signs of growth were separated from cell debris by density centrifugation as described previously in the generation of T cell lines. The T cell clone was then expanded with additional IL-2 and IL-4 at 10 units/ml until there 15 were significant numbers to assay for proliferation.

Assessment for T cell proliferation to selected peptides from those shown in Figure 25 was performed as described previously for T cell lines. The results of a representative assay using this AMB 2-10.1 specific T cell clone are shown in Figure 29. The data indicates that the T cell epitope recognized by this clone is contained by all the peptides except AMB 2-34.1 and AMB 2-35.1. Thus, a substitution of the tryptophan at position 208 eliminates the ability of the T cell clone to respond to these peptides. This data suggests that substitutions on the N and C terminal ends of the truncated peptides does not effect the recognition of the peptides by the T cell clone and, thus, does not alter the residues which comprise the T cell epitope.

Histamine Release Analysis With Purified Amb a I.1 and With Peptides Example XII Derived From Amb a 1.1

The objective of the histamine release analysis was to compare the effects of Amb a I.1 or Amb a I.1-derived peptides in an in vitro allergic response system. The release of histamine via IgE recognition and IgE receptor crosslinking on viable cells directly assays the allergic potential of a protein antigen.

The histamine release assay used for these studies is based on the detection of an acylated derivative of histamine using a specific monoclonal antibody (Morel, A.M. and Delaage, M.A. (1988) J. Allergy Clin. Immunol. 82:646-654). The assay was performed in two steps: 1) the release of histamine from basophils present in heparinized whole blood in the presence of different concentrations of protein or peptide; and 2) the assay of histamine present in the supernatants of the release reactions following cell removal by centrifugation.

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The reagents for this second step are available commercially as a competitive radioimmunoassay from AMAC Inc. (Westbrook, ME).

Heparinized whole blood was drawn from ragweed allergic patients. The test protein Amb a I.1 and peptides AMB 1-2.1, AMB 2-9.1, AMB 4-9.1, and AMB 3-5.1 were each diluted to 2X the final release concentration in PACM buffer (PIPES 25mM, NaCl 110mM, KCl 5.0 mM, human serum albumin 0.003% (w/v), CaCl₂ 5mM, MgCl₂ 2mM, pH7.3) and 0.2 ml of each dilution was added to a 1.5 ml polypropylene tube. A 0.2ml aliquot of blood was then added to each tube and the release reactions started by inversion of the tubes. A negative control of 0.2 ml blood incubated with 0.2 ml PACM buffer was also included. The release reactions were performed for 30 minutes at 37°C. The tubes were then centrifuged in a microfuge at 1500 RPM for 3 minutes and the supernatants were carefully removed and analyzed or stored at -20°C for later analysis. To analyze the total histamine released, 0.1 ml blood was diluted with 0.9 ml PACM buffer and boiled 3 minutes. This tube was then centrifuged 2 minutes at 12,000 RPM and the supernatant was removed and save for analysis.

For the competition radioimmunoassay, a 50 ml aliquot of each release supernatant was mixed with 150 ml of histamine release buffer (supplied with the kit from AMAC). The diluted supernatant (100 ml) was added to a kit tube containing an acylation reagent. A 50 ml aliquot of acylation buffer was then immediately added and the tube mixed by vortexing. The acylation reactions were incubated at least 30 minutes at room temperature. A set of histamine standards supplied with the kit were acylated at the same time. 50 ml of each acylation reaction was then placed in a tube coated with a monoclonal antibody which specifically recognizes the acylated form of histamine. A 0.5 ml aliquot of 125I-labelled tracer was added and the tubes were incubated at least 18 hours at 4°C. The solutions were aspirated from the tubes and the radioactivity bound to the tube was measured on a gamma counter (Cobra 5005, Beckman, Inc.) for two minutes per tube.

A standard curve was generated from the histamine standard counts and graphed on a semi-log plot. Since this is a competitive assay (the ¹²⁵-labelled tracer competes with acylated histamine in the samples for binding to the antibody-coated tubes), the lower the number of radioactive counts measured, the greater the amount of histamine in the sample. The amount of histamine in each sample data point was extrapolated from the standard curve using a computer statistical program (StatView for the MacIntosh). The assay is sensitive to 0.2 nM histamine.

To study whether peptides derived from the Amb a I.1 sequence could induce histamine release, blood samples from 8 ragweed-allergic patients who exhibited histamine release to native Amb a I.1 were analyzed with AMB 1-2.1, AMB 2-9.1, AMB 4-9.1 and AMB 3-5.1. Six or more 10-fold dilutions of Amb a I.1 starting at 10 mg/ml (approximately 0.66 mM) were analyzed. Five five-fold dilutions of each peptide were also analyzed (ranging from 50 mg/ml to 0.08 mg/ml, this is approximately 15 mM to 24 nM in concentration for each peptide). The peptide concentrations were selected to encompass the

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higher end of the concentration curve. Figure 30 shows the representative results from 1 of the patients (#1273). The graph shows the concentration of each antigen in mg/ml versus the percent of total histamine released. The results are presented as the percent of the total histamine released for that patient, since patients vary greatly in their overall releasable histamine levels. This variability is likely to result from variation in the number of basophils per ml between patients and the histamine content of each basophil. As Figure 30 shows, Amb a I.1, at all the concentrations tested, yields a high level of histamine release in this patient. Similar experiments using blood samples from other patients have shown that for some patients the Amb a I.1 concentration does not become limiting until 10-5-10-6 mg/ml. By contrast, there is no discernible histamine release to any of the four Amb a I.1-derived peptides which were used. Similar results were obtained for seven other patients similarly tested.

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In another experiment, plasma samples from 94 ragweed-allergic individuals, who tested positive for IgE binding to Amb a I.1 were analyzed for IgE binding to peptides Amb 1-2.1, Amb 2-36.1 and Amb 4-9.1 by direct binding ELISA as described above. Plasma samples were studied at a 1:3 and 1:30 dilution. None of the 94 patients tested had measurable IgE binding to either Amb 1-2.1 or Amb 2-36.1 (data not shown). Six patients had a low level of IgE binding to Amb 4-9.1 (data not shown). These results indicate that approximately 7% of ragweed-allergic patients have low plasma IgE levels to Amb 4-9.1

In order to assess the relative contribution of Amb 4-9.1-specific IgE to the overall level of IgE to Amb a I.1, plasma from one patient with IgE to Amb 4-9.1 was depleted of IgE to Amb 4-9.1 by repeatedly absorbing the plasma on plates coated with Amb 4-9.1-03 conjugated ovalbumin. A mock absorption was performed in parallel using plates coated with Amb 1-2.1 conjugated to ovalbumin. Previous experiments have shown that a somewhat higher O.D. signal can be obtained when the ovalbumin conjugate is used for plate coating. After absorption, the original, non-absorbed plasma samples were analyzed for IgE binding to Amb a I.1 in a direct assay. The efficiency of absorption was verified by also analyzing binding to Amb 4-9.1. The results indicated that absorption with Amb 4-9.1 efficiently removed any detectable binding to Amb 4-9.1. The mock absorption with 1-2.1 yielded a slight decrease in the O.D. signals seen in the unabsorbed sample, as indicated by the shift curve for Amb 1-2.1 absorbed plasma to the left of the curve for unabsorbed plasma The curve for Amb 4-9.1 shifted very slightly to the left of the Amb 1-2.1-absorbed plasma curve. This result suggests that IgE binding to Amb 4-9.1 represents a minor but measurable component of the overall level of IgE to Amb a I.1.

To further assess the relative contribution of anti-AMB 4-9.1-03 IgE to the overall anti-Amb a I.1 response, a competition ELISA was performed using Amb 4-9.1 and Amb a I.1 in solution to compete for binding to Amb a I.1. For a negative control, competition was also performed using Amb 1-2.1. This experiment shows that Amb 4-9.1 does not detectable compete for IgE binding to Amb a I.1 at concentrations where very effective competition is

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seen with soluble $Amb\ a\ I.1$. The results (data not shown) indicate that the change in IgE binding detected to Amb 4-9.1 when the plasma IgE response is competed with $Amb\ a\ I.1$ and Amb 4-9.1. As expected, incubation of the plasma sample with soluble Amb 4-9.1 can compete for binding to Amb 4-9.1. However, soluble $Amb\ a\ I.1$ competes much more efficiently since much lower concentrations (in μ /mL) of $Amb\ a\ I.1$ protein than Amb 4-9.1 peptide are required to compete for binding to Amb 4-9.1. This difference is even more distinct if the comparison is made on an equimolar basis since the peptide is in greater than ten-fold excess relative to $Amb\ a\ I.1$ at equivalent μ g/mL concentrations. This result suggests that in this patient sample, the portion of the IgE response detected which is directed to an epitope defined by the Amb 4-9.1 peptide sequence has lower affinity for Amb 4-9.1 than for $Amb\ a\ I.1$.

All peptide candidates were also examined for their ability to release histamine *in vitro* from basophils of ragweed-allergic patients. The objective of the histamine release analysis was to measure the effects of *Amb a* I.1 peptides Amb 1-2.1, Amb 2-36.1 and Amb 4-9.1 *in vitro* allergic response system. Heparinized whole blood from ragweed-allergic patients was incubated with different concentrations of each peptide (at $0.08-50 \,\mu g/mL$) or *Amb a* I.1 (at 10^{-7} to $10 \,\mu g/mL$). The levels of histamine release were measured using commercially available radioimmunoassay (Amac, Inc., Westbrook, ME). No patient with histamine release to native *Amb a* I.1 (n=19), released histamine to peptides Amb 1-2.1 (n=19), Amb 2-36.1 (n=19) and Amb 4-9.1 (n=14) (data not shown). Histamine release to Amb 4-9.1 was also not detected when blood from two patients with anti Amb 4-9.1 antibodies was analyzed (data not shown). The failure of Amb 4-9.1 to induce histamine release in two patients with IgE to Amb 4-9.1 is possibly due to the inability of the peptide to mediate the crosslinking of the IgE receptors on blood basophils.

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Example XIIIT Cell Responses to Ragweed Pentides

Peripheral blood mononuclear cells (PBMC) were purified by lymphocyte separation medium (LSM) centrifugation of 60 ml of heparinized blood from ragweed-allergic patients who exhibited clinical symptoms of seasonal rhinitis and were skin prick test positive for ragweed. T cell lines were established from these cells by stimulation of 1-2 x 106 PBMC/ml in RPMI-1640 containing 5% human AB serum (complete medium) with native Amb a I.1 at 20 mg/ml for 5-6 days at 37°C in a humidified CO₂ incubator. Viable cells were purified by LSM centrifugation and cultured in complete medium supplemented with 5 units recombinant human IL-2/ml and 5 units recombinant human IL-4/ml for up to three weeks until the cells no longer responded to lymphokines and were considered "rested". The ability of the T cells to proliferate to Amb a I.1 sequence-derived synthetic peptides was then assessed.

For assay, 2 x 10⁴ rested cells were restimulated in the presence of 2 x 10⁴ autologous Epstein-Barr virus (EBV)-transformed B cells (gamma-irradiated with 25,000

RADS) or 5 x 10⁴ autologous PBMC (3,500 RADS) with various concentrations of Amb a I.1 synthetic peptides in a volume of 200 ml complete medium in duplicate or triplicate wells in 96-well round bottom plates for 3 days. Each well then received 1 mCi tritiated thymidine for 16-20 hours. The counts incorporated were collected onto glass fiber filter mats and processed for liquid scintillation counting. Table III shows the results of a representative 5 assay. The maximum response in a titration of each peptide is expressed as the S.I. or stimulation index. The S.I. is the CPM incorporated by cells in response to peptide divided by the CPM incorporated by cells in medium only. An S.I. value greater than the background level is considered "positive" and indicates that the peptide contains a T cell epitope. However, only individual S.I. values above 2.0 (a response two-fold or greater over 10 background) were used in calculating mean stimulation indices for each peptide for the group of patients tested. The results shown in Table VI demonstrate that this patient (#1715) responds very well to peptides RAE 74.1, Amb 4-9.1, RAE 61.1, RAE 62.1. This indicates that these peptides contain Amb a I.1 T cell epitopes recognized by T cells from this particular allergic patient. . 15

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TABLE VI

	Antigen	Concentration (mg/ml)	СРМ	S.I.
5	Medium		452	1
	Amb a I.1	10	47822	
		100	63494	139.9
10	RAE 61.1	5	767	
	·	50	5910	13.0
	RAE 80.1	. 5	640	
		50	1067	2.4
15	RAE 45.1	5	589	
		50	1106	2.4
	RAE 75.1	5	1094	2.4
20		50	521	2.4
	RAE 62.1	5	2017	•
	10.12 02.1	50	4923	10.8
25	RAE 65.1	5	1048	2.3
•		50	462	2.3
	RW 31	5	802	•
	KW 31	50	892 1309	2.9
30		_		
•	RAE 74.1	5	18992	41.8
		50	10633	7.8
	AMB 4-9.1	5	14065	31.0
35		50	8018	

The above procedure was followed with a number of other patients. Individual patient results were used in calculating the mean S.I. for each peptide if the patient responded to the Amb a I.1 protein at an S.I. of 2.0 or greater and the patient responded to at least one peptide derived from Amb a I.1 at an S.I. of 2.0 or greater. A summary of positive experiments from 57 patients is shown in Fig. 31. The bar represents the cumulative rank of the peptide response in the 57 patients. To determine the cumulative rank, the 5 peptides with the highest S.I. in each patient were determined and assigned a numerical rank in descending order with 5 representing the strongest response. The ranks for each peptide were then summed in the 57 patients to determine the cumulative rank for the peptide. The number above each bar in paranthesis is the percentage of the positive responses (S.I. of 2.0 or greater) from the group of patients to that peptide. In parentheses above each bar is the mean S.I. of the positive responses. The cumaltive rank sum represents both the strength of the response (S.I.) and the frequency of a response to a peptide in the group of patients tested. For example, peptide AMB 4-9.1 had the highest cumulative rank so it was the best peptide response in the overall population of 57 even through it did not have the highest mean S.I. Similarly, RAE 27.1 had the highest mean S.I but not the best cumulative rank. Thus, the response to RAE 27.1 was strong when it occurred but it did not occur as frequently in the population as the response to other peptides. Based on the data as shown in Fig. 31, peptides AMB 4-9.1, RAE 74.1, AMB 3-1.1, AMB 2-36.1, RAE62.1 and AMB 1-2.1 were identified as the peptides with the strongest T cell reactivity

Example XIV Selection of Peptides For Use in Human Therapy

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To determine whether a peptide (candidate peptide) or a combination of candidate peptides are likely to comprise a sufficient percentage of T cell epitopes of ragweed pollen protein to induce T cell non responsiveness in a substantial percentage of a population of individuals sensitive to ragweed pollen protein and therefore, be a suitable composition for human therapy, the following evaluation is used. Candidate peptides are selected based, at least in part on the mean human T cell stimulation index for each of the candidate peptides in the set of peptides tested and the positivity index (the mean S.I. multiplied by the percent of positive responses) of the candidate peptides in the set of peptides tested. For example, the results in Example XIII (Fig. 31) indicate a number of peptides which contain strong T cell reactivity, and three of such peptides, Amb 1-2.1, Amb 2-36.1 and Amb 4-9.1, were selected as candidate peptides for use in human therapy. The human T cell stimulation index for each peptide tested including the stimulation index for the candidate peptides are divided by the sum of the human T cell stimulation indices of the remaining peptides in the set of peptides tested to determine a percent of T cell reactivity as shown below:

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% of T cell Reactivity of candidate peptides =

Sum of S.I.s for Each Candidate Peptide x 100

Sum of S.I.s of the set of overlapping peptides

A more liberal estimate of reactivity elicited by the candidate peptide, accounting for the overlaps giving rise to the total can be calculated as follows:

% of T cell Reactivity of Candidate Peptides =

N₁ flanking peptide S.I. + Candidate S.I. + C₁ flanking peptide S.I. x100

Sum of S.I.s from overlapping peptides

In this formula, "Nt flanking peptide" refers to a peptide which comprises amino acid resides which overlap with amino acid residues located at the N-terminus of the candidate peptide in the amino acid sequence of the protein antigen from which the peptide is derived; "Ct flanking peptide refers to a peptide which comprises amino acid residues which overlap with amino acid residues located at the C-terminus of the candidate peptide in the amino acid sequence of the protein antigen from which the peptide is derived. In this calculation, stimulation indices for the candidate peptide, the N-terminal flanking peptide and the C-terminal flanking peptide are added and divided by the sum total of the stimulation indices for the entire set of overlapping peptides to obtain a percent of T cell reactivity for the candidate peptides. If a combination of two or more candidate peptides is selected each of which contains amino acid residues which overlap, this calculation cannot be used to determine a percent of T cell reactivity for each candidate separately. However this is not the case for selected candidates, Amb 1-2.1, Amb 2-36.1 and Amb 4-9.1.

The values obtained for the percentage of T cell reactivity for the candidate peptides in each individual tested can be expressed as a range of lower and higher values of the results of the above described calculations. The mean percentage of T cell reactivity elicited by the candidate peptides can then be determined by averaging the values obtained for the individual responses to the candidate peptides.

For candidate peptides Amb 1-2.1, Amb 2-36.1 and Amb 4-9.1, secondary Amb a 1.1 T cell cultures derived from 57 ragweed-allergic subjects were analyzed for reactivity to an overlapping set of Amb a 1.1 peptides, and the highest stimulation index greater than or equal to 2.0 in response to each peptide was recorded for each individual tested. The data were then analyzed by the equation above for each individual tested, and the mean percentage of T cell reactivity was determined by averaging all the values obtained for the individual responses to the candidate peptides, Amb 1-2.1, Amb 2-36.1 and Amb 4-9.1, with the following results.

PCT/US95/14362

Candidate Peptides

Range of mean percentage of T cell reactivity

Frequency of response to at least one peptide

Amb 1-2.1 + Amb 2-36.1 + 26-35%

91%

Amb 4-9.1

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These values meet the criteria for a combination of candidate peptides described earlier.

5 Example XV Administration of Peptides to Humans for Treatment of Allergy to Ragweed A. Composition of Phase I Formulation

For Phase I clinical trials, the drug product was a multipeptide formulation comprising three peptides, Amb 1-2.1, Amb 2-36.1 and Amb 4-9.1, which together were determined to cover a sufficient percentage of T cell epitopes of a population of individuals as discussed in Example XIV. Each peptide was purified to homogeneity (at least 97% pure) by known methods and a multipeptide formulation was prepared in accordance with procedures discussed earlier. The multipeptide formulation used in this Phase I clinical Study was in the form of a freeze-dried powder cake comprising 1500 µg/L (when reconstituted) of each of the peptides, Amb 1-2.1, Amb 2-36.1 and Amb 4-9.1, in a single vial. The excipients used were 5% mannitol and 0.05 M sodium phosphate buffer system in a single vial. The formulation was reconstituted just prior to use with sterile water for injection resulting in a solution with a pH of ca. 7.5. Normal saline (0.9%) was used for any dilutions beyond initial reconstitution.

20 B. Human Phase I Clinical Studies

This study was a double blind placebo-controlled safety study of thirty-six patients conducted at Johns Hopkins Asthma and Allergy Center Baltimore. Twenty-four patients received active therapy while twelve received placebo.

After selection by demonstration skin test sensitivity to ragweed extract, the patients were tested for allergic sensitivity to study medication (active or placebo) prior to the first treatment by intracutaneous tests with increasing concentrations of the therapy. first Cohort (Cohort A) of sixteen patients received increasing weekly doses of 7.5, 75, 250, 750, and 1500 µg (per peptide) of the multipeptide formulation and four received placebo for five weeks. Cohort B of ten patients (6 active, 4 placebo) initiated a five week course of weekly fixed dose therapy (250 µg per peptide or placebo). After all patients in Cohort A completed the fourth week of therapy,, a third Cohort (Cohort C) of ten patients (6 active, 4 placebo) began a five-week course of weekly fixed dose therapy (750 µg per peptide or placebo). All patients underwent skin testing to ragweed extract and study medication (active or placebo) three weeks post treatment. Physical evaluation and clinical laboratory safety studies were done at baseline and at eight weeks.

Preliminary data indicates that all doses were tolerated and the multipeptide formulation appears to be safe. Although this study was designed to evaluate safety, preliminary data suggests that the multipeptide formulation may possibly be reducing late phase responses to ragweed allergen in the skin. Further statistical analysis of the Phase I Clinical studies are pending. Phase II Clinical Studies are pending in the United States and Canada

Equivalents:

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Although the invention has been described with reference to its preferred embodiments, other embodiments can achieve the same results. Variations and modifications to the present invention will be obvious to those skilled in the art and it is intended to cover in the appended claims all such modifications and equivalents that follow in the true spirit and scope of this invention.

- 54 -

SEQUENCE LISTING

_	(1) GENERAL INFORMATION:
5	(i) APPLICANT: Kuo, Mei-chang Garman, Richard Greenstein, Julia Evans, Sean
0	Amsberry, Kent Shaked, Ze'ev
5	(ii) TITLE OF INVENTION: T CELL EPITOPES OF THE MAJOR ALLERGEN FROM AMBROSIA ARTEMISIIFOLIA
3	(iii) NUMBER OF SEQUENCES: 150
	(iv) CORRESPONDENCE ADDRESS: (A) ADDRESSEE: IMMULOGIC PHARAMCEUTICAL CORPORATION
20	(B) STREET: 610 Lincoln Street (C) CITY: Waltham
v-	(D) STATE: MA (E) COUNTRY: USA (F) ZIP: 02154
25	(r) 21r. 0213.
	(v) COMPUTER READABLE FORM: (A) MEDIUM TYPE: Floppy disk
	(B) COMPUTER: IBM PC compatible
	(C) OPERATING SYSTEM: PC-DOS/MS-DOS
30	(D) SOFTWARE: ASCII Text
	(vi) CURRENT APPLICATION DATA: (A) APPLICATION NUMBER:
	(B) FILING DATE:
35	(C) CLASSIFICATION:
	(vi) PRIOR APPLICATION DATA:
	(A) APPLICATION NUMBER: US08/298,542
	(B) FILING DATE: August 30, 1994
40	(viii) ATTORNEY/AGENT INFORMATION:
	(A) NAME: Darlene A. Vanstone
	(B) REGISTRATION NUMBER: 35,729
	(C) REFERENCE/DOCKET NUMBER: 001.4 US
45	(ix) TELECOMMUNICATION INFORMATION:
	(A) TELEPHONE: (617) 466-6000
	(B) TELEFAX: (617) 466-6040
50	(2) INFORMATION FOR SEQ ID NO:1:
	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 1223 base pairs
55	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: single
	(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

5 (B) LOCATION: 1..1188

		(x	i) SI	EQUEN	ICE I	DESCI	RIPT	ON:	SEQ	ID 1	NO:1	:					
10	` Met	G GGG C Gl	G ATO	C AAA E Lys	CAC His	Cys	TG1 Cys	TAC	ATC	TTC Leu	туз	r TT	T ACC	C TT	A GCC	CTT a Leu	48
15	GT(Va)	Thi	r TTC	CTG Leu 20	Gln	CCI Pro	GTI Val	CG1	TCI Ser 25	Ala	GAA	A GAT	CTC Lev	CAC Glr 30	ı Glı	A ATC	96
20				Asn					Leu					Ala		AAC Asn	144
25			Asp										Ala			CGA Arg	192
.:	AAA Lys 65	Ala	TTA Leu	GCC Ala	GAT Asp	TGT Cys 70	GCC Ala	CAA Gln	GGT Gly	TTT Phe	GGG Gly 75	AAG Lys	GGA Gly	ACA Thr	GTG Val	GGC Gly 80	240
30	GGA Gly	AAA Lys	GAT Asp	GGT Gly	GAT Asp 85	ATA Ile	TAC Tyr	ACG Thr	GTC Val	ACC Thr 90	AGT Ser	GAG Glu	CTA Leu	GAT Asp	GAT Asp 95	GAT A sp	288
35				CCA Pro 100													336
40				TGG Trp										Arg			384
 45				GTG Val													432
				ATC Ile								Asn					480
50				CAT . His .					His								528
55			Ile	AAG ' Lys : 180				Gly					Arg .				576
	GAT Asp	GGT Gly	GAT Asp	GCT A	ATA .	AGT . Ser	ATT I	TCT (GGT :	AGT Ser	TCA Ser	CAA . Gln	ATA Ile	TGG Trp	ATC Ile	GAC Asp	624

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- 56 -

			195					200					205				
5		Cys 210	Ser	Leu	Ser	Lys	Ser 215	Val	Asp	GIY	Den	220	AU P	,,,,	-,-		672
	GGC I	Thr	Thr	Arg	Leu	Thr 230	Val	Ser	Asn	ser	235	PILE	****	011		240	720
10	TTT (Val	Leu	Leu	Phe 245	Gly	Ala	Gly	Asp	250	ASII	116	GIU	rsp	255	U _1	768
15 ⁶		Leu	Ala	Thr 260	Val	Ala	Phe	Asn	265	Pne	Int	Asp	YOU	270		-	816
20	AGA Arg	ATG Met	CCT Pro 275	AGA Arg	TGT Cys	CGA Arg	CAT His	GGG Gly 280	TTT Phe	TTC Phe	CAA Gln	GTC Val	GTT Val 285	21011	AAC Asn	AAC	864
25	TAT Tyr	GAT Asp 290	Lys	TGG Trp	GGA Gly	TCG Ser	TAT Tyr 295	GCC Ala	ATC	GGT Gly	GGT Gly	AGC Ser 300	AIG	TCC	CCA Pro	ACC	912
	ATA Ile 305	CTC Leu	AGC Ser	CAA Gln	GGG	AAC Asn 310	Arg	TTC Phe	TGC Cys	GCC Ala	Pro	ASP	GAA Glu	. CGC . Arg	AGC Ser	Lys 320	960
30	AAA Lys	AAT Asn	GTC Val	CTA Leu	GGA Gly 325	Arg	CAT His	GGT	GAA Glu	GCC Ala 330	Ald	GCA Ala	GAG	TCG Ser	ATG Met	AAG Lys	1008
35	TGG Trp	AAC Asn	TGG Trp	AGA Arg	Thr	TAA E	AAA Lys	GAC Asp	GT0 Val	r rer	GAA Glu	TAA A	GG7	GC7 Ala 350		TTT Phe	1056
10 40	GTT Val	GC# Ala	A TCC a Sea 35!	c Gly	GT(GA7 L Asi	CCA Pro	GTG Val	Let	A ACC	C CCT	GAC Glv	G CAI Glr 36		c GCA	A GGG a Gly	1104
45	ATG Met	AT:	e Pro	A GCC	GAI a Gl	A CC	A GG/ o Gl; 37	λ GT	TC Se	C GC' r Ala	r CTI a Le	A AGG u Se: 38		C AC	T AG' r Se:	T AGT r Ser	1152
	GCT Ala	a Gl	T GT. y Va	A CTO	C TC. u Se	A TG r Cy 39	s Gl	A CC	C GG o Gl	À YT	A CC a/Pr / 39	o cy	C TA	AGCA	CCCG		1198
50			TACT	AAG	CACT	TAT	AATG	A									1223

(2) INFORMATION FOR SEQ ID NO:2:

55 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 396 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

260

275

(ii)	MOLECULE	TYPE:	protein
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: 5 Met Gly Ile Lys His Cys Cys Tyr Ile Leu Tyr Phe Thr Leu Ala Leu Val Thr Leu Leu Gln Pro Val Arg Ser Ala Glu Asp Leu Gln Glu Ile 10 25 Leu Pro Val Asn Glu Thr Arg Arg Leu Thr Thr Ser Gly Ala Tyr Asn Ile Ile Asp Gly Cys Trp Arg Gly Lys Ala Asp Trp Ala Glu Asn Arg 50 Lys Ala Leu Ala Asp Cys Ala Gln Gly Phe Gly Lys Gly Thr Val Gly 20 Gly Lys Asp Gly Asp Ile Tyr Thr Val Thr Ser Glu Leu Asp Asp Asp 90 Val Ala Asn Pro Lys Glu Gly Thr Leu Arg Phe Gly Ala Ala Gln Asn 25 105 Arg Pro Leu Trp Ile Ile Phe Glu Arg Asp Met Val Ile Arg Leu Asp 120 30 Lys Glu Met Val Val Asn Ser Asp Lys Thr Ile Asp Gly Arg Gly Ala 130 Lys Val Glu Ile Ile Asn Ala Gly Phe Thr Leu Asn Gly Val Lys Asn 155 35 Val Ile Ile His Asn Ile Asn Met His Asp Val Lys Val Asn Pro Gly 165 170 Gly Leu Ile Lys Ser Asn Asp Gly Pro Ala Ala Pro Arg Ala Gly Ser 40 190 Asp Gly Asp Ala Ile Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp 45 His Cys Ser Leu Ser Lys Ser Val Asp Gly Leu Val Asp Ala Lys Leu 210 Gly Thr Thr Arg Leu Thr Val Ser Asn Ser Leu/Phe Thr Gln His Gln 230 235 50 Phe Val Leu Leu Phe Gly Ala Gly Asp Glu Asn Ile Glu Asp Arg Gly 245 250

Met Leu Ala Thr Val Ala Phe Asn Thr Phe Thr Asp Asn Val Asp Gln

Arg Met Pro Arg Cys Arg His Gly Phe Phe Gln Val Val Asn Asn Asn

280

- 58 -

	Tyr	Asp 290	Lys	Trp	Gly	Ser	Tyr 295	Ala	Ile	Gly	Gly	/ Se 30	r A	la :	Ser	Pro	Tì	nr		
5	305			Gln		310					31:	,								
				Leu	325					330	'									
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20	Ala 385		Val	Leu	Ser	Cys 390	Gln	Pro	Gly	/ Ala	a Pr 39	o C; 5	ys		. •					
t- <u>.</u>	(2)	INF	ORMA	TION	FOR	SEC	DI	NO : 3	:											
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40	Me	G GG t Gl 1	G AT y Il	C AA e Ly	s Hi	c TG s Cy 5	T TG s Cy	T TA	C AT	e re	rg T. eu T	AT T	rtt Phe	ACC	TTA Lev		C a 5	CTT Leu		48
45	GT Va	C AC	T TI	G CT u Le	G CA u Gl 0	A CC n Pr	T GT O Va	T CG 1 Ar	g se	T GO er Al	CA G la G	AA (GAT Asp	GTI Val	GAI Glu		A u	TTC Phe		96
50	TI Le	A CC	o Se	CA GC er Al	T AA .a As	C GF	A AC	r Ar	G A0	eg A	GC C er L	TG . Leu	AAA Lys	GCF Ala	3	r GA s Gl	u.u	GCA Ala		144
55	C# H:	is A		TT AT	TA GA	C Al	ys C)	C TO	G AC	GG T rg C	GC F ys I	ràs ryy	GCC Ala 60	, AJ	r TG p Tr	g gc p Al	CG La	TAA neA		192
33	A			AA GO	CG T	eu A	CC GA la As	AT TO	GT G ys A	CC C	AA (GGT Gly 75	TTT Phe	GC:	A AA a Ly	G G 's G	GA ly	ACC Thr 80		240

						s Gly				Thi	val				p Ly	A GAT s Asp	288
5	CNT			. aa						90					9!	_	
					a Ası					Thi					Ala	GCC A Ala	336
10				g Pro					Phe					. Val		CAT His	384
15			ı Glr					Asn					.Ile			CGA	432
20		Va]					Val					Thr				GTC Val 160	480
25						CAT His											528
25					Ile	AAG Lys											576
30				Gly		GCT Ala											624
35						CTC Leu											672
40						CAC His 230									Thr		720
45						TTG Leu											768
						ACG Thr											816
50						AGA Arg	Cys					Phe					864
55	AAC Asn					Trp					Ile						912
	CCA .																960

- 60 -

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	305					310					315						32		
5	ATC Ile	AAG Lys	AAA Lys	AAT Asn	GTC Val 325	TTA Leu	GCG Ala	AGG Arg	ACT Thr	GGT Gly 330	ACT Thr	GGC Gly	AAC	C GO	ra .	SAG Slu 335	TC Se	G r	1008
	ATG Met	TCG Ser	TGG Trp	AAC Asn 340	TGG Trp	AGA Arg	ACA Thr	GAT Asp	AGA Arg 345	GAC Asp	TTG Leu	CTT Leu	GAJ Glu	4 24	AT (sn (GGT	GC Al	T a	1056
10	ATT Ile	TTT Phe	CTC Leu 355	CCA Pro	TCC Ser	GGG Gly	TCT Ser	GAT Asp 360	CCA Pro	GTG Val	CTA Leu	ACC Thr	CC'	o G	AG (CAA Gln	AA Ly	A 's	1104
15	GCA Ala	GGG Gly 370	Met	ATT	CCA Pro	GCT Ala	GAA Glu 375	CCA Pro	GGA Gly	GAA Glu	GCC Ala	GTT Val 380	Le	A A u A	.GA .rg	CTC Leu	AC Th	T ir	1152
20	AGT Ser 385	Ser	GCT Ala	GGT	GTA Val	CTC Leu 390	TCA Ser	TGC Cys	CAT His	CAA Gln	GGA Gly 395	GCA Ala	CC Pr	T T	GC ys	TAA	-		1197
	GCA	CCTG	GCC	AATT	CCTA	AG C	TTTT	ATAA	AA T	TCAI	TAAA	ACI	TAT	TTI	T A7	TTT.	ATT	TTTT	1260
25	GAT	ATTI	TAT	ATGA	ACCA	A TT.	CGTT	CAAG	T AC	TCT	AATT.	CAT	GTT	TT	AA A	TTC	ΑTĄ	AAGA	1320
	GTT	TATI	GAT	АААА	AAAA	A AA	AAAC	CGAA	T TC	:									1349
30	(2)	INF					ID			_									•
35			(i)	() ()	4) LE 3) TY	ENGTH PE:	RACT : 39 amin)GY:	8 an	nino cid	acio	ls	i							
			(ii)	MOL	ECULI	E TY	PE: p	rote	ein										
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- 61 -

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				100)				109	5				110)	
5	Glı	n As	n Arg) Leu	Tr	o Ile	120		E Ly:	s Arg	Asn	Met 125		l Ile	His
-	Let	13:		a Glu	ı Lev	Val	. Val		ı Ser	. Ası	o Lys	Thr 140		e Asp	Gly	Arg
10	Gly 145		l Lys	3 Val	. Asn	11e		. Asn	Ala	Gly	/ Leu 155		Leu	Met	Asn	Val 160
	Lys	Ası	ı Ile	: Ile	11e		Asn	Ile	: Asn	11e		Asp	Ile	. Lys	Val 175	Сув
15	Pro	Gly	/ Gly	Met 180		Lys	Ser	Asn	Asp 185		Pro	Pro	Ile	Leu 190	Arg	Gln
20	Gln	Ser	195		Asp	Ala	Ile	Asn 200		Ala	Gly	Ser	Ser 205		Ile	Trp
	Ile	Asp 210		Cys	Ser	Leu	Ser 215	Lys	Ala	Ser	Asp	Gly 220	Leu	Leu	Asp	Ile
25	Thr 225	Leu	Gly	Ser	Ser	His 230	Val	Thr	Val	Ser	Asn 235	Cys	Lys	Phe	Thr	Gln 240
	His	Gln	Phe	Val	Leu 245	Leu	Leu	Gly	Ala	Asp 250	Asp	Thr	His	Tyr	Gln 255	Asp
30	Lys	Gly	Met	Leu 260	Ala	Thr	Val	Ala	Phe 265	Asn	Met	Phe	Thr	Asp 270	His	Val
35	Asp	Gln	Arg 275		Pro	Arg	Cys	Arg 280	Phe	Gly	Phe	Phe	Gln 285	Val	Val	Asn
	Asn	Asn 290	Tyr	Asp	Arg	Trp	Gly 295	Thr	Tyr	Ala	Ile	Gly 300	Gly	Ser	Ser	Ala
40	Pro 305	Thr	Ile	Leu	Ser	Gln 310	Gly	Asn	Arg	Phe	Phe 315	Ala	Pro	Asp	Asp	Ile 320
	Ile	Lys	Lys	Asn	Val 325	Leu	Ala	Arg	Thr	Gly 330	Thr	Gly	Asn	Ala	Glu :	Ser
45	Met	Ser	Trp	Asn 340	Trp	Arg	Thr	Asp	Arg 345	Asp	Leu	Leu	Glu	Asn 350	Gly	Ala
50	Ile	Phe	Leu 355	Pro	Ser	Gly	Ser	Asp 360	Pro	Val	Leu /		Pro 365	Glu	Gln 1	Lys
	Ala	Gly 370	Met	Ile	Pro	Ala	Glu 375	Pro	Gly	Glu		Val 380	Leu	Arg	Leu :	Thr
55	Ser 385	Ser	Ala	Gly		Leu 390	Ser	Cys	His	Gln	Gly . 395	Ala :	Pro	Cys		

PCT/US95/14362

WO 96/13589

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1320 base pairs(B) TYPE: nucleic acid

- 62 -

5				STF TOP					re									
		(ii)	MOLI	ECULI	E TY	PE: C	DNA											
10		(ix)	(A)	TURE:) NAI	ME/K	EY: (CDS	191										
•	-																	
15				UENC													•.*	
	ATG	GGG	ATC .	AAA (CAA	TGT '	TGT '	TAC :	ATC	TTG	TAT	TTT	ACC	TTA	GCA	CTT	4,8	
	Met 1	Gly	Ile	Lys	Gln 5	Cys	Cys '	Tyr	Ile	Leu 10	Tyr	Phe	Thr	Leu	Ala 15	Leu :		
20	GTC						amar .	ССТ	ጥርጥ	GCC	GAA	GGT	GTC	GGG	GAA	ATC	. 96	
	GTC Val	GCT Ala	TTG Leu	Leu 20	Gln	Pro	Val	Arg	Ser 25	Ala	Glu	Gly	Val	Gly 30	Glu	Ile		
				GTT			> 00	200	NGC.	CTG	CAA	GCA	TGT	GAA	GCA	CTC	144	
25	TTA Leu	Pro	TCA Ser 35	GTT Val	AAC Asn	GAA	Thr	Arg 40	Ser	Leu	Gln	Ala	Cys 45	Glu	Ala	Leu		
				GAC	220	TCC	TGG	NGG	GGC	AAA	GCC	GAT	TGG	GAG	AAC	AAC	192	
30	AAC Asn	Ile 50	Ile	Asp	Lys	Суѕ	Trp 55	Arg	Gly	Lys	Ala	Asp 60	Trp	Glu	Asn	Asn	. •	
				TTA		~1.0	mom	ccc	CAA	GGT	الململة :	GCA	AAG	GGA	ACC	TAC	240	
35	Arg	CAA Gln	GCG Ala	TTA Leu	GCC Ala	Asp 70	Cys	Ala	Gln	Gly	Phe 75	Ala	Lys	Gly	Thr	Tyr 80		
33										ama	7.00	NCC	አእጥ	СТА	CAT	GAT	288	
the the s	GGC	GGA	AAA	TGG Trp	GGT	GAT	GTC	TAC	ACG Thr	Val	Thr	Ser	Asn	Leu	Asp	Asp		
443	Gly	Gly	Lys	Trp	61y 85	Asp	Vai	I y I		90					95		-	
40												mma		CCC	GCC	ממי	336	;
	GAT Asp	GTT Val	GCA Ala	AAT Asn 100	Pro	AAA Lys	GAA Glu	GGC Gly	ACA Thr 105	Leu	CGG Arg	Phe	Ala	Ala 110		CAA Gln		
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45	AAC Asn	AGG	g Pro	Leu	TGG	ATC lle	ATT Ile	Phe	rys	AA1 ASD	Asp	Met	Val 125		Asn	TTG Leu		
			115								/						432	,
	LAA	CA	A GAG	CTI	GTO	GTA	AAC	AGC	GAC	: AAC	ACC	ATO	GAT	GGC	CGF	GGG Gly	434	•
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				: Ile	Lys				Gly	Pro				ı Arg	Glr	GCA Ala	576
5	3.00	. ~		180					185					190			
				/ Asp					Ala					Île		ATA Ile	624
10			Cys										Val			ACC	672
15		Gly										Lys		ACC Thr			720
20											Thr			CAA Gln			768
25														AAC Asn 270			816
	_													GTT Val			864
30														TCG Ser			912
35														GAT Asp			960
40														GAG Glu		Met	1008
45														GGT Gly 350			1056
43							Asp							CAA Gln			1104
50						Glu								CTC .			1152
55				GTA Val										TAAG	CACC	CT	1201
	GCCA	ATTC	TC C	TAAG	СТТТ	T GC.	AATG:	ATCA	AAA	ATAC	TTT	TTTA	TTTT	'AT T	TTTA	ATATT	1261

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- 64 -

TTATATGTAC TGGAAATGAA CCATTACCTT CTAGTACTCT ATAACATGTT TTGCATTTA (2) INFORMATION FOR SEQ ID NO:6: 5 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 397 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear 10 (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6: Met Gly Ile Lys Gln Cys Cys Tyr Ile Leu Tyr Phe Thr Leu Ala Leu 15 10 5 1 Val Ala Leu Leu Gln Pro Val Arg Ser Ala Glu Gly Val Gly Glu Ile 25 20 Leu Pro Ser Val Asn Glu Thr Arg Ser Leu Gln Ala Cys Glu Ala Leu 45 40 Asn Ile Ile Asp Lys Cys Trp Arg Gly Lys Ala Asp Trp Glu Asn Asn 25 Arg Gln Ala Leu Ala Asp Cys Ala Gln Gly Phe Ala Lys Gly Thr Tyr Gly Gly Lys Trp Gly Asp Val Tyr Thr Val Thr Ser Asn Leu Asp Asp 30 90 Asp Val Ala Asn Pro Lys Glu Gly Thr Leu Arg Phe Ala Ala Ala Gln 105 35 Asn Arg Pro Leu Trp Ile Ile Phe Lys Asn Asp Met Val Ile Asn Leu 120 Asn Gln Glu Leu Val Val Asn Ser Asp Lys Thr Ile Asp Gly Arg Gly 40 130 Val Lys Val Glu Ile Ile Asn Gly Gly Leu Thr Leu Met Asn Val Lys 150 Asn Ile Ile Ile His Asn Ile Asn Ile His Asp Val Lys Val Leu Pro 45 170 165 Gly Gly Met Ile Lys Ser Asn Asp Gly Pro Pro Ile Leu Arg Gln Ala 185 50 Ser Asp Gly Asp Thr Ile Asn Val Ala Gly Ser Ser Gln Ile Trp Ile Asp His Cys Ser Leu Ser Lys Ser Phe Asp Gly Leu Val Asp Val Thr 55 210

Leu Gly Ser Thr His Val Thr Ile Ser Asn Cys Lys Phe Thr Gln Gln

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- 65 -

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	Se	r Ly:	s Ala	a Ile	245		Gly	Ala	a Asp	250		His	Val	Gln	Asp 255	Lys	
5	Gly	y Mei	t Lei	260		Val	Ala	Phe	265		Phe	Thr	Asp	Asn 270		Asp	
	Glr	ı Arg	7 Met 275		Arg	Cys	Arg	Phe 280		Phe	Phe	Gln	Val 285	Val	Asn	Asn	
10	Asn	Туз 290	Asp	Arg	Trp	Gly	Thr 295	Туг	Ala	Ile	Gly	Gly 300	Ser	Ser	Ala	Pro	
15	Thr 305	Ile _	e Leu	Cys	Gln	Gly 310	Asn	Arg	Phe	Leu	Ala 315	Pro	Asp	Asp	Gln	Ile 320	
	Lys	Lys	Asn	Val	Leu 325	Ala	Arg	Thr	Gly	Thr 330	Gly	Ala	Ala	Glu	Ser 335	Met	
20	Ala	Trp	Asn	Trp 340	Arg	Ser	Asp	Lys	Asp 345	Leu	Leu	Glu	Asn	Gly 350	Ala	Ile	
	Phe	Val	Thr 355	Ser	Gly	Ser	Asp	Pro 360	Val	Leu	Thr	Pro	Val 365	Gln	Ser	Ala	
25	Gly	Met 370	Ile	Pro	Ala	Glu	Pro 375	Gly	Glu	Ala	Ala	Ile 380	Lys	Leu	Thr	Ser	
30	Ser 385	Ala	Gly	Val	Phe	Ser 390	Cys	Arg	Pro	Gly	Ala 395	Pro	Cys				
	(2)	INF	ORMAT	rion	FOR	SEO	TD N	·0 · 7 ·									
35) SE((<i>I</i> (E	QUENC A) LE B) TY	E CH NGTH PE: RAND	ARAC : 11 nucl EDNE	TERI 87 b eic SS:	STIC ase acid sing	S: pair	s		ì					
40		(ii)	MOL	ECUL	E TY	PE:	cDNA								•		
45	·	(ix)		.) NA	: ME/KI CATIO			176									
		(xi)	SEQ	UENC	E DES	SCRII	PTIO	1: S	EQ II	ONO:	: 7 :						
50	ATG Met																4.8
55	GTC .																96
	TTA	CCT	TCA (GCT A	AAC G	AA A	CA A	.GG A	AGC C	CTG A	.CA A	CA I	GT G	GA A	CA I	AC	144

Leu Pro Ser Ala Asn Glu Thr Arg Ser Leu Thr Thr Cys Gly Thr Tyr

•									-	66 -								
			35					40					45					
5	AAC Asn	ATT Ile 50		GAC Asp	GGG Gly	TGC '	TGG Trp 55	AGG Arg	GGC Gly	AAA Lys	GCC Ala	GAT Asp 60	TGG Trp	GCG Ala	GAA Glu	AAC Asn		192
	CGA Arg 65	AAA Lys	GCG Ala	TTA Leu	GCC Ala	GAT Asp 70	TGT Cys	GCC Ala	CAA Gln	GGT Gly	TTT Phe 75	GCA Ala	AAG Lys	GGA Gly	ACA Thr	ATC Ile 80		240
10	GGC Gly	GGA Gly	AAA Lys	GAT Asp	GGT Gly 85	GAT Asp	ATA Ile	TAC Tyr	ACG Thr	GTC Val 90	ACC Thr	AGT Ser	GAG Glu	CTA Leu	GAT Asp 95	GAT Asp		288
15 	GAT Asp	GTT Val	GCA Ala	AAT Asn 100		AAA Lys	GAA Glu	GGC Gly	ACA Thr 105	CTC Leu	CGG Arg	TTT Phe	GGT Gly	GCC Ala 110	GCC Ala	CAA Gln		336
20	AAC Asn	AGG Arg	CCC Pro	Leu	TGG	ATT Ile	ATT Ile	TTT Phe 120	GAA Glu	AGA Arg	GAT Asp	ATG Met	GTG Val 125	ATT	CGT	TTG Leu		384
25	GAT Asp	AGA Arg 130	Glu	TTG Lev	GCT Ala	ATA Ile	AAC Asn 135	AAC Asn	GAC Asp	AAG Lys	ACC Thr	ATC Ile 140	ASP	GGC	CGA Arg	GGG		432
	GCG Ala 145	Lys	GTI Val	GAF Glu	ATC	ATT Ile 150	Asn	GCT Ala	GGT	TTC Phe	GCC Ala 155	. 116	TAT Tyr	AAT Asn	GTC Val	Lys 160		480
30	AAT Asn	ATA	ATO	ATT	CAT His	Asn	ATA Ile	ATT	ATG Met	CAT His	Asp	ATT Ile	GTA Val	GTG Val	AAT Asr 175	CCA Pro		528
35	GG# Gly	GGG Gly	CTC Let	3 AT u Il 18	e Lys	TCC Ser	CAC His	GAT Asp	GGT Gly 185	PIC	CCA Pro	GTI Val	CCA Pro	AGA Arg 190	, ~,-	GGT Gly		576
40	AG: Sei	r GA'	r GG p Gl	y As	T GCT p Ala	ATA 11e	A GGT	7 AT7 7 Ile 200	e Sei	r GGT	r GG:	r TC! y Sei	CAA Gli 20		A TG(ATC o Ile		624
45	GA(As)	C CA p Hi 21	s Cy	C TC	C CTO	C AG	r AA0	s Ale	r GT a Vai	r GA' l As	r GG(p Gl	G CTA y Let 22	u 11.	C GA' e As	T GC' p Al	T AAA a Lys		672
ų.	CA Hi 22	s Gl	C AG y Se	C AC	A CA	C TT s Ph	e Th	C GT r Va	T TC	T AA r As	C TG n/Cy / 23	a ne	A TT u Ph	C AC e Th	c ca r Gl	A CAC n His 240	·	720
50	CA G1	A TA n Ty	T TI	TA T	TA TT eu Le 24	u Ph	C TG e Tr	G GA p As	T TT p Ph	T GA e As 25	b Gr	G CG u Ar	A GG g Gl	c Al y Me	G CT t Le 25	A TGT u Cys		768
55	AC TT	G G	rc Go	la P	IC AA he As	c AA	G TI	C AC	T GA ir As 26	ip As	C GI	T GA	C CA		SA AI ig Me	G CCT	; >	816

AAC TTA CGA CAT GGG TTT GTC CAA GTC GTT AAC AAC AAC TAC GAA AGA

260

										- 07	-						
	Ası	n Le	u Arg 275		Gly	'Phe	. Val	Gln 280		Val	. Asn	Asn	Asn 285	_	Glu	Arg	
5	TG(G GG G1 ₂ 29	y Ser	TAC Tyr	GCC Ala	CTC Leu	GGT Gly 295	Gly	AGC Ser	GCA Ala	GGC	CCA Pro 300	Thr	ATA Ile	CTI Leu	AGC Ser	912
10	CAZ Glr 305	Gl	AAC Asn	AGA Arg	TTC Phe	TTA Leu 310	GCC Ala	TCC Ser	GAT Asp	ATC Ile	AAG Lys 315	AAA Lys	GAG Glu	GTC Val	GTA Val	GGG Gly 320	960
15	. AGG Arg	TAT	GGT Gly	GAA Glu	TCC Ser 325	GCC Ala	ATG Met	TCA Ser	GAG Glu	TCG Ser 330	ATT Ile	AAT Asn	TGG Trp	AAC Asn	TGG Trp 335	AGA Arg	1008
	TCG Ser	TAT	ATG Met	GAC Asp 340	GTA Val	TTT Phe	GAA Glu	AAT Asn	GGT Gly 345	GCT Ala	ATT	TTT Phe	GTT Val	CCA Pro 350	TCC Ser	GGG Gly	1056
20	GTT Val	GAT Asp	CCA Pro 355	GTG Val	CTA Leu	ACC Thr	CCT Pro	GAG Glu 360	CAA Gln	AAC Asn	GCA Ala	GGG Gly	ATG Met 365	ATT Ile	CCA Pro	GCC Ala	1104
25	GAA Glu	CCA Pro 370	GGA Gly	GAA Glu	GCC Ala	GTT Val	CTA Leu 375	AGA Arg	CTC Leu	ACT Thr	AGT Ser	AGT Ser 380	GCT Ala	GGT Gly	GTC Val	CTC Leu	1152
30			CAA Gln						TAAG	CACT	GC A						1187
35	(2)		ORMAT	EQUE (A) (B)	NCE LEN TYP		ACTE 392 mino	RIST ami aci	ICS: no a d	cids		1					
40		(:	ii) M	OLEC	ULE	TYPE	: pr	otei:	n							•	
45	Met 1		ci) S Ile										Thr 1	Leu <i>i</i>	Ala 1	Leu	
		Thr	Leu	Leu (20		Pro 1	Val 1	Arg :	Ser 1 25		Glu <i>I</i>	Asp I	Ceu (Gln (Ile	
50	Leu	Pro	Ser . 35	Ala i	Asn (3lu 1	Thr 1	Arg S	Ser I	Leu 1	Thr I	Thr (Cys (Sly ?	Thr T	Tyr	
55	Asn	Ile 50	Ile i	Asp (Gly (Cys T	Trp #	Arg (Sly I	ys I	Ala A	sp T	rp A	lla (Slu <i>I</i>	Asn	
	Arg 65	Lys	Ala 1	Leu A	Ala A	Asp C 70	ys A	Ala G	Sln G	Sly F	Phe A 75	la I	ys G	ly 1	hr 1	le 80	

Gly Gly Lys Asp Gly Asp Ile Tyr Thr Val Thr Ser Glu Leu Asp Asp

- 68 -

					85					90					95	
	Asp	Val	Ala	Asn 100	Pro	Lys	Glu	Gly	Thr 105	Leu	Arg	Phe	Gly	Ala 110	Ala	Gln
5	Asn	Arg	Pro 115	Leu	Trp	Ile	Ile	Phe 120	Glu	Arg	Asp	Met	Val 125	Ile	Arg	Leu
10.	Asp	Arg 130	Glu	Leu	Ala	Ile	Asn 135	Asn	Asp	Lys	Thr	Ile 140	Asp	Gly	Arg	Gly
	Ala 145	Lys	Val	Glu	Ile	Ile 150	Asn	Ala	Gly	Phe	Ala 155	Ile	Tyr	Asn	Val	Lys 160
15	Asn	Ile	Ile	Ile	His 165	Asn	Ile	Ile	Met	His 170	Asp	Ile	Val	Val	Asn 175	Pro
	Gly	Gly	Leu	Ile 180	Lys	Ser	His	Asp	Gly 185	Pro	Pro	Val	Pro	Arg 190	Lys	Gly
20	Ser	Asp	Gly 195	Asp	Ala	Ile	Gly	11e 200	Ser	Gly	Gly	Ser	Gln 205	Ile	Trp	Ile
25	Asp	His 210		Ser	Leu	Ser	Lys 215	Ala	Val	Asp	Gly	Leu 220	Ile	Asp	Ala	Lys
	His 225	Gly	Ser	Thr	His	Phe 230	Thr	Val	Ser	Asn	Cys 235	Leu	Phe	Thr	Gln	His 240
30	Gln	Tyr	Leu	Leu	Leu 245	Phe	Trp	Asp	Phe	Asp 250	Glu	Arg	Gly	Met	Leu 255	Cys.
25	Thr	Val	Ala	Phe 260		Lys	Phe	Thr	Asp 265	Asn	Val	Asp	Gln	Arg 270	Met	Pro
35 .2197 .2197	Asn	Leu	Arg 275		Gly	Phe	Val	Gln 280		Val	Asn	Asn	Asn 285	Tyr	Glu	. Arg
40	Trp	Gl ₃		Tyr	Ala	Leu	Gly 295	Gly	Ser	Ala	Gly	Pro 300	Thr	lle	Leu	Ser
	Gln 305		/ Asr	n Arg	Phe	Leu 310		Ser	Asp) Ile	215	Lys	Glu	val	Val	Gly 320
45	Arg	ту:	c Gly	y Glu	325		Met	. Ser	Glu	330	Ile	. Asn	Trp) Asn	335	Arg
	Sei	r Ty:	r Met	Asp 340		L Phe	e Glu	ı Ası	34!	/ Ala	4/11e	Phe	· Val	350	Ser	Gly
50	Va:	l As	p Pro		l Le	ı Thi	r Pro	360	ı Glı	n Ası	n Ala	a Gly	7 Met 36!	t Ile	Pro	Ala
55	Gl	u Pr 37		y Gli	ı Ala	a Va:	1 Let 37!	ı Arç	g Le	u Th	r Sei	380	Ala O	a Gly	y Val	l Leu
	Se	r Cy	s Gl	n Pro	o G1	y Ala	a Pr	о Су	s							

390

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145

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(2	INFORMATION	FOR	SEQ	ID	NO:	9 :	:
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(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1395 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

15 (B) LOCATION: 1..1191

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

		(11	., SE	ZOEN	CE L	ESCR	IPTI	ON:	SEQ	ID V	0:9:						
20		Gly				Cys					Tyr					CTT Leu	48
25					Gln										Ile	TTA Leu	96
30	CCT Pro	TCA Ser	CCT Pro 35	Asn	GAT A sp	ACA Thr	AGG Arg	AGG Arg 40	AGC Ser	CTG Leu	CAA Gln	GGA Gly	TGT Cys 45	GAA Glu	GCA Ala	CAC	144
35			Ile	GAC A sp													192
33				TTA Leu													240
40				TGG Trp													288
45				AAT Asn 100													336
50				TTG Leu			Ile										384
55	Gln			ATG Met		Val											432

GCG AAA GTT GAG CTC GTT TAT GGA GGT ATC ACC CTC ATG AAT GTC AAG

Ala Lys Val Glu Leu Val Tyr Gly Gly Ile Thr Leu Met Asn Val Lys

155

150

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							•		_	, 0								
	AAT Asn	GTA Val	ATC Ile	ATT Ile	CAC His	AAC Asn	ATA (GAT Asp	ATC Ile	CAT His 170	GAT Asp	GTT Val	AGA Arg	GTG Val	CTT Leu 175	CCA Pro		528
5	GGA Gly	GGT Gly	AGG Arg	ATT Ile 180	AAG Lys	TCC Ser	AAT Asn	GGT Gly	GGT Gly 185	CCA Pro	GCC Ala	ATA Ile	CCA Pro	AGA Arg 190	CAT His	CAG Gln		576
10	AGT Ser	GAT Asp	GGT Gly 195	GAT Asp	GCT Ala	ATC Ile	CAT His	GTT Val 200	ACG Thr	GGT Gly	AGT Ser	TCA Ser	GAC Asp 205	ATA Ile	TGG Trp	ATC Ile		624
15	GAC Asp	CAT His 210	TGC Cys	ACG Thr	CTC Leu	AGT Ser	AAG Lys 215	TCA Ser	TTT Phe	GAT Asp	GGG Gly	CTC Leu 220	GTC Val	GAT Asp	GTC Val	AAC Asn		672
	TGG Trp 225	GGC Gly	AGC Ser	ACA Thr	GGA Gly	GTA Val 230	ACC Thr	ATT Ile	TCC Ser	AAC Asn	TGC Cys 235	AAA Lys	TTC Phe	ACC	CAC	CAC His 240		720
20	GAA Glu	AAA Lys	GCT Ala	GTT Val	TTG Leu 245	CTC Leu	GGG Gly	GCT Ala	AGT Ser	GAC Asp 250	Thr	CAT	TTT Phe	CAA Gln	GAT Asp 255	CTG Leu	٠	768
25	AAA Lys	ATG Met	CAT His	GTA Val 260	Thr	CTT Leu	GCA Ala	TAC Tyr	AAC Asn 265	He	TTC Phe	ACC	AAT Asn	ACC Thr 270	vai	CAC His		816
30	GAA Glu	AGA Arg	ATO Met	Pro	AGA Arg	TGC Cys	CGA Arg	TTT Phe 280	Gly	TTT Phe	TTC Phe	CAF	ATC 116 285	; val	AAC Asr	AAC Asn		864
35	TTC Phe	TAC Ty:	Ası	AGA	TGG Trp	GAT Asp	AAG Lys 295	Tyr	GCC Ala	ATC	GGT Gly	GG1 Gly 300	/ SE1	TCC Sea	AAG ASI	C CCT		912
K.A.	ACT Thr	: Ile	r CTO	C AGO	C CAF	GGG Gly 310	ASD	AAA Lys	TTC Phe	GTC Val	GCC Ala	1 PI	C GAT	r TTO	C ATT	TAC Tyr 320		960
40	AAC Lys	AA Ly:	A AA s As	C GT(n Va	C TG:	s Lev	A AGG	ACI	r GG7 r Gly	GC/ Ala 330	. G11	G GA	g CC	A GA	A TGG u Trj 33	G ATG p Met 5		1008
45	AC:	r TG r Tr	G AA p As	C TG n Tr	p Ar	A ACI	A CAI	A AA(C GAG n Asj 34	o va.	G CT	T GA u Gl	A AA' u As	T GG n Gl 35	,	T ATC	! !	1056
50	TT Ph	T GT e Va	G GC	a Se	T GG r Gl	G TC y Se	T GA' r As	r cc p Pr 36	o va	G CT	A AC u Th	C GC r Al	T GA a Gl 36	u 01	A AA n As	T GCA n Ala	A A	1104
55	GG G1	C A1 y Me 37	t Me	G CA	A GC n Al	T GA a Gl	A CC u Pr 37	O GI	A GA y As	T AT p Me	G GT t Va	T CC 1 Pr 36		A CI n Le	C AC	C ATO	3 t	1152
	AA As 36	n Al	CA GO	ST GI Ly Va	TA CI	C AC	r Cy	C TC	G CC	T GG	A GC y Al 39	a F	CT TO	C TA	AAGC#	ACCTG		1201

- 71 -

	GC	CAAT	TCCT	ATG	CAAC	GAT	CATA	AATA	ст т	GCTC	ACCA	T AA	GTGT	TCAT	TTG	ATTAGAT	1261
5	TT	GGAC	ACGA	ATG	ATGT.	AAC	CGAT	TCGT	CT G	AATT	ATGA	T TT	GTTT	TGAT	TCT	CAGTTTC	1321
J	AT	AATA	rggc	TTC	rtga(GAG (CAAA	ATTA	GA G	AAGA	GTGT	C TT	rgat	CAAC	TAC	ATTTTAT	1381
	GGT	ETTT?	TATA	TTA	A.												1395
10	` (0)																
	(2)	INI					Q ID										•
15			(1)	(<i>)</i>	A) LI 3) TY	ENGTI (PE :	ARACT H: 39 amir OGY:	97 an	nino cid		ls			. •.			
		((ii)	MOLE	CULE	TY	E: p	rote	in					٠			
20		(xi)	SEQU	ENCE	DES	CRIP	MOIT	: SE	Q II	NO:	10:					
	Met 1		Ile	. Lys	His 5		: Cys	Tyr	Ile	Leu 10	_	Phe	Thr	Leu	Ala 15	Leu	
25	Val	Thr	Leu	Val 20		Ala	Gly	Arg	Leu 25	_	Glu	Glu	Val	Asp 30		Leu	·
30	Pro	Ser	Pro 35		Asp	Thr	Arg	Arg 40		Leu	Gln	Gly	Cys 45		Ala	His	
	Asn	Ile 50		Asp	Lys	Cys	Trp 55	Arg	Cys	Lys	Pro	Asp 60	_	Ala	Glu	Asn	
35	Arg 65	Gln	Ala	Leu	Gly	Asp 70		Ala	Gln	Gly	Phe 75	Gly	Lys	Ala	Thr	His 80	
	Gly	Gly	Lys	Trp	Gly 85	Asp	Ile	Tyr	Met	Val 90	Thr	Ser	Asp	Gln	Asp 95	Asp	
40	Asp	Val	Val	Asn 100	Pro	Lys	Glu	Gly	Thr 105	Leu	Arg	Phe	Gly	Ala 110	Thr	Gln	
45	Asp	Arg	Pro 115	Leu	Trp	Ile	Ile	Phe 120	Gln	Arg	Asp	Met	Ile 125	Ile	Tyr	Leu	
	Gln	Gln 130	Glu	Met	Val	Val	Thr 135	Ser	Asp	Thr	Thr	Ile 140	Asp	Gly	Arg	Gly	
50	Ala 145	Lys	Val	Glu	Leu	Val 150	Tyr	Gly	Gly	Ile	Thr 155	Leu	Met	Asn	Val	Lys 1 ⁰	
	Asn	Val	Ile	Ile	His 165	Asn	Ile	Asp	Ile	His 170	Asp	Val	Arg	Val	Leu 175	Pro	
55	Gly	Gly	Arg	Ile 180	Lys	Ser	Asn	Gly	Gly 185	Pro	Ala	Ile	Pro	Arg 190	His	Gln	
	Ser	Asp	Gly	Asp	Ala	Ile	His	Val	Thr	Gly	Ser	Ser	Asp	Ile	Trp	Ile	

200

205

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	Asp	His 210	Cys	Thr	Leu	Ser	Lys 215	Ser	Phe	Asp	Gly	Leu 220	Val	Asp	Val	Asn
5	Trp 225	Gly	Ser	Thr	Gly	Val 230	Thr	Ile	Ser	Asn	Cys 235	Lys	Phe	Thr	His	His 240
	Glu	Lys	Ala	Val	Leu 245	Leu	Gly	Ala	Ser	Asp 250	Thr	His	Phe	Gln	Asp 255	Leu
10	Lys	Met	His	Val 260	Thr	Leu	Ala	Tyr	Asn 265	Ile	Phe	Thr	Asn	Thr 270	Val	His
15	GĪu	Arg	Met 275	Pro	Arg	Cys	Arg	Phe 280	Gly	Phe	Phe	Gln	11e 285	Val	Asn	Asn
	Phe	Tyr 290		Arg	Trp	Asp	Lys 295	Tyr	Ala	Ile	Gly	Gly 300	Ser	Ser	Asn	Pro
20	Thr 305		Leu	Ser	Gln	Gly 310	Asn	Lys	Phe	Val	Ala 315	Pro	Asp	Phe	Ile	Tyr 320
	Lys	Lys	Asn	Val	Cys 325		Arg	Thr	Gly	Ala 330	Gln	Glu	Pro	Glu	Trp 335	Met
25	Thr	Trp	Asn	Trp 340		Thr	Gln	Asn	Asp 345	val	Leu	Glu	Asn	Gly 350	Ala	Ile
30	Ph∈	e Val	Ala 355		Gly	/ Ser	Asp	360	val	Leu	Thr	Ala	365	Gln	Asn	Ala
	Gly	y Met 370		Gl:	n Ala	a Glu	375	Gly	/ Asp) Met	: Val	380	Glr	. Lev	Thr	Met
35 	As :		a Gly	y Va	l Lei	u Thi 390	r Cys	s Sei	r Pro	o Gly	/ Ala 39	a Pro	Cys	5		
<i></i>	(2						Q ID								•	
		(i) S	(A) (B) (C)	LENG TYPE STRA	TH: : nu NDED	ACTE 312 clei NESS : li	base c ac : si	pai id ngle							
45		(i	.i) M				: cD				,					
50		(i	ix) E	(A)	NAMI	E/KEY	7: CI)S 276	;		<i>!</i>					

ATG TCG ATT CTT GGA GGA ATT ACC GAA GTT AAA GAC AAT GAT AAC AGC Met Ser Ile Leu Gly Gly Ile Thr Glu Val Lys Asp Asn Asp Asn Ser 1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

5					Glu					Ala					s As	C AAG n Lys	96
J	AAC Lys	GAG Glu	AAT Asn 35	Ala	GCT Ala	CTG Leu	GAG Glu	TTT Phe	Gly	AAI Lys	A GTA s Val	A ATA	GAZ Glu	ı Lys	A AAG	G CAG S Gln	144
10	CAG `Gln	GCG Ala 50	Val	CAG Gln	GGC Gly	ACC	ATG Met 55	Tyr	TAT Tyr	ATA Ile	A AAA e Lys	GTG Val	Glu	A GCA	AA? ASI	GAT Asp	192
15	GGT Gly 65	Gly	GAG Glu	AAG Lys	AAA Lys	ACT Thr 70	TAT Tyr	GAA Glu	GCC Ala	AAG Lys	GTG Val 75	Trp	GTT Val	Lys	CTA Lev	TGG Trp 80	240
20				AAG Lys							Leu	GTT Val	TGA	TGTT	GCC		286
	ACC	TCAC	CTTA	ACTO	CAT	ATG (GACG	3									312
25	(2)	INFO	ORMAT	NOIT	FOR	SEQ	ID 1	NO:12	2:								
30			(i) S	(B)	LEN	CHAP IGTH: PE: a POLOG	92 mino	amir aci	no ac								
				OLEC								1					
35	Met 1			EQUE Leu								l2: Asp	Asn	Asp	Asn 15	Ser	
40	Val	Asp	Phe	Asp 20	Glu	Leu	Ala	Lys	Phe 25	Ala	Ile	Ala	Glu	His 30	Asn	Lys	
	Lys	Glu	Asn 35	Ala .	Ala	Leu	Glu	Phe 40	Gly	Lys	Val	Ile	Glu 45	Lys	Lys	Gln	
45	Gln		Val	Gln (Gly	Thr	Met 55	Tyr	Tyr	Ile	Lys	Val 60		Ala	Asn	Asp	·
50	Gly 65	Gly	Glu	Lys :	Lys	Thr '	Tyr	Glu /	Ala	Lys	Val 75	Trp	Val	Lys	Leu	Trp 80	
	Glu	Asn	Phe :	Lys (Glu : 85	Leu (Gln (Glu 1	Leu :	Lys 90	Leu	Val					

- 55 (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 27 amino acids
 - (B) TYPE: amino acid

	<i>-</i> 74 <i>-</i>
	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
5	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:
10	Tyr Ile Leu Tyr Phe Thr Leu Ala Leu Val Thr Leu Leu Gln Pro Val 1 5 10 15
15	Arg Ser Ala Glu Asp Leu Gln Glu Ile Leu Pro 20 25
	(2) INFORMATION FOR SEQ ID NO:14:
20	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 amino acids (B) TYPE: amino acid
;	(C) STRANDEDNESS: single(D) TOPOLOGY: linear
25	(ii) MOLECULE TYPE: peptide
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:
30	Pro Val Arg Ser Ala Glu Asp Leu Gln Glu Ile Leu Pro Val Asn Glu 1 5 10 15
	Thr Arg Arg Leu Thr Thr Ser Gly Ala Tyr Asn Ile 20 25
35	
	(2) INFORMATION FOR SEQ ID NO:15:
40	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
45	(ii) MOLECULE TYPE: peptide
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
50	Glu Thr Arg Arg Ser Leu Lys Thr Ser Gly Ala Tyr Asn Ile Ile Asy 1 5 10 15
	Cly Cyc Trp Arg Gly Lys Ala Asp

(2) INFORMATION FOR SEQ ID NO:16:

55

(i) SEQUENCE CHARACTERISTICS:

	(A) LENGTH: 23 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
5	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
	Glu Thr Arg Arg Leu Thr Thr Ser Gly Ala Tyr Asn Ile Ile Asp Gly 1 5 10 15
15	Cys Trp Arg Gly Lys Ala Asp
20	(2) INFORMATION FOR SEQ ID NO:17:
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
25	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
35	Ala Tyr Asn Ile Ile Asp Gly Cys Trp Arg Gly Lys Ala Asp Trp Ala 1 5 10 15
33	Glu Asn Arg Lys Ala Leu Ala Asp Cys Ala Gln Gly 20 - 25
40	(2) INFORMATION FOR SEQ ID NO:18:
45	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
45	(ii) MOLECULE TYPE: peptide
50	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
55	Arg Gly Lys Ala Asp Trp Ala Glu Asn Arg Lys Ala Leu Ala Asp Cys 1 5 10 15
	Ala Gln Gly

	(2) INFORMATION FOR SEQ ID NO:19:	
	(i) SEQUENCE CHARACTERISTICS:	
5	(A) LENGTH: 10 amino acids	
	(B) TYPE: amino acid	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
10	(v) FRAGMENT TYPE: internal	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
15		
	Gly Lys Ala Asp Trp Ala Glu Asn Arg Cys	
	1 5 10	
20	(2) INFORMATION FOR SEQ ID NO:20:	
20		
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 10 amino acids (B) TYPE: amino acid	
25	(D) TOPOLOGY: linear	
25	(D) 10F0D031. 12M0D1	
	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
30		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
	(X1) SEQUENCE DESCRIPTION. SEQ 15 1001	
	Gly Lys Ala Asp Trp Ala Glu Asn Arg Lys	
35	1 5 10	
	-	
	(2) INFORMATION FOR SEQ ID NO:21:	
40	(i) SEQUENCE CHARACTERISTICS:	
40	(A) LENGTH: 13 amino acids	
	(B) TYPE: amino acid	
	(D) TOPOLOGY: linear	
45	(ii) MOLECULE TYPE: peptide	
	() and Gumm mype, internal	
	(v) FRAGMENT TYPE: internal	
	<i>,</i>	
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
	Ala Glu Asn Arg Lys Ala Leu Ala Asp Cys Ala Gln Gl	У
	Ala Giu Ash Arg bys Ala 201	

55 (2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 17 amino acids

```
(B) TYPE: amino acid
                 (D) TOPOLOGY: linear
          (ii) MOLECULE TYPE: peptide
  5
           (v) FRAGMENT TYPE: internal
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:
 10
           Lys Ala Leu Ala Asp Cys Ala Gln Gly Phe Gly Lys Gly Thr Val Gly
                                                                     15
         - Gly
15
      (2) INFORMATION FOR SEQ ID NO:23:
20
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 18 amino acids
                (B) TYPE: amino acid
                (D) TOPOLOGY: linear
25
          (ii) MOLECULE TYPE: peptide
           (v) FRAGMENT TYPE: internal
30
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:
          Gly Phe Gly Lys Gly Thr Val Gly Gly Lys Asp Gly Asp Ile Tyr Ser
35
          Val Thr
     (2) INFORMATION FOR SEQ ID NO:24:
40
          (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 18 amino acids
               (B) TYPE: amino acid
               (D) TOPOLOGY: linear
45
         (ii) MOLECULE TYPE: peptide
          (v) FRAGMENT TYPE: internal
```

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Gly Phe Gly Lys Gly Thr Val Gly Gly Lys Asp Gly Asp Ile Tyr Thr

10

55 Val Thr

```
(2) INFORMATION FOR SEQ ID NO:25:
          (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 17 amino acids
                (B) TYPE: amino acid
 5
                (D) TOPOLOGY: linear
         (ii) MOLECULE TYPE: peptide
          (v) FRAGMENT TYPE: internal
-10
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:
          Lys Asp Gly Asp Ile Tyr Ser Val Thr Ser Glu Leu Asp Asp Val
15
           Ala
20
      (2) INFORMATION FOR SEQ ID NO:26:
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 17 amino acids
 25
                (B) TYPE: amino acid
                (D) TOPOLOGY: linear
          (ii) MOLECULE TYPE: peptide
 30
           (v) FRAGMENT TYPE: internal
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:
           Lys Asp Gly Asp Ile Tyr Thr Val Thr Ser Glu Leu Asp Asp Asp Val
 35
                            5
            Ala
 40
       (2) INFORMATION FOR SEQ ID NO:27:
            (i) SEQUENCE CHARACTERISTICS:
                 (A) LENGTH: 11 amino acids
  45
                 (B) TYPE: amino acid
                 (D) TOPOLOGY: linear
           (ii) MOLECULE TYPE: peptide
  50
             (v) FRAGMENT TYPE: internal
            (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
  55
             Ile Tyr Thr Val Thr Ser Glu Leu Asp Asp Asp
                             5
```

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- 79 -
      (2) INFORMATION FOR SEQ ID NO:28:
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 14 amino acids
  5
                (B) TYPE: amino acid
                (D) TOPOLOGY: linear
          (ii) MOLECULE TYPE: peptide
 10
          (v) FRAGMENT TYPE: internal
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
 15
          Ile Tyr Thr Val Thr Ser Glu Leu Asp Asp Asp Val Ala Asn
      (2) INFORMATION FOR SEQ ID NO:29:
 20
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 17 amino acids
                (B) TYPE: amino acid
                (D) TOPOLOGY: linear
25
         (ii) MOLECULE TYPE: peptide
          (v) FRAGMENT TYPE: internal
30
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:
          Ile Tyr Thr Val Thr Ser Glu Leu Asp Asp Val Ala Asn Pro Lys
35
          Glu
40
     (2) INFORMATION FOR SEQ ID NO:30:
          (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 amino acids
               (B) TYPE: amino acid
45
               (D) TOPOLOGY: linear
         (ii) MOLECULE TYPE: peptide
          (v) FRAGMENT TYPE: internal
50
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:
         Ile Tyr Ser Val Thr Ser Glu Leu Asp Asp Val Ala Asn Pro Lys
55
```

Glu Gly Thr Leu

5	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear 	
10	(ii) MOLECULE TYPE: peptide	
10	(v) FRAGMENT TYPE: internal	
1.6	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:	
.15	Ile Tyr Thr Val Thr Ser Glu Leu Asp Asp Asp Val Ala Asn Pro Lys 1 10 15	
20	Glu Gly Thr Leu 20	
	(2) INFORMATION FOR SEQ ID NO:32:	
25	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 amino acids	
	(B) TYPE: amino acid (D) TOPOLOGY: linear	
30	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:	
	Asn Pro Lys Glu Gly Thr Leu Arg Phe Gly Ala Ala Gln Asn Arg Pro 1 5 10 15	
40	Leu Trp Ile Ile Phe Glu Arg Asp Met Val Ile Arg Leu 20 25	
45	(2) INFORMATION FOR SEQ ID NO:33:	
43	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 amino acids	
	(B) TYPE: amino acid (D) TOPOLOGY: linear	
50	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
55	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:	
	Trp Ile Ile Phe Glu Arg Asp Met Val Ile Arg Leu Asp Lys Glu Met 1 5 10	t

Val Val Asn Ser 20

5

10

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 19 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

15 (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Leu Asp Lys Glu Met Val Val Asn Ser Asp Lys Thr Ile Asp Gly Arg

1 5 10 15

Gly Ala Lys

25

30

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 26 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

35

40

45

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

Asp Lys Thr Ile Asp Gly Arg Gly Ala Lys Val Glu Ile Ile Asn Ala 1 5 10 15

Gly Phe Thr Leu Asn Gly Val Lys Asn Val 20 25

(2) INFORMATION FOR SEQ ID NO:36:

50 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

55 (ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

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	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:	
	Gly Phe Thr Leu Asn Gly Val Lys Asn Val Ile Ile His Asn Ile Asn 1 10 15	
5	Met His Asp Val Lys Val Asn Pro Gly Gly Leu 20 25	
10	(2) INFORMATION FOR SEQ ID NO:37:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
15	(ii) MOLECULE TYPE: peptide	
20	(v) FRAGMENT TYPE: internal	ı
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:	
25	His Asp Val Lys Val Asn Pro Gly Gly Leu Ile Lys Ser Asn Asp Gly 1 5 10 15	•
	Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala 20 25	
30	(2) INFORMATION FOR SEQ ID NO:38:	
35	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 23 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
40	(v) FRAGMENT TYPE: internal	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:	
45	Asn Pro Gly Gly Leu Ile Lys Ser Asn Asp Gly Pro Ala Ala Pro Ar 1 5 10	g
	Ala Gly Ser Asp Gly Asp Ala	
50	(2) INFORMATION FOR SEQ ID NO:39:	
55	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 amino acids (B) TYPE: amino acid	

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

	(v) FRAGMENT TYPE: internal
5	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:
	Ile Lys Ser Asn Asp Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly 1 5 10 15
10	Asp Ala
15	(2) INFORMATION FOR SEQ ID NO:40:
13	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 13 amino acids(B) TYPE: amino acid
20	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
25	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:
30	Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala 1 5 10
	(2) INFORMATION FOR SEQ ID NO:41:
35	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 13 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
40	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:
	Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile 1 5 10
50	(2) INFORMATION FOR SEQ ID NO:42:
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 18 amino acids(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

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	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:
5	Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile Ser Gly Ser 1 10 15
	Ser Gln
10	
	(2) INFORMATION FOR SEQ ID NO:43:
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 23 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
20	(v) FRAGMENT TYPE: internal
	CEO ID NO.43:
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:
	Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile Ser Gly Ser 1 15
30	Ser Gln Ile Trp Ile Asp His 20
	(2) INFORMATION FOR SEQ ID NO:44:
35	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
40	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:
	Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile Ser Gly Ser 10 15
50	Ser Gln Ile Trp Ile Asp His Cys Ser Leu Ser Lys 20 25
•	(2) INFORMATION FOR SEQ ID NO:45:

(i) SEQUENCE CHARACTERISTICS:

(B) TYPE: amino acid
(D) TOPOLOGY: linear

(A) LENGTH: 28 amino acids

	(ii) MOLECULE TYPE: peptide
5	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:
10	Trp Ile Asp His Cys Ser Leu Ser Lys Ser Val Asp Gly Leu Val Asp 1 5 10 15
	Ala Lys Leu Gly Thr Thr Arg Leu Thr Val Ser Asn 20 25
15	(2) INFORMATION FOR SEQ ID NO:46:
20	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
25	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:
30	Leu Gly Thr Thr Arg Leu Thr Val Ser Asn Ser Leu Phe Thr Gln His 1 5 10 15
35	Gln Phe Val Leu Phe Gly Ala Gly Asp Glu 20 25
	(2) INFORMATION FOR SEQ ID-NO:47:
40	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
45	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:
	Phe Val Leu Leu Phe Gly Ala Gly Asp Glu Asn Ile Glu Asp Arg Gly 1 5 10 15
55	Met Leu Ala Thr Val Ala Phe Asn Thr Phe Thr Asp 20 25

(2) INFORMATION FOR SEQ ID NO:48:

	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 18 amino acids (B) TYPE: amino acid
	(D) TOPOLOGY: linear
5	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:
	Leu Ala Thr Val Ala Phe Asn Thr Phe Thr Asp Asn Val Asp Gln Arg
15	Met Pro
20	(2) INFORMATION FOR SEQ ID NO:49:
20	(i) SEQUENCE CHARACTERISTICS:
••	(A) LENGTH: 20 amino acids (B) TYPE: amino acid
	(D) TOPOLOGY: linear
25	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:
	Phe Thr Asp Asn Val Asp Gln Arg Met Pro Arg Cys Arg His Gly Phe 1 5 10 15
35	Phe Gln Val Val
n vida April I	20
40	(2) INFORMATION FOR SEQ ID NO:50:
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 amino acids (B) TYPE: amino acid
45	(D) TOPOLOGY: linear
43	(ii) MOLECULE TYPE: peptide
50	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:
55	Arg Cys Arg His Gly Phe Phe Gln Val Val Asn Asn Asn Tyr Asp Ly 1 5 10 15
	Trp Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro

	(2) INFORMATION FOR SEQ ID NO:51:
5	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 25 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
10	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
15	-4xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:
	His Gly Phe Phe Gln Val Val Asn Asn Tyr Asp Lys Trp Gly Set 1 5 10 15
20	Tyr Ala Ile Gly Gly Ser Ala Ser Pro 20 25
	(2) INFORMATION FOR SEQ ID NO:52:
25	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 22 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
30	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:
	Phe Gln Val Val Asn Asn Asn Tyr Asp Lys Trp Gly Ser Tyr Ala Ile 1 5 10 15
40	Gly Gly Ser Ala Ser Pro
45	(2) INFORMATION FOR SEQ ID NO:53:
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
50	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
55	(vi) SEQUENCE DECORIDATION, SEC. ID NO. 52

Val Asn Asn Tyr Asp Lys Trp Gly Ser Tyr Ala Ile Gly Gly Ser

10

15

Ala Ser Pro

Gly Asn Arg Phe

5	(2) INFORMATION FOR SEQ ID NO:54:	
ю	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 12 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
15	(v) FRAGMENT TYPE: internal	•
, •	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:	<i>t</i> ,
20	Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr 1 5 10	
	(2) INFORMATION FOR SEQ ID NO:55:	
25	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 16 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
30	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:	
40	Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr Ile Leu Ser G 1 5 10 15	lln
	(2) INFORMATION FOR SEQ ID NO:56:	
45	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 20 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
50	(ii) MOLECULE TYPE: peptide	
50	(v) FRAGMENT TYPE: internal	
_	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:	
55	Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr Ile Leu Ser 1 5 10 15	Glr

5	(2) INFORMATION FOR SEQ ID NO:57:
10	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
15	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:
20	Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr Ile Leu Ser Gln 1 5 10 15
	Gly Asn Arg Phe Cys Ala Pro Asp Glu Arg Ser Lys 20 25
25	(2) INFORMATION FOR SEQ ID NO:58:
30	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
35	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:
40	Arg Phe Cys Ala Pro Asp Glu Arg Ser Lys Lys Asn Val Leu Gly Arg 1 10 .15
	His Gly Glu Ala Ala Glu Ser Met Lys Trp Asn 20 25
45	(2) INFORMATION FOR CEO ID NO.50.
•	(2) INFORMATION FOR SEQ ID NO:59:
50	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 26 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
55	(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

	- 70	
	Glu Ala Ala Ala Glu Ser Met Lys Trp Asn Trp Arg Thr Asn Lys As 1 5 10 15	φ
5	Val Leu Glu Asn Gly Ala Ile Phe Val Ala 20 25	
	(2) INFORMATION FOR SEQ ID NO:60:	
19	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 23 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
15	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
	· ····	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:	
	Glu Ser Met Lys Trp Asn Trp Arg Thr Asn Lys Asp Val Leu Glu A 1 5 10 15	sr.
25	Gly Ala Ile Phe Val Ala Ser 20	
20	(2) INFORMATION FOR SEQ ID NO:61:	
30	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
35	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
40	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:	
	Glu Phe Gly Trp Arg Thr Asn Lys Asp Val Leu Glu Asn Gly Ala 1 5 10 15	Il
45	Phe Val Ala Ser 20	
50	(2) INFORMATION FOR SEQ ID NO:62:	
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 20 amino acids(B) TYPE: amino acid	

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:
5	Lys Trp Asn Trp Arg Thr Asn Lys Asp Val Leu Glu Asn Gly Ala Ile 1 10 15
10	Phe Val Ala Ser 20
	(2) INFORMATION FOR SEQ ID NO:63:
15	- (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
20	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:
	Lys Trp Asn Trp Arg Thr Asn Lys Asp Val Leu Glu Asn Gly Ala Ile 1 5 10 15
30	Phe Val Ala Ser Gly Val Asp Pro Val Leu Thr Pro Glu Gln 20 25 30
	(2) INFORMATION FOR SEQ ID NO:64:
35	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 17 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
40	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:
	Trp Arg Thr Asn Lys Asp Val Leu Glu Asn Gly Ala Ile Phe Val Ala 1 5 10 15
50	Ser
55	(2) INFORMATION FOR SEQ ID NO:65:
JJ	(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 14 amino acids

(B) TYPE: amino acid(D) TOPOLOGY: linear

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55

	(ii) MOLECULE TYPE: peptide
_	(v) FRAGMENT TYPE: internal
5	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:
ю	Asn Lys Asp Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser 1 5 10
	(2) INFORMATION FOR SEQ ID NO:66:
15	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 11 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
20	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:
	Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser 1 5 10
30	(2) INFORMATION FOR SEQ ID NO:67:
	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 11 amino acids
35	(B) TYPE: amino acid (D) TOPOLOGY: linear
	-
	(ii) MOLECULE TYPE: peptide
40	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:
45	Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly 1 5 10
	(2) INFORMATION FOR SEQ ID NO:68:
50	
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 14 amino acids
	(B) TYPE: amino acid
	(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:

Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro
1 5 10

(2) INFORMATION FOR SEQ ID NO:69:

(2) INCOMENTION FOR DEG 1D NO.03.

10 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 amino acids

(B) TYPE: amino acid(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:

Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro Val Leu

1 10 15

25 Thr

(2) INFORMATION FOR SEQ ID NO:70:

30

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

35

- (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:70:

Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro Val Leu

1 10 15

45

Thr Pro Glu Gln

- 50 (2) INFORMATION FOR SEQ ID NO:71:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
- 55 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

Pro

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:71:
5	Gly Val Asp Pro Val Leu Thr Pro Glu Gln Ser Ala Gly Met Ile 1 5 10 15
	Ala Glu Pro Gly 20
19	
•	(2) INFORMATION FOR SEQ ID NO:72:
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 9 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
••	(ii) MOLECULE TYPE: peptide
20	(v) FRAGMENT TYPE: internal
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:
23	Gly Val Asp Pro Val Leu Thr Pro Glu 1 5
30	(2) INFORMATION FOR SEQ ID NO:73:
35	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 12 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
33	(ii) MOLECULE TYPE: peptide
40	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:
45	Gly Val Asp Pro Val Leu Thr Pro Glu Gln Ser Ala 1 5 10
	(2) INFORMATION FOR SEQ ID NO:74:
50	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 16 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
55	(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:

Gly Val Asp Pro Val Leu Thr Pro Glu Gln Ser Ala Gly Met Ile Pro 1 5 10 15

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- (2) INFORMATION FOR SEQ ID NO:75:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - -(ii) MOLECULE TYPE: peptide

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- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:75:

Ser Ala Gly Met Ile Pro Ala Glu Pro Gly Glu Ser Ala Leu Ser Leu

1 10 15

Thr Ser Ser Ala 25 20

- (2) INFORMATION FOR SEQ ID NO:76:
- 30 (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- 35 (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal
- 40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:76:

Glu Pro Gly Glu Ser Ala Leu Ser Leu Thr Ser Ser Ala Gly Val Leu 1 5 10 15

45 Ser Cys

- (2) INFORMATION FOR SEQ ID NO:77:
- 50 (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- 55 (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

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	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:77:
5	Thr Ser Ser Ala Gly Val Leu Ser Cys Gln Pro Gly Ala Pro Cys 1 5 10 15
	(2) INFORMATION FOR SEQ ID NO:78:
10	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
15	(v) FRAGMENT TYPE: internal
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:78:
	Leu Ala Thr Val Ala Phe Asn Met Phe Thr Asp His Val Asp Gln Arg 1 5 10 15
25	Met Pro
	(2) INFORMATION FOR SEQ ID NO:79:
30	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
35	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
40	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:
	Tyr Ile Leu Tyr Phe Thr Leu Ala Leu Val Ala Leu Leu Gln Pro Val 1 5 10
45	Arg Ser Ala Glu Gly Val Gly Glu Ile Leu Pro 20 25
50	(2) INFORMATION FOR SEQ ID NO:80:
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:80:
5	Leu Asn Gln Glu Leu Val Val Asn Ser Asp Lys Thr Ile Asp Gly Arg 1 5 10 15
	Gly Val Lys
10	(2) INFORMATION FOR SEQ ID NO:81:
15	(i) SEQUENCE CHARACTERISTICS: — (A) LENGTH: 21 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
20	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:81:
25	Gly Met Leu Ala Thr Val Ala Phe Asn Met Phe Thr Asp Asn Val Asp 1 10 15
30	Gln Arg Met Pro Arg 20
	(2) INFORMATION FOR SEQ ID NO:82:
35	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
40	(ii) MOLECULE TYPE: peptide
••	(v) FRAGMENT TYPE: internal
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:82:
	Arg Gly Lys Ala Asp Trp Ala Glu Asn Arg Lys Ala Leu Ala Asp Cys 1 10 15
50	Ala Gln Gly Phe Gly Lys Gly Thr Val Gly Gly Lys 20 25
	(2) INFORMATION FOR SEQ ID NO:83:

(i) SEQUENCE CHARACTERISTICS:

(B) TYPE: amino acid(D) TOPOLOGY: linear

(A) LENGTH: 22 amino acids

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	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
5	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:83:
	Ala Glu Asn Arg Lys Ala Leu Ala Asp Cys Ala Gln Gly Phe Gly Lys 1 10 15
0	Gly Thr Val Gly Gly Lys 20
15	(2) INFORMATION FOR SEQ ID NO:84:
20	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 25 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
25	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:84:
30	Ala Glu Asn Arg Lys Ala Leu Ala Asp Cys Ala Gln Gly Phe Gly Lys 1 5 10 15
	Gly Thr Val Gly Gly Lys Asp Gly Asp 20 25
35	(2) INFORMATION FOR SEQ ID NO:85:
40	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
45	(v) FRAGMENT TYPE: internal
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:85:
-	Gly Phe Gly Lys Gly Thr Val Gly Gly Lys Asp Gly Asp Ile Tyr Th 1 5 10
55	Val Thr Ser Glu Leu Asp Asp Asp Val Ala Asn 20 25
	(2) INFORMATION FOR SEQ ID NO:86:

(i) SEQUENCE CHARACTERISTICS:

	**
	(A) LENGTH: 24 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
5	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:86:
15	Lys Gly Thr Val Gly Gly Lys Asp Gly Asp Ile Tyr Thr Val Thr Ser
	Glu Leu Asp Asp Val Ala Asn 20
20	(2) INFORMATION FOR SEQ ID NO:87:
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 amino acids(B) TYPE: amino acid
25	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
30	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:87:
35	Gly Lys Asp Gly Asp Ile Tyr Thr Val Thr Ser Glu Leu Asp Asp Asp 1 5 10 15
	Val Ala Asn
40	(2) INFORMATION FOR SEQ ID NO:88:
45	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 25 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
50	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:88:
55	Glu Asn Asn Arg Gln Ala Leu Ala Asp Cys Ala Gln Gly Phe Ala Lys 1 5 10 15

Gly Thr Tyr Gly Gly Lys Trp Gly Asp

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	(2) INFORMATION FOR SEQ 25 STORY
5	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
10	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
15	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:89:
	Lys Gly Thr Tyr Gly Gly Lys Trp Gly Asp Val Tyr Thr Val Thr Ser
20	Asn Leu Asp Asp Val Ala Asn 20
	(2) INFORMATION FOR SEQ ID NO:90:
25	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 22 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
30	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:90:
	Lys Ser Asn Asp Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp 1 15
40	Ala Ile Ser Ile Ser Gly 20
45	(2) INFORMATION FOR SEQ ID NO:91:
43	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 25 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
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	(v) FRAGMENT TYPE: internal
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	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:91:

Lys Ser Asn Asp Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp

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Ala Ile Ser Ile Ser Gly Ser Ser Gln
20 25

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- (2) INFORMATION FOR SEQ ID NO:92:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 amino acids

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(B) TYPE: amino acid(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

15 (v) FRAGMENT TYPE: internal

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:92:

Lys Ser Asn Asp Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp
1 10 15

Ala Ile Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp His 25 20 25 30

- (2) INFORMATION FOR SEQ ID NO:93:
- (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 35 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

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(v) FRAGMENT TYPE: internal

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:93:

Lys Ser Asn Asp Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp

1 10 15

Ala Ile Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp His Cys Ser
20 25 30

Leu Ser Lys 35

- 50 (2) INFORMATION FOR SEQ ID NO:94:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 amino acids
 - (B) TYPE: amino acid
- 55 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

5	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:94:
	Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile 1 5 10 15
10	Ser Gly
	(2) INFORMATION FOR SEQ ID NO:95:
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 21 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
20	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:95:
	Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile 1 5 10 15
30	Ser Gly Ser Ser Gln 20
	(2) INFORMATION FOR SEQ ID NO:96:
35	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
40	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:96:
50	Lys Lys Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile Se 1 5 10 15
	Gly Ser Ser Gln Ile Trp Ile Asp His Lys Lys 20 25
55	(2) INFORMATION FOR SEQ ID NO:97:
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 31 amino acids

(B) TYPE: amino acid
(D) TOPOLOGY: linear

	(ii) MOLECULE TYPE: peptide	
5	(v) FRAGMENT TYPE: internal	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:97:	
10	Lys Lys Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile 1 5 10 15	Ser
15	Gly Ser Ser Gln Ile Trp Ile Asp His Cys Ser Leu Ser Lys Lys 20 25 30	
	(2) INFORMATION FOR SEQ ID NO:98:	
20	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 31 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
25	(v) FRAGMENT TYPE: internal	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:98:	
	Lys Lys Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile 1 10 15	Ser
35	Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu Ser Lys Lys 20 25 30	
	(2) INFORMATION FOR SEQ ID NO:99:	
40	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 31 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
45	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:99:	
	Lys Lys Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile S 1 5 10 15	Ser
55	Gly Ser Ser Gln Ile Trp Ile Asp His Leu Ser Leu Ser Lys Lys 20 25 30	

(2) INFORMATION FOR SEQ ID NO:100:

	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 31 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
5	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
ю	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:100:
15	Lys Lys Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile Ser 1 10 15
	Gly Ser Ser Gln Ile Trp Ile Asp His Glu Ser Leu Ser Lys Lys 20 25 30
20	(2) INFORMATION FOR SEQ ID NO:101:
25	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
30	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:101:
35	Gly Pro Pro Ile Leu Arg Gln Ala Ser Asp Gly Asp Thr Ile Asn Val
	Ala Gly Ser Ser Gln 20
40	(2) INFORMATION FOR SEQ ID NO:102:
45	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
50	(v) FRAGMENT TYPE: internal
55	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:102:
رر	Arg His Gly Phe Phe Gln Val Val Asn Asn Asn Tyr Asp Lys Trp Gly

Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr

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(2) INFORMATION FOR SEQ ID NO:103:

5 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- 10 (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:103:

Phe Phe Gln Val Val Asn Asn Asn Tyr Asp Lys Trp Gly Ser Tyr Ala

1 10 15

Ile Gly Gly Ser Ala Ser Pro Thr 20

- (2) INFORMATION FOR SEQ ID NO:104:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:104:

Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr Ile Leu Ser Gln
40 1 5 10 .15

Gly Asn Arg Phe Cys Ala Pro Asp 20

- 45 (2) INFORMATION FOR SEQ ID NO:105:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 amino acids(B) TYPE: amino acid
- 50 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:105:

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- 106 -Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr Ile Leu Ser Gln Gly Asn Arg Phe Cys Ala Pro Asp Glu Arg 5 (2) INFORMATION FOR SEQ ID NO:106: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 amino acids -10 (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide 15 (v) FRAGMENT TYPE: internal (xi) SEQUENCE DESCRIPTION: SEQ ID NO:106: 20 Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr Ile Leu Ser Gln Gly Asn Arg Phe Cys Ala Pro Asp Glu Arg Ser Lys 25 (2) INFORMATION FOR SEQ ID NO:107: (i) SEQUENCE CHARACTERISTICS: 30 (A) LENGTH: 24 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide 35 (v) FRAGMENT TYPE: internal 40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:107: Phe Phe Gln Val Val Asn Asn Tyr Asp Arg Trp Gly Thr Tyr Ala 10 45 Ile Gly Gly Ser Ser Ala Pro Thr (2) INFORMATION FOR SEQ ID NO:108: 50 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:108:
5	Gly Thr Tyr Ala Ile Gly Gly Ser Ser Ala Pro Thr Ile Leu Cys Gln 1 5 10 15
10	Gly Asn Arg Phe Leu Ala Pro Asp Asp Gln Ile Lys 20 25
	(2) INFORMATION FOR SEQ ID NO:109:
15	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
20	(v) FRAGMENT TYPE: internal
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25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:109:
	Lys Asp Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp 1 5 10 15
30	Pro Val Leu Thr Pro Glu Gln Ser Ala Gly Met Ile Pro 20 25
	(2) INFORMATION FOR SEQ ID NO:110:
35	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 26 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
40	(ii) MOLECULE TYPE: peptide
40	(v) FRAGMENT TYPE: internal
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:110:
	Lys Asp Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp 1 5 10 15
50	Pro Val Leu Thr Pro Glu Gln Ser Ala Gly 20 25
	(2) INFORMATION FOR SEQ ID NO:111:
55	(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 31 amino acids

(B) TYPE: amino acid(D) TOPOLOGY: linear

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	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
5	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:111:
10	Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro Val 1 5 10 15
	Leu Thr Pro Glu Gln Ser Ala Gly Met Ile Pro Ala Glu Pro Gly 20 25 30
15	(2) INFORMATION FOR SEQ ID NO:112:
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 29 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
20	(ii) MOLECULE TYPE: peptide
25	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:112:
30	Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro Val 1 5 10 15
	Leu Thr Pro Glu Gln Ser Ala Gly Met Ile Pro Ala Glu 20 25
35	(2) INFORMATION FOR SEQ ID NO:113:
40	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
45	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:113:
50	Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro Val 1 10 15
55	Leu Thr Pro Glu Gln Ser Ala Gly Met Ile Pro 20 25
	(2) INFORMATION FOR SEQ ID NO:114:

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 24 amino acids
                 (B) TYPE: amino acid
                 (D) TOPOLOGY: linear
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          (ii) MOLECULE TYPE: peptide
           (v) FRAGMENT TYPE: internal
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          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:114:
           Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro Val
                           5
                                                                     15
 15
           Leu Thr Pro Glu Gln Ser Ala Gly
                       20
      (2) INFORMATION FOR SEQ ID NO:115:
 20
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 21 amino acids
                (B) TYPE: amino acid
                (D) TOPOLOGY: linear
25
          (ii) MOLECULE TYPE: peptide
           (v) FRAGMENT TYPE: internal
30
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:115:
          Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro Val
35
          1
          Leu Thr Pro Glu Gln
                      20
     (2) INFORMATION FOR SEQ ID NO:116:
40
          (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 27 amino acids
               (B) TYPE: amino acid
45
               (D) TOPOLOGY: linear
         (ii) MOLECULE TYPE: peptide
          (v) FRAGMENT TYPE: internal
50
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:116:
55
         Leu Leu Glu Asn Gly Ala Ile Phe Val Thr Ser Gly Ser Asp Pro Val
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Leu Thr Pro Val Gln Ser Ala Gly Met Ile Pro

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	(2) INFORMATION FOR SEQ ID NO:117:
5	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 60 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
ю.	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
<u></u> .	- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:117:
15	
of the control of the	Ala Tyr Asn Ile Ile Asp Gly Cys Trp Arg Gly Lys Ala Asp Trp Ala 1 5 10 15
20	Glu Asn Arg Lys Ala Leu Ala Asp Cys Ala Gln Gly Phe Gly Lys Gly 20 25 30
	Thr Val Gly Gly Lys Asp Gly Asp Ile Tyr Thr Val Thr Ser Glu Leu 35 40 45
25	Asp Asp Asp Val Ala Asn Pro Lys Glu Gly Thr Leu 50 55 60
30	(2) INFORMATION FOR SEQ ID NO:118:
35	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 46 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
40	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:118:
45	His Asp Val Lys Val Asn Pro Gly Gly Leu Ile Lys Ser Asn Asp Gl 1 5
	Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp Ala Ile Ser Ile Se 20 25 / 30
50	Gly Ser Ser Gln Ile Trp Ile Asp His Cys Ser Leu Ser Lys 35 40 45
55	(2) INFORMATION FOR SEQ ID NO:119:

(i) SEQUENCE CHARACTERISTICS:

(B) TYPE: amino acid

(A) LENGTH: 45 amino acids

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- 5 (v) FRAGMENT TYPE: internal
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:119:

'Arg Cys Arg His Gly Phe Phe Gln Val Val Asn Asn Asn Tyr Asp Lys

Trp Gly Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr Ile Leu Ser
15 20 25 30

Gln Gly Asn Arg Phe Cys Ala Pro Asp Glu Arg Ser Lys 35 40 45

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- (2) INFORMATION FOR SEQ ID NO:120:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 47 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
- 30 (v) FRAGMENT TYPE: internal
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:120:
- 35 Glu Ala Ala Glu Ser Met Lys Trp Asn Trp Arg Thr Asn Lys Asp 1 5 10 15

Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp Pro Val 20 25 30

Leu Thr Pro Glu Gln Ser Ala Gly Met Ile Pro Ala Glu Pro Gly
35 40 45

- 45 (2) INFORMATION FOR SEQ ID NO:121:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 amino acids
 - (B) TYPE: amino acid
- 50 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:121:

Arg Gly Lys Ala Asp Trp Ala Glu Asn Arg Lys Ala Leu Ala Asp Cys

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Ala Gln Gly Phe Gly Lys Gly Thr Val Gly Gly Lys Asp Gly Asp Ile 20 25 30

Tyr Thr Val Thr Ser Glu Leu Asp Asp Asp Val Ala Asn

10 (2) INFORMATION FOR SEQ ID NO:122:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 35 amino acids
 - (B) TYPE: amino acid
- 15 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

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- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:122:
- Lys Ser Asn Asp Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly Asp

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Ala Ile Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp His Cys Ser

30 Leu Ser Lys

(2) INFORMATION FOR SEQ ID NO:123:

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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 43 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

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- (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:123:

Arg His Gly Phe Phe Gln Val Val Asn Asn Tyr Asp Lys Trp Gly

Ser Tyr Ala Ile Gly Gly Ser Ala Ser Pro Thr Ile Leu Ser Gln Gly
20 25 30

Asn Arg Phe Cys Ala Pro Asp Glu Arg Ser Lys 35 40

(2) INFORMATION FOR SEQ ID NO:124:

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	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 36 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
5	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:124:
15	Arg Thr Asn Lys Asp Val Leu Glu Asn Gly Ala Ile Phe Val Ala Ser 1 10 15
	Gly Val Asp Pro Val Leu Thr Pro Glu Gln Ser Ala Gly Met Ile Pro 20 25 30
20	Ala Glu Pro Gly 35
	(2) INFORMATION FOR SEQ ID NO:125:
25	(i) SEQUENCE CHARACTERS:(A) LENGHT 24 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
30	(ii) MOLECULAR TYPE: peptide
	(v) FRAGMENT TYPE: internal
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:125:
40	Ser Asp Gly Asp Ala Ile Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile 1 10 15
	Asp His Cys Ser Leu Ser Lys Ser 20
45	(2) INFORMATION FOR SEQ ID NO:126:
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 16 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
50	(ii) MOLECULE TYPE: peptide
	(v) FRAGMENT TYPE: internal
55	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:126:

Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu Ser Lys Ser

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15 10 1 (2) INFORMATION FOR SEQ ID NO:127: (i) SEQUENCE CHARACTERISTICS: 5 (A) LENGTH: 18 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide 10 (v) FRAGMENT TYPE: internal 15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:127: Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu Ser 10 20 Lys Ser (2) INFORMATION FOR SEQ ID NO:128: 25 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 21 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear 30 (ii) MOLECULE TYPE: peptide (v) FRAGMENT TYPE: internal 35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:128: Asp Ala Ile Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser 10 40 Ser Leu Ser Lys Ser 20 (2) INFORMATION FOR SEQ ID NO:129: 45 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear 50 (ii) MOLECULE TYPE: peptide (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:129:

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Ser Asp Gly Asp Ala Ile Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile

1 10 15

Asp His Ser Ser Leu Ser Lys Ser 5

- (2) INFORMATION FOR SEQ ID NO:130:
 - (i) SEQUENCE CHARACTERISTICS:

10 (A) LENGTH: 18 amino acids

- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- _(ii) MOLECULE TYPE: peptide
- (v) FRAGMENT TYPE: internal
- 20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:130:

Ser Glu Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu Ser 1 10 15

- 25 Lys Ser
 - (2) INFORMATION FOR SEQ ID NO:131:
- 30 (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- 35 (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:131:

Ser Lys Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu Ser 1 10 15

Lys Ser

- 50 (2) INFORMATION FOR SEQ ID NO:132:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 amino acids
 - (B) TYPE: amino acid
- 55 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:132:	
5	Asp Lys Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu S 1 10 15	er:
10	Lys Glu	
	(2) INFORMATION FOR SEQ ID NO:133:	
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
20	(v) FRAGMENT TYPE: internal	
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:133:	
	Asp Lys Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser S 1 10 15	ei
30	Leu Ser Lys Glu 20	
	(2) INFORMATION FOR SEQ ID NO:134:	
35	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
40	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:134:	
50	Asp Lys Glu Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser 1 5 10 15	Leu
50	Ser Lys Glu	
. 55	(2) INFORMATION FOR SEQ ID NO:135:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 amino acids (B) TYPE: amino acid	

(D) TOPOLOGY: linear

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	(ii) MOLECULE TYPE: peptide
5	(v) FRAGMENT TYPE: internal
•	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:135:
10	Asp Lys Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu Se
	Lys Lys
15	(2) INFORMATION FOR SEQ ID NO:136:
20	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
25	(v) FRAGMENT TYPE: internal
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:136:
30	Asp Lys Glu Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu 1 5 10 15
35	Ser Lys Glu Lys 20
	(2) INFORMATION FOR SEQ ID NO:137:
40	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 16 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
45	(v) FRAGMENT TYPE: internal
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:137:
٠	Ser Gly Ser Ser Gln Ile Ala Ile Asp His Ser Ser Leu Ser Lys Ser
55	 -

(2) INFORMATION FOR SEQ ID NO:138:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids

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	(B) TYPE: amino acid (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: peptio	le
(v)	FRAGMENT TYPE: intern	al

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:138:

Asp Lys Glu Ser Gly Ser Ser Gln Ile Ala Ile Asp His Ser Ser Leu
1 5 10 15

15 Ser Lys Ser Glu 20

(2) INFORMATION FOR SEQ ID NO:139:

20 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

30
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:139:
Asp Lys Glu Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Ser Leu

35 Ser Lys Glu Lys Asp 20 -

(2) INFORMATION FOR SEQ ID NO:140:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 13 amino acids(B) TYPE: amino acid

(D) TOPOLOGY: linear

45 (ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:140:

Ser Gln Ile Trp Ile Asp His Ser Ser Leu Ser Lys Ser

(2) INFORMATION FOR SEQ ID NO:141:

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 23 amino acids(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:141:

Asp Lys Glu Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser

Ser Leu Ser Lys Glu Lys Asp 20

(2) INFORMATION FOR SEQ ID NO:142:

20

15

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

25

- (ii) MOLECULE TYPE: peptide
- (v) FRAGMENT TYPE: internal

30

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:142:
- Lys Asp Val Leu His Asn Gly Ala Ile Phe Val Ala Ser Gly Val Glu
 1 5 10 15

Pro Val Leu Thr Pro His Gln Ser Ala Gly
20 __ 25

(2) INFORMATION FOR SEQ ID NO:143:

40

35

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 amino acids
 - (B) TYPE: amino acid

45

- (D) TOPOLOGY: linear
- (v) FRAGMENT TYPE: internal

(ii) MOLECULE TYPE: peptide

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:143:

Lys Asp Val Leu His Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asn
1 5 10 15

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Pro Val Leu Thr Pro His Gln Ser Ala Gly
20 25

(2) INFORMATION FOR SEQ ID NO:144:

5	(i) SEQUENCE CHARACTERISTICS:(A) LENGHT: 26 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
•	(ii) MOLECULAR TYPE: peptide
10	(v) FRAGMENT TYPE: internal
	(xi) SEQUENCE DESCRIPTION SEQ ID NO:144:
15	Lys Asp Val Leu His Asn Gly Ala Ile Phe Val Ala Ser Gly Val Ala 1 5 10 15
20	Pro Val Leu Thr Pro His Gln Ser Ala Gly 20 25
20	(2) INFORMATION FOR SEQ ID NO:145:
25	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 26 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
30	(v) FRAGMENT TYPE: internal
	i i
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:145:
33	Lys Asp Val Leu His Asn Gly Ala Ile Phe Val Ala Ser Gly Val Ser 1 5- 10 15
40	Pro Val Leu Thr Pro His Gln Ser Ala Gly 20 25
	(2) INFORMATION FOR SEQ ID NO:146:
45	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 26 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
50	(v) FRAGMENT TYPE: internal
55	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:146:
	Lys Asp Val Leu His Asn Gly Ala Ile Phe Val Ala Ser Gly Val Gl

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Pro Val Leu Thr Pro His Gln Ser Ala Gly 20 25

(2) INFORMATION FOR SEQ ID NO:147:

5

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

10

- (ii) MOLECULE TYPE: peptide
- (v) FRAGMENT TYPE: internal

15

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:147:
- Lys Asp Val Leu His Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp
 1 5 10 15

Ala Val Leu Thr Pro His Gln Ser Ala Gly
20 25

- 25 (2) INFORMATION FOR SEQ ID NO:148:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 amino acids
 - (B) TYPE: amino acid
- 30 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (v) FRAGMENT TYPE: internal

35

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:148:
- Lys Asp Val Leu His Asn Gly Ala Ile Phe Val Ala Ser Gly Val Asp

 1 5 10 15

Ser Val Leu Thr Pro His Gln Ser Ala Gly

45

- (2) INFORMATION FOR SEQ ID NO:149:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 amino acids
 - (B) TYPE: amino acid(D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
- 55 (v) FRAGMENT TYPE: internal
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:149:

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	Lys Asp Val Leu His Asn Gly Ala Ile Phe Val Ala Ser Gly Val As 1 5 10 15	р
5	Gly Val Leu Thr Pro His Gln Ser Ala Gly 20 25	
	(2) INFORMATION FOR SEQ ID NO:150:	
10	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 35 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
15	(ii) MOLECULE TYPE: peptide	
	(v) FRAGMENT TYPE: internal	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:150:	
25	Lys Ser Asn Asp Gly Pro Ala Ala Pro Arg Ala Gly Ser Asp Gly As 1 5 10 15	p
25	Ala Ile Ser Ile Ser Gly Ser Ser Gln Ile Trp Ile Asp His Ser Se 20 25 30	er
30	Leu Ser Lys 35	

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Claims

1. All or a portion of an isolated peptide of Amb a I.1, said peptide or portion thereof comprising at least one T cell epitope of Amb a I.1, said peptide comprising an amino acid sequence selected from the group consisting of:

```
AMB 1-1.1 (SEQ ID NO:85);
                    a)
                    b)
                          AMB 1-2.1 (SEQ ID NO:86);
                    c)
                          AMB 1-3.1 (SEQ ID NO:87);
                    d)
                          AMB 1-4.1 (SEQ ID NO:84);
 10
                    e)
                          AMB 1-5.1 (SEQ ID NO:83);
                    f)
                          AMB 1-6.1 (SEQ ID NO:82);
                          AMB 2-4.1 (SEQ ID NO:90);
                    g)
                   h)
                          AMB 2-3.1 (SEQ ID NO:91);
                   i)
                          AMB 2-5.1 (SEQ ID NO:92);
 15
                   j)
                          AMB 2-6.1 (SEQ ID NO:93);
                   k)
                          AMB 2-2.1 (SEQ ID NO:94);
                   1)
                          AMB 2-1.1 (SEQ ID NO:95);
                   m)
                          RAE 70.1-1 (SEQ ID NO:43);
                   n)
                          AMB 2-7.1 (SEQ ID NO:96);
20
                   0)
                          AMB 2-8.1 (SEQ ID NO:97);
                          AMB 2-9.1 (SEQ ID NO:98);
                   p)
                          AMB 2-10.1 (SEQ ID NO:99);
                   q)
                          AMB 2-11.1 (SEQ ID NO:100);
                   r)
                   s)
                          RAE 70.1 (SEQ ID NO:44):
25
                          AMB 3-4.1 (SEQ ID NO:103);
                   t)
                   u)
                          AMB 3-5.1 (SEQ ID NO:102);
                          AMB 3-3.1 (SEQ ID NO:104);
                   v)
                   w)
                         AMB 3-2.1 (SEQ ID NO:105);
                   x)
                         AMB 3-1.1 (SEQ ID NO:106);
30
                         AMB 4-8.1 (SEQ ID NO:109);
                  y)
                   z)
                         AMB 4-9.1 (SEQ ID NO:110);
                  a')
                         AMB 4-6.1 (SEQ ID NO:111);
                  b')
                         AMB 4-5.1 (SEQ ID NO:112);
                  c')
                         AMB 4-3.1 (SEQ ID NO:113);
35
                  d')
                         AMB 4-2.1 (SEQ ID NO:114);
                  e')
                         AMB 4-1.1 (SEQ ID NO:115);
                  f)
                         AMB 2-38.1 (SEQ ID NO:141);
                         RA-02.1 (SEQ ID NO:150); and
                  g')
```

h')

AMB 2-36.1 (SEQ ID NO:139).

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- 2. All or a portion of an isolated peptide of claim 1 which has a stimulation index of at least 2.0.
- All or a portion of an isolated peptide of claim 1 which has minimal immunoglobulin E stimulating activity.
 - 4. All or a portion of an isolated peptide of claim 1 which does not bind immunoglobulin E specific for Amb a I.1 in a substantial percentage of individuals sensitive to Amb a I.1, or if binding of the peptide to said immunoglobulin E occurs, such binding does not result in release of mediators from mast cells or basophils in a substantial percentage of individuals sensitive to Amb a 1.1.
- 5. All or a portion of an isolated peptide of claim 1 which binds immunoglobulin E to a substantially lesser extent than Amb a I.1 binds said immunoglobulin E.
 - 6. All or a portion of an isolated peptide of claim 1 which modifies, in a ragweed pollen-sensitive individual to whom it is administered, the allergic response of the individual to ragweed pollen.
 - 7. A portion of an isolated peptide of claim 1 wherein the portion comprises at least fifteen amino acid residues.
 - 8. An isolated peptide of Amb a 1.1 comprising an amino acid sequence selected from the group consisting of:
 - a) AMB 1-1.1 (SEQ ID NO:85);
 - b) AMB 1-2.1 (SEQ ID NO:86);
 - c) AMB 1-3.1 (SEQ ID NO:87);
 - d) AMB 1-4.1 (SEQ ID NO:84);
 - e) AMB 1-5.1 (SEQ ID NO:83);
 - f) AMB 1-6.1 (SEQ ID NO:82);
 - g) AMB 2-4.1 (SEQ ID NO:90);
 - h) AMB 2-3.1 (SEQ ID NO:91);
 - i) AMB 2-5.1 (SEQ ID NO:92);
 - j) AMB 2-6.1 (SEQ ID NO:93);
 - k) AMB 2-2.1 (SEQ ID NO:94);
 - l) AMB 2-1.1 (SEQ ID NO:95);
 - m) RAE 70.1-1 (SEQ ID NO:43);
 - n) AMB 2-7.1 (SEQ ID NO:96);

- AMB 2-8.1 (SEQ ID NO:97); 0) p) AMB 2-9.1 (SEQ ID NO:98); AMB 2-10.1 (SEQ ID NO:99); q) r) AMB 2-11.1 (SEQ ID NO:100); 5 s) RAE 70.1 (SEQ ID NO:44); t) AMB 3-4.1 (SEQ ID NO:103); u) AMB 3-5.1 (SEQ ID NO:102); v) AMB 3-3.1 (SEQ ID NO:104); w) AMB 3-2.1 (SEQ ID NO:105); 10 AMB 3-1.1 (SEQ ID NO:106); x) y) AMB 4-8.1 (SEQ ID NO:109); AMB 4-9.1 (SEQ ID NO:110); Z) a') AMB 4-6.1 (SEQ ID NO:111); b') AMB 4-5.1 (SEQ ID NO:112); 15 c') AMB 4-3.1 (SEQ ID NO:113); d') AMB 4-2.1 (SEQ ID NO:114); e') AMB 4-1.1 (SEQ ID NO:115); f) AMB 2-38.1 (SEQ ID NO:141); g') RA-02.1 (SEQ ID NO:150); and 20 h') AMB 2-36.1 (SEQ ID NO:139).
 - 9. All or a portion of an isolated peptide of claim 1, wherein said peptide or a portion thereof comprises at least two T cell epitopes Amb a 1.1.
- 25 10. An isolated nucleic acid having a sequence encoding all or a portion of a peptide of claim 1 or the functional equivalent of said nucleic acid sequence.
 - 11. An isolated peptide which is immunologically cross-reactive with antibodies specific for all or a portion of a peptide of claim 1.
 - 12. An isolated peptide which is immunologically cross-reactive with T cells reactive with all or a portion of a peptide of claim 1.
 - 13. All or a portion of a modified peptide of claim 1.
 - 14. A modified peptide of claim 8.
 - 15. All or a portion of a modified peptide of claim 13 which has minimal immunoglobulin E stimulating activity.

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- 16. All or a portion of a modified peptide of claim 13 which does not bind immunoglobulin E specific for Amb a I.1 in a substantial percentage of individuals sensitive to Amb a I.1, or if binding of the peptide to said immunoglobulin E occurs, such binding does not result in release of mediators from mast cells or basophils in a substantial percentage of individuals sensitive to Amb a I.1.
- 17. All or a portion of a modified peptide of claim 13 which binds immunoglobulin E to a substantially lesser extent than Amb a I.1 binds said immunoglobulin E.
- 18. All or a portion of a modified peptide of claim 13 which modifies, in a ragweed pollen-sensitive individual to whom it is administered, the allergic response of the individual to Amb a I.1.
- 19. All or a portion of an isolated peptide of Ambrosia artemisiifolia, said peptide or portion thereof comprising at least one T cell epitope of Ambrosia artemisiifolia, said peptide comprising an amino acid sequence selected from the group consisting of:
 - a) RAE 67.1 (SEQ ID NO:13);
- 20 b) RAE 57.1 (SEQ ID NO:14);
 - c) RAE 24.E (SEQ ID NO:15);
 - d) RAE 24.1 (SEQ ID NO:16);
 - e) RAE 22.E (SEQ ID NO:17);
 - f) RAE 22.E-1 (SEQ ID NO:18);
- 25 g) RAE 3.1 (SEQ ID NO:20);
 - h) RAE 22.E-2 (SEQ ID NO:21);
 - i) RAE 5.D (SEQ ID NO:22);
 - j) RAE 6.D (SEQ ID NO:23);
 - k) RAE 6.1 (SEQ ID NO:24);
 - l) RAE 7.D (SEQ ID NO:25);
 - m) RAE 7.D-1 (SEQ ID NO:26);
 - n) RAE 40.1-6 (SEQ ID NO:27);
 - o) RAE 40.1-5 (SEQ ID NO:28);
 - p) RAE 40.1-4 (SEQ ID NO:29);
 - q) RAE 40.D (SEQ ID NO:30);
 - r) RAE 40.1 (SEQ ID NO:31);
 - s) RAE 61.1 (SEQ ID NO:32);
 - t) RAE 80.1 (SEQ ID NO:33);
 - u) RAE 45.1 (SEQ ID NO:34);

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v)
                    RAE 75.1 (SEQ ID NO:35);
             w)
                    RAE 62.1 (SEQ ID NO:36);
                    RAE 69.1 (SEQ ID NO:37);
             x)
             y)
                    RAE 69.1-1 (SEQ ID NO:38);
  5
             z)
                   RAE 69.1-2 (SEQ ID NO:39);
             a')
                   RAE 69.1-3 (SEQ ID NO:40);
             b')
                   RAE 70.1-3 (SEQ ID NO:41);
            c')
                   RAE 70.1-2 (SEQ ID NO:42);
            d')
                   RAE 71.1 (SEQ ID NO:45);
 10 -
            e')
                   RAE 65.1 (SEQ ID NO:46);
            f)
                   RAE 63.1 (SEQ ID NO:47);
            g')
                   RAE 76.1 (SEQ ID NO:48);
            h')
                  RAE 27.1 (SEQ ID NO:49);
            i')
                   RAE 66.1 (SEQ ID NO:50);
15
            j')
                   RAE 66.1-1 (SEQ ID NO:51);
            k')
                   RAE 66.1-2 (SEQ ID NO:52);
            ľ)
                   RAE 66.1-3 (SEQ ID NO:53);
                   RAE 64.1-3 (SEQ ID NO:54);
            m')
                   RAE 64.1-2 (SEQ ID NO:55);
            n')
20
            o')
                   RAE 64.1-1 (SEQ ID NO:56);
            p')
                   RAE 64.1 (SEQ ID NO:57);
                   RAE 73.1 (SEQ ID NO:58);
            q')
            r')
                   RAE 74.1 (SEQ ID NO:59);
            s')
                   RAE 74.1-1 (SEQ ID NO:60);
25
            t')
                  RAE 29.1 (SEQ ID NO:61);
            u')
                  RAE 29.1-1 (SEQ ID NO:62);
            v')
                  RAE 28+29 (SEQ ID NO:63);
            w')
                  RAE 29.1-2 (SEQ ID NO:64);
            x')
                  RAE 29.1-3 (SEQ ID NO:65);
30
            y')
                  RAE 29.1-4 (SEQ ID NO:66);
            z')
                  RAE 28.1-3 (SEQ ID NO:67);
                  RAE 28.1-2 (SEQ ID NO:68);
            a")
           b")
                  RAE 28.1-1 (SEQ ID NO:69);
           c")
                  RAE 28.1 (SEQ ID NO:70);
           d")
                  RAE 20.1 (SEQ ID NO:71);
35
           e")
                  RAE 20.1-3 (SEQ ID NO:72);
           f'')
                  RAE 20.1-2 (SEQ ID NO:73);
           g")
                  RAE 20.1-1 (SEQ ID NO:74);
           h")
                  RAE 21.1 (SEQ ID NO:75);
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i") RAE 17.1 (SEQ ID NO:76);
j") RAE 55.1 (SEQ ID NO:77);
k") RAE 76.6 (SEQ ID NO:78);
l") RAE 67.15 (SEQ ID NO:79);
m") RAE 45.15 (SEQ ID NO:80); and
n") RAE 27.15 (SEQ ID NO:81).
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20. An isolated peptide comprising at least two regions, each region comprising at least one T cell epitope of a protein allergen of Ambrosia artemisiifolia, said regions derived from the same or from different protein allergens of Ambrosia artemisiifolia, said regions each comprising all or a portion of an amino acid sequence selected from the group consisting of:

AMB 1-1.1 (SEQ ID NO:85); · a) AMB 1-2.1 (SEQ ID NO:86); b) 15 AMB 1-3.1 (SEQ ID NO:87); c) AMB 1-4.1 (SEQ ID NO:84); d) AMB 1-5.1 (SEQ ID NO:83); e) AMB 1-6.1 (SEQ ID NO:82); f) AMB 2-4.1 (SEQ ID NO:90); g) 20 AMB 2-3.1 (SEQ ID NO:91); h) AMB 2-5.1 (SEQ ID NO:92); i) AMB 2-6.1 (SEQ ID NO:93); j) AMB 2-2.1 (SEQ ID NO:94); k) AMB 2-1.1 (SEQ ID NO:95); 1) RAE 70.1-1 (SEQ ID NO:43): m) AMB 2-7.1 (SEQ ID NO:96); n) AMB 2-8.1 (SEQ ID NO:97); o) AMB 2-9.1 (SEQ ID NO:98); p) AMB 2-10.1 (SEQ ID NO:99); q) 30 AMB 2-11.1 (SEQ ID NO:100); r) RAE 70.1 (SEQ ID NO:44); s) AMB 3-4.1 (SEQ ID NO:103); t) AMB 3-5.1 (SEQ ID NO:102); u) AMB 3-3.1 (SEQ ID NO:104); v) 35 AMB 3-2.1 (SEQ ID NO:105); w) AMB 3-1.1 (SEQ ID NO:106); x) AMB 4-8.1 (SEQ ID NO:109); y)

z)

AMB 4-9.1 (SEQ ID NO:110);

	a')	AMB 4-6.1 (SEQ ID NO:111);
	b')	AMB 4-5.1 (SEQ ID NO:112);
	c')	AMB 4-3.1 (SEQ ID NO:113);
	d')	AMB 4-2.1 (SEQ ID NO:114);
5	e')	AMB 4-1.1 (SEQ ID NO:115);
	f)	AMB 2-38.1 (SEQ ID NO:141);
	. g')	RA-02.1 (SEQ ID NO:150); and
	h')	AMB 2-36.1 (SEQ ID NO:139).
10	– 21. An i	solated peptide of claim 20 wherein said peptide comprises various
	combinations of reg	gions, said combinations of regions selected from the group consisting of
	a)	AMB 4-6.1 and RAE 70.1 (SEQ ID NO:111 and SEQ ID NO:44);
	b)	AMB 4-6.1 and AMB 2-5.1 (SEQ ID NO:111 and SEQ ID NO:92);
15	c)	AMB 4-9.1 and AMB 2-5.1 (SEQ ID NO:110 and SEQ ID NO:92);
	d)	AMB 4-9.1 and RAE 70.1 (SEQ. ID NO:110 and SEQ ID NO:44);
	e)	AMB 4-6.1, AMB 2-5.1 and AMB 1-2.1 (SEQ ID NO:111, SEQ ID
		NO:92 and SEQ ID NO:86);
	f)	AMB 4-9.1, AMB 2-5.1 and AMB 1-2.1 (SEQ ID NO:110, SEQ ID
20		NO:92 and SEQ ID NO:86);
	g)	AMB 4-6.1, RAE 70.1 and AMB 1-2.1 (SEQ ID NO:111. SEQ ID
		NO:44 and SEQ ID NO:86);
	h)	AMB 4-9.1, RAE 70.1 and AMB 1-2.1 (SEQ ID NO:110, SEQ ID
2.5	•	NO:44 and SEQ ID NO:86);
25	i)	AMB 4-6.1, RAE 70.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:111,
	• ` `	SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:102);
	j)	AMB 4-9.1, RAE 70.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:110,
	1.5	SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:102);
20	k)	AMB 4-6.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID
30		NO:111, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:102);
	1)	AMB 4-9.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID
	-,	NO:110, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID
		NO:102);
35	m)	AMB 4-6.1, RAE 70.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:111,
	•	SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:103);
	n)	AMR 4-9.1 RAF 70.1 AMR 1-2.1 and AMR 2.4.1 (SEO ID NO.110

SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:103);

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		- 130 -
	o)	AMB 4-6.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID
	•	NO:111, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID
		NO:103);
	p)	AMB 4-9.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID
5	-	NO:110, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID
		NO:103);
•	q')	AMB 2-1.15 and AMB 4-3.15 (SEQ ID NO:101, and SEQ ID
		NO:116);
	r')	AMB 1-2.15, AMB 2-1.15 and
10	-	AMB 4-3.15 (SEQ ID NO:89, SEQ ID NO:101, and SEQ ID NO:116);
		and
(Area)	s')	AMB 1-2.15, AMB 2-1.15,
ŧ		AMB 4-3.15 and AMB 3-4.15 (SEQ ID NO:89, SEQ ID NO:101, SEQ
	•	ID NO:116 and SEQ ID NO:107).
15	*	
	22. An iso	lated peptide of claim 20, wherein the regions each comprise an amino
	acid sequence selecte	d from the group consisting of:
	a)	AMB 1-1.1 (SEQ ID NO:85);
20	b)	AMB 1-2.1 (SEQ ID NO:86);
	c)	AMB 1-3.1 (SEQ ID NO:87);
	d)	AMB 1-4.1 (SEQ ID NO:84);
	e)	AMB 1-5.1 (SEQ ID NO:83);
	f)	AMB 1-6.1 (SEQ ID NO:82);
25	g)	AMB 2-4.1 (SEQ ID NO:90);
27.	h)	AMB 2-3.1 (SEQ ID NO:91);
	i)	AMB 2-5.1 (SEQ ID NO:92);
	j)	AMB 2-6.1 (SEQ ID NO:93);
	k)	AMB 2-2.1 (SEQ ID NO:94);
30	l)	AMB 2-1.1 (SEQ ID NO:95);
	m)	RAE 70.1-1 (SEQ ID NO:43);
	n)	AMB 2-7.1 (SEQ ID NO:96);
	0)	AMB 2-8.1 (SEQ ID NO:97);
	p)	AMB 2-9.1 (SEQ ID NO:98);
35	q)	AMB 2-10.1 (SEQ ID NO:99);
	r)	AMB 2-11.1 (SEQ ID NO:100);
	s)	RAE 70.1 (SEQ ID NO:44);
	t)	AMB 3-4.1 (SEQ ID NO:103);
	u)	AMB 3-5.1 (SEQ ID NO:102);

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- v) AMB 3-3.1 (SEQ ID NO:104);
- w) AMB 3-2.1 (SEQ ID NO:105);
- x) AMB 3-1.1 (SEQ ID NO:106);
- y) AMB 4-8.1 (SEQ ID NO:109);
- z) AMB 4-9.1 (SEQ ID NO:110);
- a') AMB 4-6.1 (SEQ ID NO:111);
- b') AMB 4-5.1 (SEQ ID NO:112);
- c') AMB 4-3.1 (SEQ ID NO:113);
- d') AMB 4-2.1 (SEQ ID NO:114); and
- e') AMB 4-1.1 (SEQ ID NO:115).
 - 23. An isolated nucleic acid having a sequence encoding all or a portion of a peptide of claim 20, or the functional equivalent of said nucleic acid sequence.
- 15 24. An isolated peptide produced in a host cell transformed with the nucleic acid of claim 23.
- 25. All or a portion of an isolated peptide of Amb a I.3, said peptide or portion thereof comprising at least one T cell epitope of Amb a I.3, said peptide comprising an amino acid sequence selected from the group consisting of:
 - a) AMB 1-4.15 (SEQ ID NO:88);
 - b) AMB 1-2.15 (SEQ ID NO:89);
 - c) AMB 2-1.15 (SEQ ID NO:101);
 - d) AMB 3-4.15 (SEQ ID NO:107);
 - e) AMB 3-1.15 (SEQ ID NO:108); and
 - f) AMB 4-3.15 (SEQ ID NO:116).
- 26. A therapeutic composition comprising all or a portion of an isolated peptide of claim 1 and a pharmaceutically acceptable carrier or diluent.
 - 27. A method of treating sensitivity to ragweed pollen in an individual, comprising administering to the individual a therapeutically effective amount of the composition of claim 26.
 - 28. A method of treating sensitivity to ragweed pollen in an individual, comprising administering sequentially to the individual a therapeutically effective amount of two different compositions of claim 26.

- 29. A therapeutic composition comprising all or a portion of an isolated peptide of claim 8 and a pharmaceutically acceptable carrier or diluent.
- 30. A therapeutic composition comprising all or a portion of an isolated peptide of claim 19 and a pharmaceutically acceptable carrier or diluent.
 - 31. A therapeutic composition comprising all or a portion of an isolated peptide of claim 20 and a pharmaceutically acceptable carrier or diluent.
- 32. A method of treating sensitivity to ragweed pollen in an individual, comprising administering to the individual a therapeutically effective amount of the composition of claim 31.
- 33. A method of detecting sensitivity to ragweed pollen in an individual, comprising combining a blood sample obtained from the individual with all or a portion of peptide of claim 1, under conditions appropriate for binding of blood components with the peptide, and determining the extent to which such binding occurs as indicative of sensitivity in the individual to ragweed pollen.
- 20 34. A method of claim 33 wherein the extent to which binding occurs is determined by assessing T cell function, T cell proliferation or a combination thereof.
 - 25. A method of detecting sensitivity to ragweed pollen in an individual, comprising combining a blood sample obtained from the individual with a peptide of claim 20, under conditions appropriate for binding of blood components with the peptide, and determining the extent to which such binding occurs as indicative of sensitivity in the individual to ragweed pollen.
- 36. A method of claim 35 wherein the extent to which binding occurs is determined by assessing T cell function, T cell proliferation or a combination thereof.
 - 37. A composition comprising at least two peptides, said peptides each comprising at least one T cell epitope of a protein allergen of Ambrosia artemisiifolia, said peptides derived from the same or from different protein allergens of Ambrosia artemisiifolia, said composition selected from the group consisting of:
 - a) AMB 4-6.1 and RAE 70.1 (SEQ ID NO:111 and SEQ ID NO:44);
 - b) AMB 4-6.1 and AMB 2-5.1 (SEQ ID NO:111 and SEQ ID NO:92);
 - c) AMB 4-9.1 and AMB 2-5.1 (SEQ ID NO:110 and SEQ ID NO:92);

AMB 4-9.1 and RAE 70.1 (SEQ ID NO:110 and SEQ ID NO:44); d) AMB 4-6.1, AMB 2-5.1 and AMB 1-2.1 (SEQ ID NO:111, SEQ ID e) NO:92 and SEQ ID NO:86); AMB 4-9.1, AMB 2-5.1 and AMB 1-2.1 (SEQ ID NO:110, SEQ ID f) 5 NO:92 and SEQ ID NO:86); AMB 4-6.1, RAE 70.1 and AMB 1-2.1 (SEQ ID NO:111, SEO ID g) NO:44 and SEQ ID NO:86); h) AMB 4-9.1, RAE 70.1 and AMB 1-2.1 (SEQ ID NO:110, SEQ ID NO:44 and SEQ ID NO:86); 10 i) AMB 4-6.1, RAE 70.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:111, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:102); j) AMB 4-9.1, RAE 70.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:110, SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:102); k) AMB 4-6.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID 15 NO:111, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:102); I) AMB 4-9.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-5.1 (SEQ ID NO:110, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:102); AMB 4-6.1, RAE 70.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:111, 20 m) SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:103): AMB 4-9.1, RAE 70.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID NO:110. n) SEQ ID NO:44, SEQ ID NO:86 and SEQ ID NO:103): o) AMB 4-6.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-4.1 (SEQ ID 25 NO:111, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:103); AMB 4-9.1, AMB 2-5.1, AMB 1-2.1 and AMB 3-4:1 (SEQ ID p) NO:110, SEQ ID NO:92, SEQ ID NO:86 and SEQ ID NO:103); AMB 2-1.15 and AMB 4-3.15 (SEQ ID NO:101, and SEQ ID 30 **q'**) NO:116); AMB 1-2.15, AMB 2-1.15 and/ r') AMB 4-3.15 (SEQ ID NO:89, SEQ ID NO:101, and SEQ ID NO:116); and AMB 1-2.15, AMB 2-1.15, 35 s') AMB 4-3.15 and AMB 3-4.15 (SEQ ID NO:89, SEQ ID NO:101, SEQ ID NO:116 and SEQ ID NO:107).

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38. A therapeutic composition comprising a pharmaceutically acceptable carrier or diluent and at least two peptides, said peptides each comprising at least one T cell epitope of a protein allergen of Ambrosia artemisiifolia, said peptides derived from the same or from different protein allergens of Ambrosia artemisiifolia.

39. A method of treating sensitivity to ragweed pollen in an individual, comprising administering to the individual a therapeutically effective amount of the composition of claim 38.

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- 40. A method of treating sensitivity to ragweed pollen in an individual, comprising administering to the individual a therapeutically effective amount of a therapeutic composition comprising a pharmaceutically acceptable carrier or diluent and a protein allergen selected from the group consisting of recombinant Amb a I.1, recombinant Amb a I.2, recombinant Amb a I.3, recombinant Amb a I.4 and recombinant Amb a II, said protein allergen which binds immunoglobulin E to a substantially lesser extent than the corresponding native protein allergen.
 - 41. An isolated peptide comprising at least two different regions, each region comprising at least one T cell epitope of *Amb a* I.1, said regions each comprising all or a portion of an amino acid sequence selected from the group consisting of: amino acid residues 48-107 (SEQ ID NO:117); amino acid residues 171-216 (SEQ ID NO:118); amino acid residues 278-322 (SEQ ID NO:119); and amino acid residues 331-377 (SEQ ID NO:120).
 - 25 42. An isolated peptide comprising at least two different regions, each region comprising at least one T cell epitope of *Amb a* 1.1, said regions each comprising all or a portion of an amino acid sequence selected from the group consisting of: amino acid residues 57-101 (SEQ ID NO:121); amino acid residues 182-216 (SEQ ID NO:122); amino acid residues 280-322 (SEQ ID NO:123); and amino acid residues 342-377 (SEQ ID NO:124).
 - 43. An isolated peptide of claim 41 wherein the regions comprise amino acid residues 171-216 (SEQ ID NO:118) of Amb a I.1 and amino acid residues 331-377 (SEQ ID NO:120) of Amb a I.1.
 - 44. An isolated peptide of Amb a I.1 comprising at least one T cell epitope of Aml a I.1, said peptide comprising all or a portion of an amino acid sequence selected from the group consisting of: amino acid residues 48-107 (SEQ ID NO:117); amino acid residues

171-216 (SEQ ID NO:118); amino acid residues 278-322 (SEQ ID NO:119); and amino acid residues 331-377 (SEQ ID NO:120).

- 45. A therapeutic composition comprising at least one peptide of Amb a I.1 and a pharmaceutically acceptable carrier or diluent, said at least one peptide comprising a sufficient percentage of the T cell epitopes of Amb a I.1 such that upon administration of the peptide to an individual sensitive to ragweed pollen, T cells of the individual are tolerized to Amb a I.1.
- 10 46. A method of treating sensitivity to ragweed pollen in an individual, comprising a therapeutically effective amount of a composition of claim 45.
 - 47. A method of preventing sensitivity to ragweed pollen in an individual, comprising administering to the individual a therapeutically effective amount of a composition of claim 45.
 - 48. A composition comprising at least two peptides, said peptides each comprising at least one T cell epitope of *Amb a* I.1, said composition selected from the group consisting of:

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- a) AMB 1-2.1 and AMB 4-9.1 (SEQ ID NO:86 and SEQ ID NO:110);
- b) AMB 1-2.1, AMB 2-38.1 and AMB 4-9.1 (SEQ ID NO:86, SEQ ID NO:141, and SEQ ID NO:110);
- c) AMB 1-2.1, AMB 2-38.1, AMB 4-9.1 and AMB 2-4.1 (SEQ ID NO:86, SEQ ID NO:141, SEQ ID NO:110, and SEQ ID NO:90);
- d) AMB 1-2.1, AMB 2-38.1, AMB 4-9.1, AMB 2-4.1, and AMB 3-5.1 (SEQ ID NO:86, SEQ ID NO:141, SEQ ID NO:110, SEQ ID NO:90, and SEQ ID NO:102);
- e) AMB 1-2.1, AMB 2-36.1 and AMB 4-9.1 (SEQ ID NO:86, SEQ ID NO:139, and SEQ ID NO:110);
- f) AMB 1-2.1, AMB 2-36.1, AMB 4-9.1 and AMB 2-4.1 (SEQ ID NO:86, SEQ ID NO:139, SEQ ID NO:110, and SEQ ID NO:90); and
- g) AMB 1-2.1, AMB 2-36.1, AMB 4-9.1, AMB 2-4.1, and AMB 3-5.1 (SEQ ID NO:86, SEQ ID NO:139, SEQ ID NO:110, SEQ ID NO:90, and SEQ ID NO:102).
- 49. A peptide of claim 8 in which at least one amino acid residue not essential for T cell receptor interaction has been substituted with another amino acid residue.

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- 50. A peptide of claim 49 wherein said at least one amino acid residue not essential to T cell receptor interaction has been substituted with an amino acid residue selected from the group consisting of a methyl amino acid, alanine and glutamic acid.
- 51. A peptide of claim 8 wherein at least one amino acid residue essential for T cell receptor interaction has been substituted with a similar amino acid residue.
- ____ 52. A peptide of claim 8 which has been modified by a process comprising the steps of:
 - a) determining the amino acid residues of said peptide essential to T cell receptor recognition;
 - b) substituting at least one amino acid residue not essential to T cell receptor recognition with an amino acid residue, selected from the group consisting of a methyl amino acid, alanine and glutamic acid; and optionally
 - c) substituting at least one amino acid residue essential for T cell interaction with a similar amino acid residue.
 - 53. A peptide of claim 8 modified by a process comprising the steps of:
 - determining the amino acid residues of said peptide which are essential to T cell receptor recognition;
 - b) substituting at least one amino acid residue essential for T cell interaction with a similar amino acid residue; and optionally
 - c) substituting at least one amino acid residue not essential to T cell receptor interaction with an amino acid residue selected from the group consisting of methyl amino acid, alanine and glutamic acid.
 - 54. A method for determining in an individual the presence of immunoglobulin E specific for a ragweed protein allergen, and the ability of T cells of the individual to respond to T cell epitope(s) of ragweed protein allergen, comprising the steps of:
 - a) administering to an individual an Immediate Type Hypersensitivity test utilizing Amb a I.1 or a portion thereof which binds immunoglobulin E specific for the human T cell reactive feline protein;
 - b) determining whether a specific Immediate Type Hypersensitivity reaction occurs;
 - c) administering to an individual prior to, simultaneously with, or subsequent to administration of the Immediate Type Hypersensitivity test in step (a), a Delayed Type

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Hypersensitivity test utilizing a peptide derived from Amb a I.1 which has human T cell stimulating activity and which does not bind immunoglobulin E specific for Amb a I.1, or if binding to said immunoglobulin E occurs, such binding does not result in release of mediators from mast cells or basophils in a substantial percentage of a population of individuals sensitive to the human T cell reactive feline protein; and

- d) determining whether a specific Delayed Type Hypersensitivity reaction occurs.
 - 55. An antibody specifically reactive with a peptide of claim 1.
 - 56. The antibody of claim 55 which is a monoclonal antibody.
 - 57. A T cell clone specifically reactive with a peptide of claim 1.
- 15 58. A soluble T cell receptor specifically reactive with a peptide of claim 1.
 - 59. An antibody specifically reactive with the T cell receptor of claim 58.
- 60. A multipeptide formulation suitable for pharmaceutical administration, comprising:at least two peptides derived from a ragweed pollen protein allergen or a combinantion thereof, having human T cell stimulating activity in an *in vitro* T cell proliferation assay and soluble at a physiologically acceptable predetermined pH and an excipient.
- The multipeptide formulation of claim 60 said at least two peptides are lyophilized.
 - 62. The multipeptide formulation of claim 60, wherein the excipient comprises sodium phosphate.
 - 63. The multipeptide formulation of claim 60, wherein the excipient comprises mannitol.
- 64. A multipeptide formulation comprising peptides Amb 1-2.1 (SEQ ID NO:86)
 35 Amb 2-36.1 (SEQ ID NO: 139), and Amb 4-9.1 (SEQ ID NO: 110).
 - 65. The multipeptide formulation of claim 64 further comprising sodium phosphate and mannitol.

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- 66. The multipeptide formulation of claim 64 wherein said formulation is in lyophilized form.
- 67. A therapeutic composition comprising at least one peptide of Amb a I.1,, at least one said peptide containing a sufficient percentage of the T cell epitopes present in ragweed pollen protein allergen to induce T cell nonresponsiveness in a substantial percentage of a population of individuals sensitive to ragweed pollen protein allergen.
- one said peptide comprising a sufficient percent of the T cell epitopes present in Amb a 1.1 to induce T cell nonresponsiveness in a substantial percent of the population of individuals sensitive to ragweed protein allergen.
- 69. A method for treating sensitivity to a ragweed pollen protein allergen in a subject sensitive to said allergen comprising orally administering at least one isolated and purified ragweed pollen protein allergen or any portion thereof.
 - 70. The composition of claim 48 comprising AMB 1-2.1, AMB 2-36.1 and AMB 4-9.1 (SEQ ID NO:86, SEQ ID NO:139, and SEQ ID NO:110)
 - 71. A method of treating sensitivity to ragweed pollen protein allergen in an individual comprising administering subcutaneously, in non-immunogenic form, a therapeutically effective amount of the mutipeptide formulation of claim 65.

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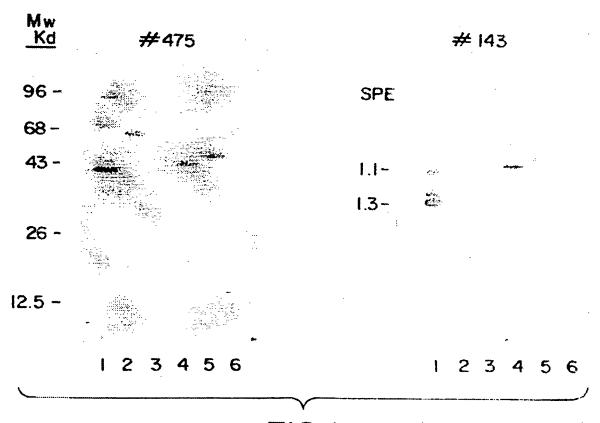
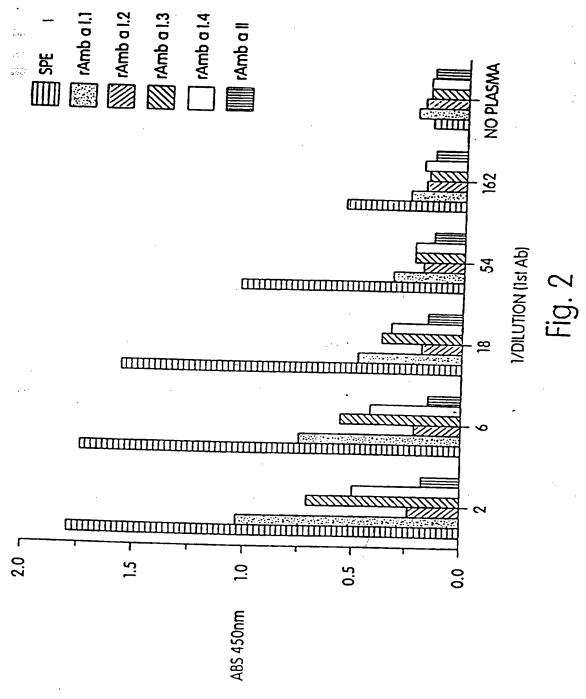


FIG. I



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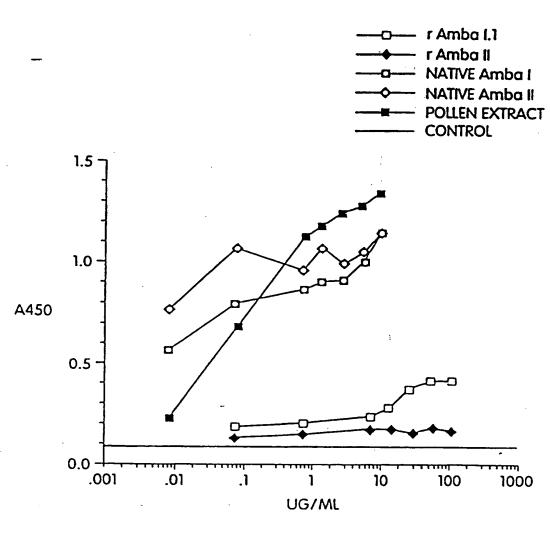


Fig. 3

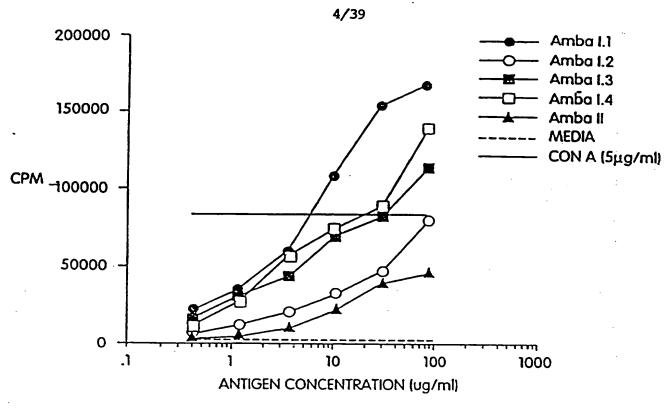
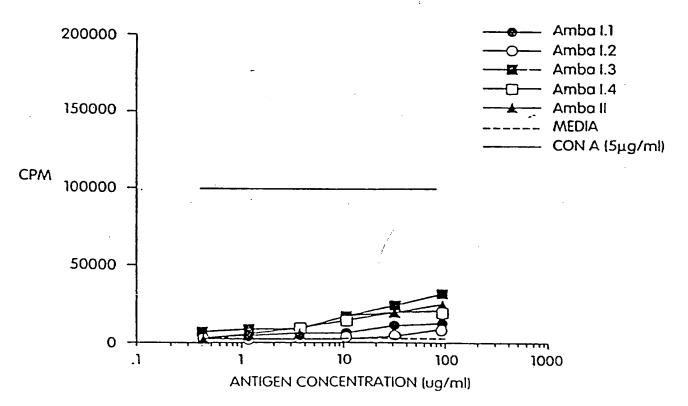


Fig. 4A



SUBSTITUTE SHEET (RULE 26) Fig. 4B

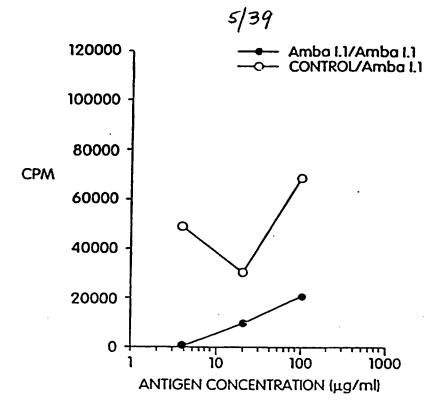


Fig. 5A

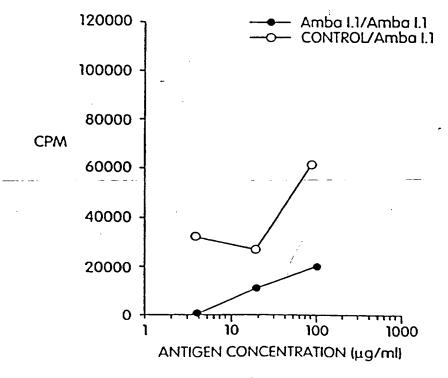
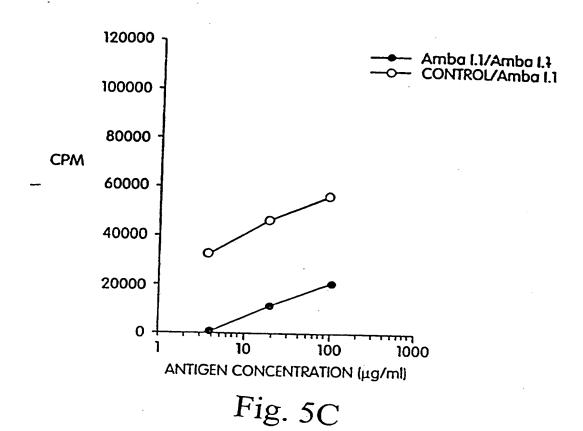


Fig. 5B

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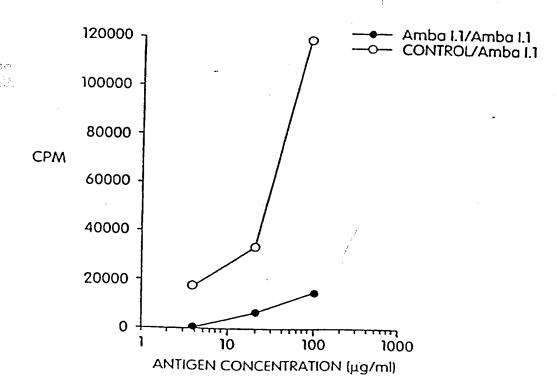


Fig. 5D

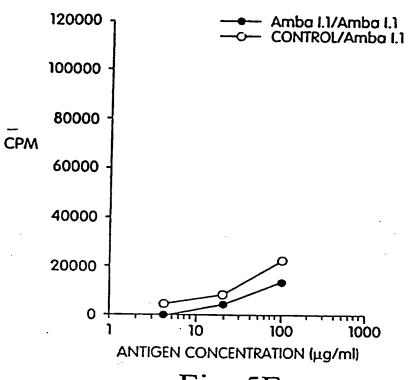
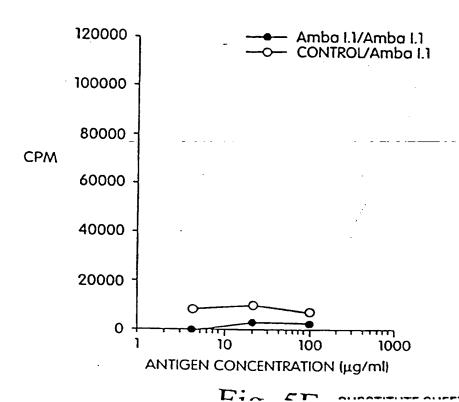
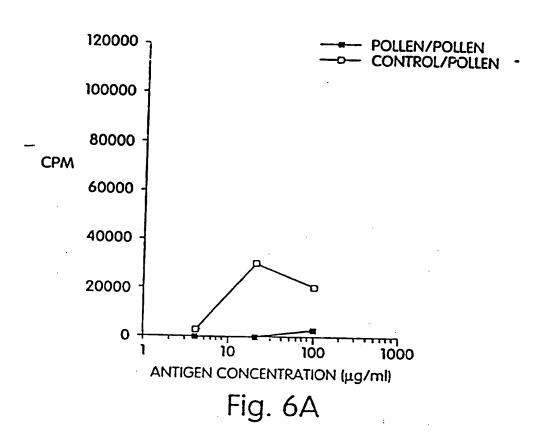


Fig. 5E

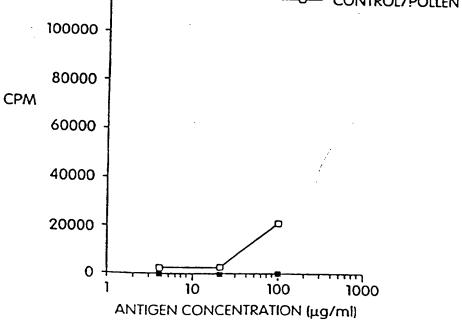


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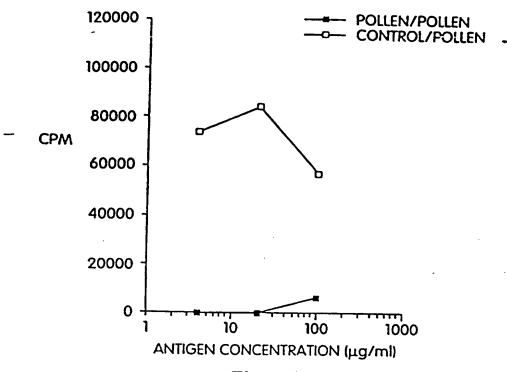
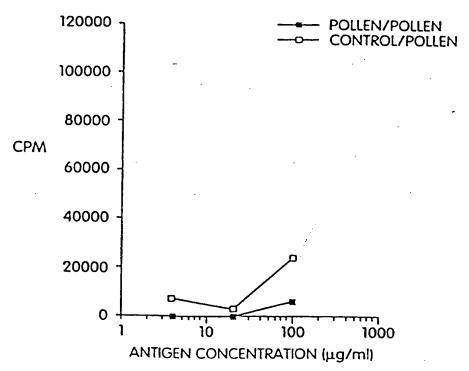
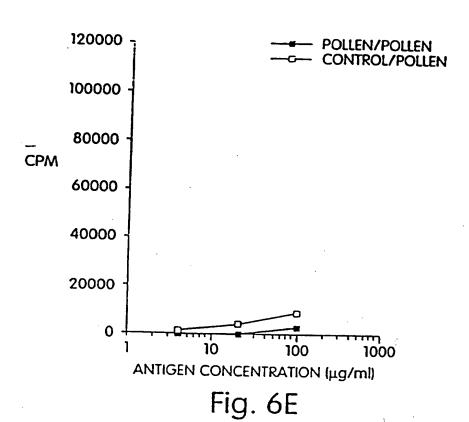


Fig. 6C



SUBSTITUTE SHEET (RULE 26)9. 6D



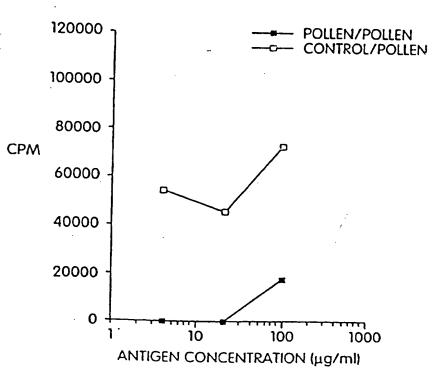


Fig. 6F

24.E(38-61)42S44K 24.1(38-61)42del.

57.1(22-51)35,

67.15(8-34)

RAE RAE

67.1(8-34)

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PVRSAEDLQEILP*VNETRR*LTTSGAYNI npregtlrfgaaqnrplwiiferdmvirl AYNIIDGCWRGKADWAENRKALADCAQG YILYFTLALVTLLQPVRSAEDLQEILP YILYFTLALVALLQPVRSAEGVGEILP ETRRSLKTSGAYNIIDGCWRGKAD ETRR*LTTSGAYNIIDGCWRGKAD **LYTVTSELDDDVANPKEGTL [YSVTSELDDDVANPKEGTL WIIFERDMVIRLDKEMVVNS** RGKADWAENRKALADCAQG LDKEMVVNSDRTIDGRGAK LNQELVVNSDKTIDGRGVK GFGKGTVGGKDGDIYSVT GFGKGTVGGKDGDIYTVT KALADCAQGFGKGTVGG KDGDIYTVTSELDDDVA YTVTSELDDDVANPKE KDGDIYSVTSELDDDVA YTVTSELDDDVAN AENRKALADCAQG YTVTSELDDD GKADWAENRC GKADWAENRK

.D(84-100)90S

RAE

7.D-1(84-100)

RAE

RAE RAE

6.D(75-92)90S

6.1(75-92)

22.E-2(63-75)

RAE

5.D(67-83)

RAE

22.E-1(57-75)

3.D(58-67C)

3.1(58-67)

22.E(48-75)

RAE RAE 40.D(88-107)90S

RAE

40.1(88-107

45.15(129-147)

61.1(101-129 80.1(118-137) 45.1(129-147)

40.1-5(88-101

40.1-4(88-104

Fig. 7

69.1-1(176-198 69.1-2(181-198

RAE

45.15(129-147

RAE

75.1(138-163) 62.1(154-180)

RAE

69.1(171-198

1-3 (186-198

RAE

RAE

70.1-2(189-206

RAE RAE RAE

1-1(189-211

0.1(189-216)

1.1(208-235

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HDVKVNPGGLIKSNDGPAAPRAGSDGDA **APRAGSDGDAISISGSSQIWIDHCSLSK** WIDHCSLSKSVDGLVDAKLGTTRLTVSN FVLLFGAGDENIEDRGMLATVAFNTFTD GFTLNGVKNVIIHNINMHDVKVNPGGL **LGTTRLTVSNSLFTQHQFVLLFGAGDE** DKTIDGRGAKVEIINAGFTLNGVKNV HGFFQVVNNNYDKWGSYAIGGSASP NPGGLIKSNDGPAAPRAGSDGDA APRAGSDGDAISISGSSQIWIDH RCRHGFFQVVNNNYDKWGSYAIG FQVVNNNYDKWGSYAIGGSASP FTDNVDQRMPRCRFGFFQVV FTDNVDQRMPRCRHGFFQVV **ENQELVVNSDKTIDGRGVK** VNNNYDKWGSYAIGGSASP APRAGSDGDAISISGSSO IKSNDGPAAPRAGSDGDA LATVAFNTFTDNVDQRMP LATVAFNMFTDHVDQRMP GPAAPRAGSDGDA APRAGSDGDAISI GSYAIGGSASPT

Fig. 7 cont

64.1-3(295-306)

66.1-1(281-305

66.1(278-305)

27.15(268-287

27.1(268-287)

76.6(260-277)

RAE RAE RAE

63.1(243-270)

RAE

76.1(260-277

RAE

64.1-2(295-310) 64.1-1(295-314)

RAE RAE RAE RAE RAE RAE RAE RAE RAE RAE

64.1(295-322 73.1(313-340) 74.1(331-356) 74.1-1(335-357)

29.1-1(338-357

28+29 (338-367

13/39

KWNWRTNKDVLENGAIFVASGVDPVLTPEQ 3SYAIGGSASPTILSQGNRFCAPDERSK RFCAPDERSKKNVLGRHGEAAAESMKWN EAAAESMKWNWRTNKDVLENGAIFVA ESMKWNWRTNKDVLENGALFVAS GSYAIGGSASPTILSQGNRF **EFGWRTNKDVLENGAIFVAS** KWNWRTNKDVLENGAIFVAS **LENGAIFVASGVDPVLTPEQ** GVDPVLTPEQSAGMIPAEPG SAGMIPAEPGESALSLTSSA PGESALSLTSSAGVLSC WRTNKDVLENGAIFVAS LENGAIFVASGVDPVLT GSYAIGGSASPTILSO GVDPVLTPEQSAGMIP SSAGVLSCQPGAPC NKDVLENGAIFVAS ENGAIFVASGVDP GVDPVLTPEQSA /LENGAIFVAS ENGAIFVASG GVDPVLTPE 29.1 (338E-357) 339F340G

29.1-4(347-357 28.1-3(348-358)

RAE RAE

RAE

28.1-2(348-361 28.1-1(348-364

RAE

RAE

RAE RAE 20.1-3(358-366

RAE

RAE RAE RAE

20.1(358-377 28.1(348-367

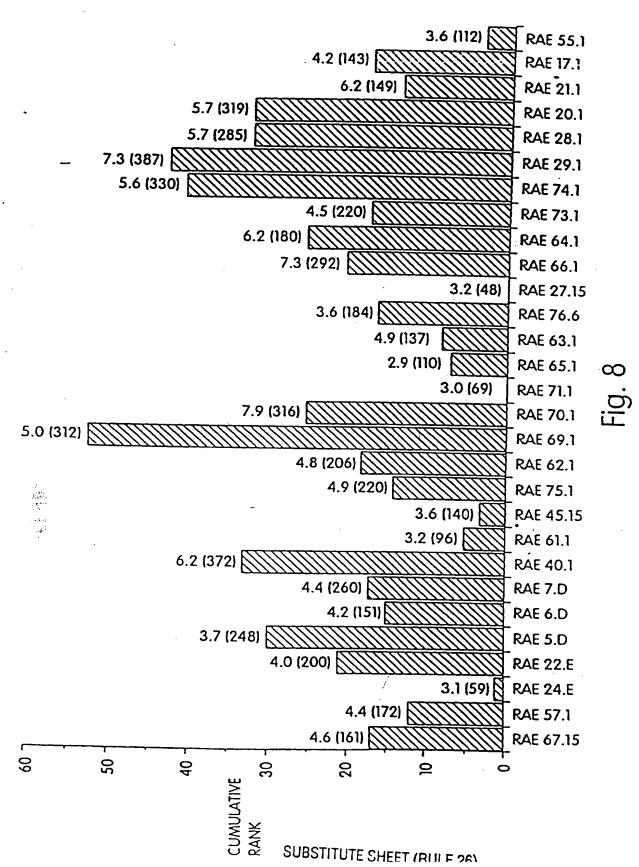
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1 (368-387

17.1(375-392 55.1(384-398

RAE

29.1-2(341-357 29.1-3 (344-357 Fig. 7 cont



AYNIIDGCWRGKADWAENRKALADCAQG HDVKVNPGGLIKSNDGPAAPRAGSDGDA **APRAGSDGDAISISGSSQIWIDHCSLSK** RCRHGFFQVVNNNYDKWGSYAIGGSASP HGFFQVVNNNXDKWGSXAIGGSASP NPGGLIKSNDGPAAPRAGSDGDA APRAGSDGDAISISGSSQIWIDH IYTVTSELDDDVANPKEGTL RGKADWAENRKALADCAQG GFGKGTVGGKDGDIYSVT IKSNDGPAAPRAGSDGDA APRAGSDGDAISISGSSQ KALADCAQGFGKGTVGG KDGDIYSVTSELDDDVA **LYTVTSELDDDVANPKE** TYTYTSELDDDVAN AENRKALADCAQG GPAAPRAGSDGDA APRAGSDGDAISI (YTVTSELDDD 7.D(84-100)90S 69.1-3(186-198 0.1-2(189-206) 69.1-1(176-198 69.1-2(181-198 0.1-1(189-211 66.1-2 (284-305 0.1-3(189-201 40.1-5(88-101) 6.D(75-92)90S 40.1-4(88-104 22.E-1(57-75 22.E-2(63-75) 40.1-6(88-98) 0.1(189-216)6.1(278-305)69.1(171-198) 40.1(88-107) 22.E(48-75) 5.D(67-83) RAE RAE

FQVVNNNYDKWGSYAIGGSASP

66.1-3(287-305

RAE RAE RAE RAE RAE RAE

64.1(295-322)

64.1-1(295-314) 64.1-2(295-310)

64.1-3(295-306

74.1(331-356)

74.1-1(335-357

29.1-2(341-357 29.1-3(344-357 29.1-4(347-357 28.1-3(348-358 28.1-2(348-361 28.1-1(348-364

> RAE RAE RAE

28+29 (338-367

16/39

KWNWRTNKDVLENGAIFVASGVDPVLTPEQ GSYAIGGSASPTILSQGNRFCAPDERSK EAAAESMKWNWRTNKDVLENGAIFVA ESMKWNWRTNKDVLENGAIFVAS GSYAIGGSASPTILSQGNRF KWNWRTNKDVLENGAIFVAS GVDPVLTPEQSAGMIPAEPG VNNNYDKWGSYAIGGSASP LENGAIFVASGVDPVLTPEQ WRTNKDVLENGAIFVAS **LENGAIFVASGVDPVLT** GSYAIGGSASPTILSQ GVDPVLTPEQSAGMIP NKDVLENGALFVAS LENGAIFVASGVDP GSYAIGGSASPT GVDPVLTPEQSA LENGAIFVASG VLENGAIFVAS GVDPVLTPE

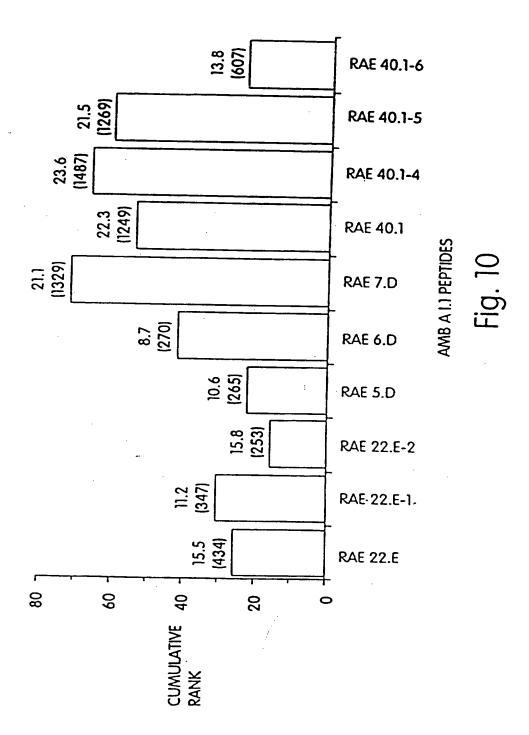
Fig. 9 cont

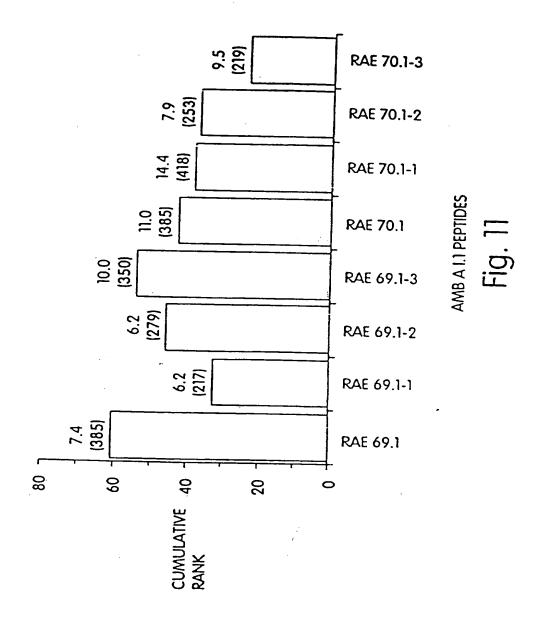
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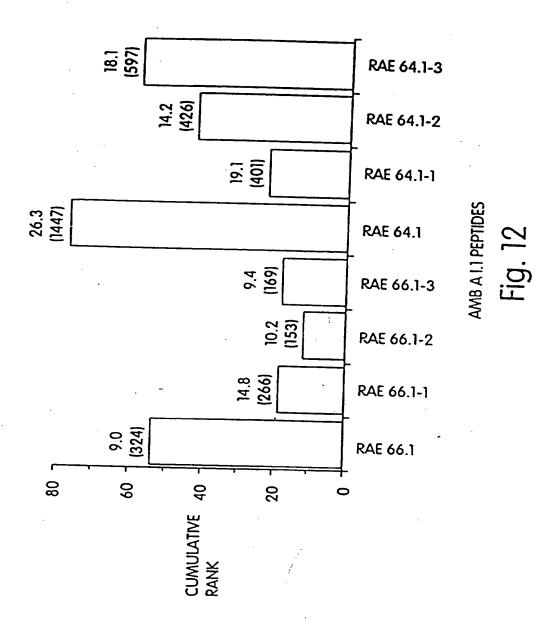
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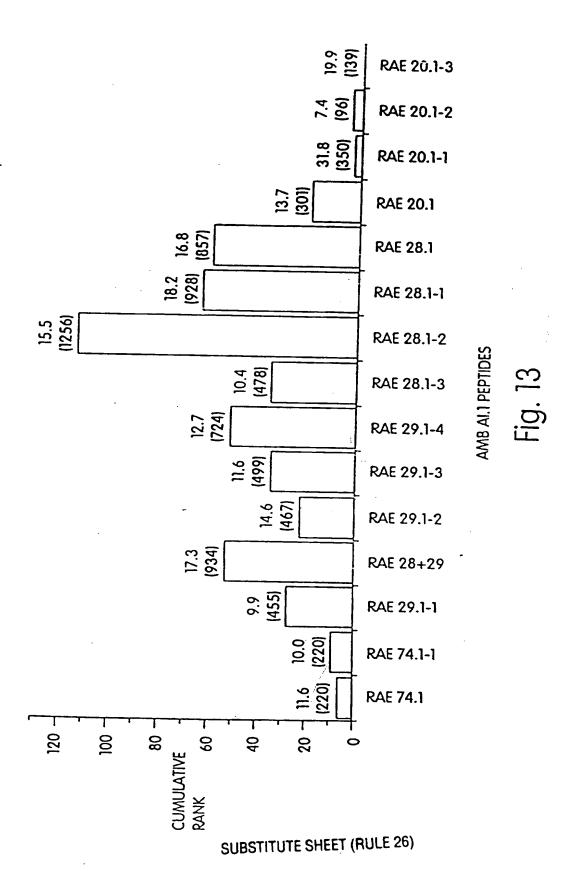
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RAE









KSNDGPAAPRAGSDGDAISISGSSQIWIDHCSLSK KKAPRAGSDGDAISISGSSQIWIDHCSLSKK KKAPRAGSDGDAISISGSSQIWIDHSSLSKK KKAPRAGSDGDAISISGSSQIWIDHLSLSKK KKAPRAGSDGDAISISGSSQIWIDHESLSKK KSNDGPAAPRAGSDGDAISISGSSQIWIDH RGKADWAENRKALADCAQGFGKGTVGGK **APRAGSDGDAISISGSSQIWIDHCSLSK** GFGKGTVGGKDGDIYTVTSELDDDVAN KKAPRAGSDGDAISISGSSQIWIDHKK **AENRKALADCAQGFGKGTVGGKDGD** ENNRQALADCAQGFAKGTYGGKWGD KSNDGPAAPRAGSDGDAISISGSSQ KGTVGGKDGDIYTVTSELDDDVAN KGTYGGKWGDVYTVTSNLDDDVAN APRAGSDGDAISISGSSQIWIDH **AENRKALADCAQGFGKGTVGGK** KSNDGPAAPRAGSDGDAISISG GPAAPRAGSDGDAISISGSSQ GKDGDIYTVTSELDDDVAN GPAAPRAGSDGDAISISG

> 2-4.1(182-203) 2-3.1(182-206) 2-5.1(182-211) 2-6.1(186-216) 2-2.1(186-203)

1-2.15(78-101

-4.15(63-87)

-3.1(83-101)

-1.1(75-101

1-5.1(63-84) 1-4.1(63-87)

AMB AMB AMB Fig. 14

2-9.1KK(189-216)K, S212 2-10.1KK(189-216)K, L212

2-8.1KK(189-216)K

70.1-1(189-211) 2-7.1KK(189-211)

70.1(189-216)

2-1.1(186-206

MB2-11.1KK(189-216)K, E212

2-1.15(186-206)

AMB AMB AMB AMB AMB

3-5.1(280-306 3-4.1(283-306) 3-3.1(295-318) 3-2.1(295-320) 3-1.1(295-322) 22/39

-3.15(347-373

AMB

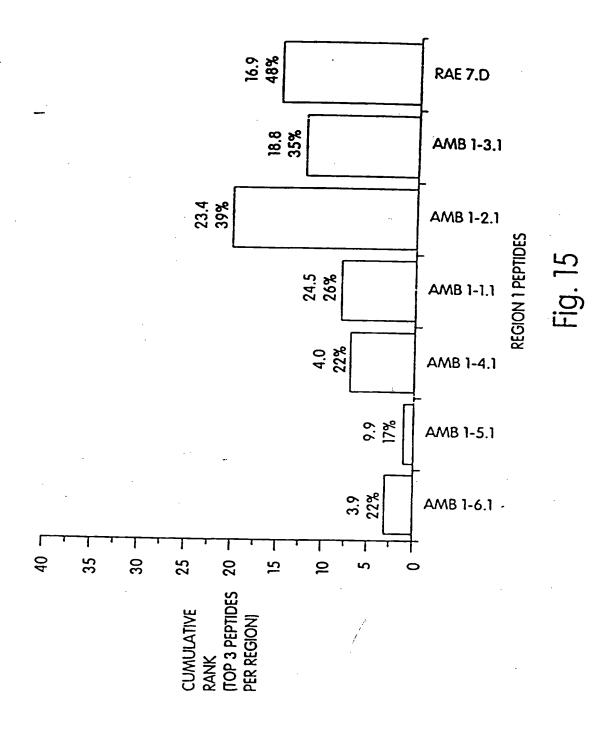
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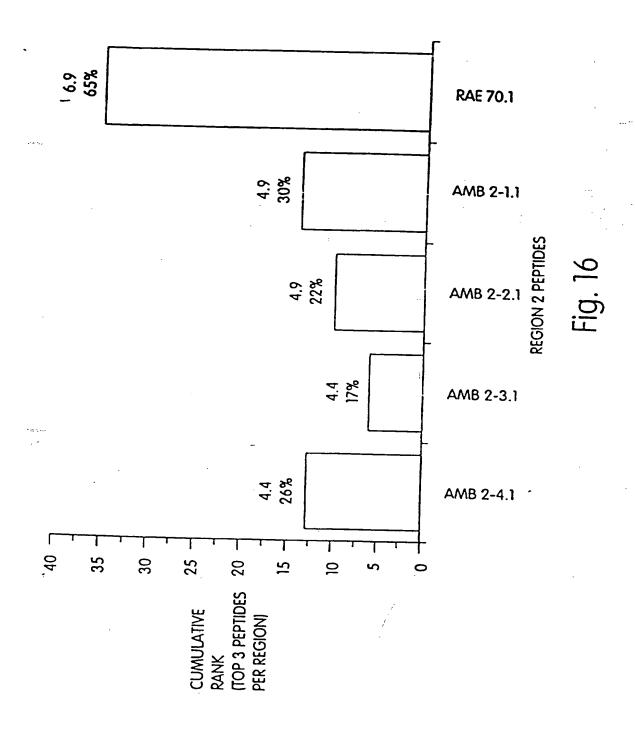
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> 3-4.15(283-306 3-1.15(295-322

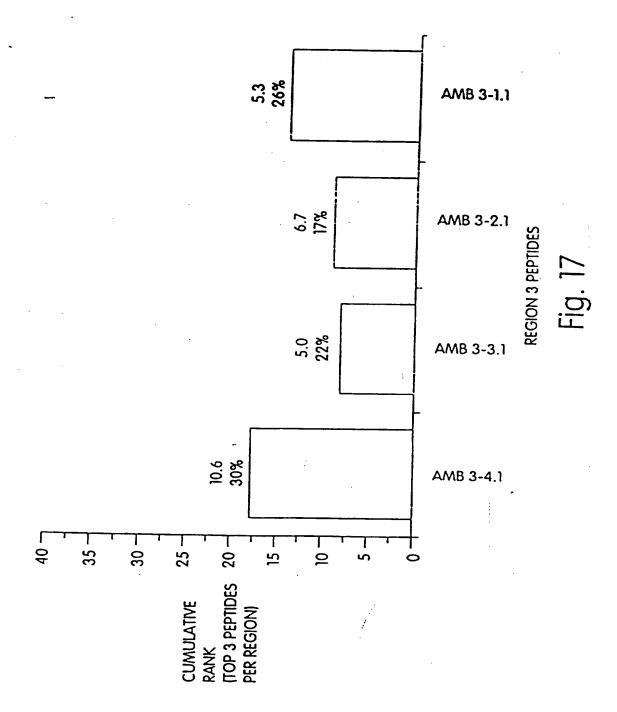
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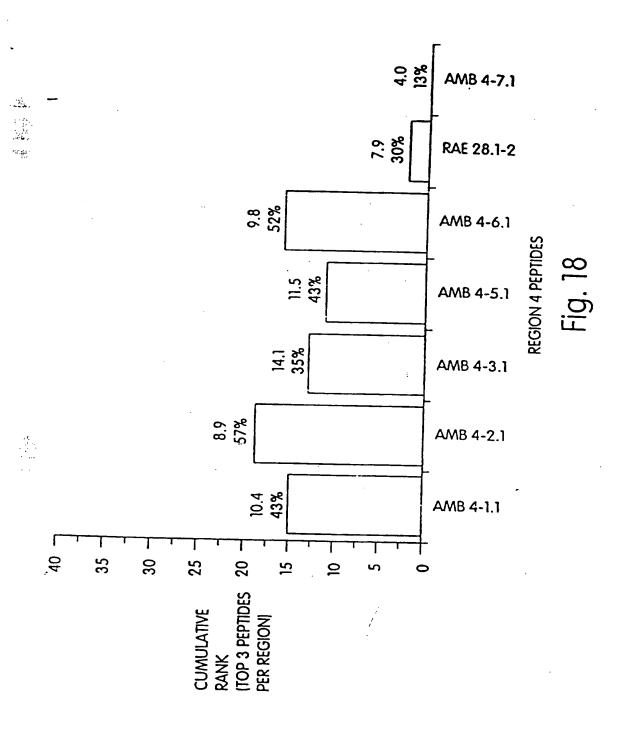
AMB AMB AMB AMB AMB AMB Fig. 14 cont



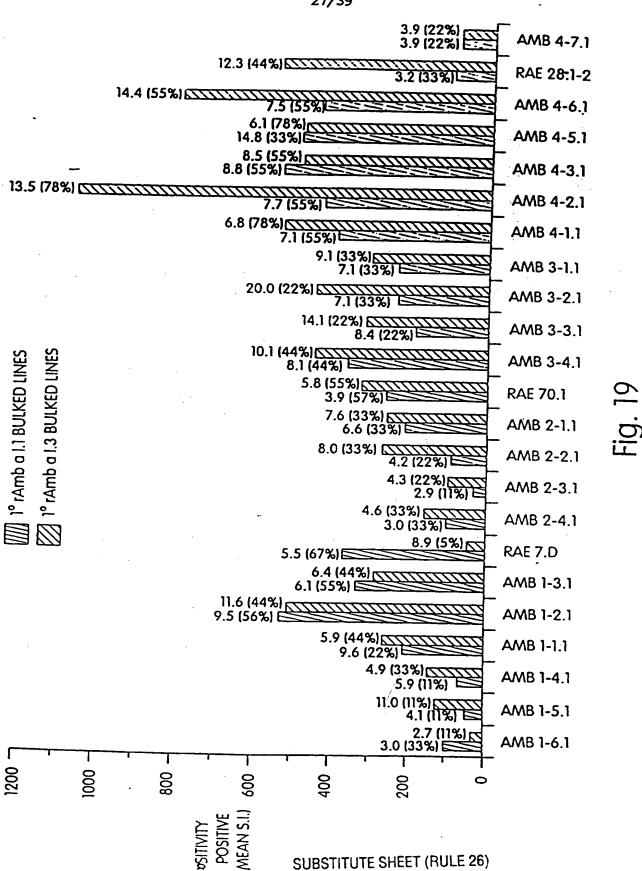


SUBSTITUTE SHEET (RULE 26)

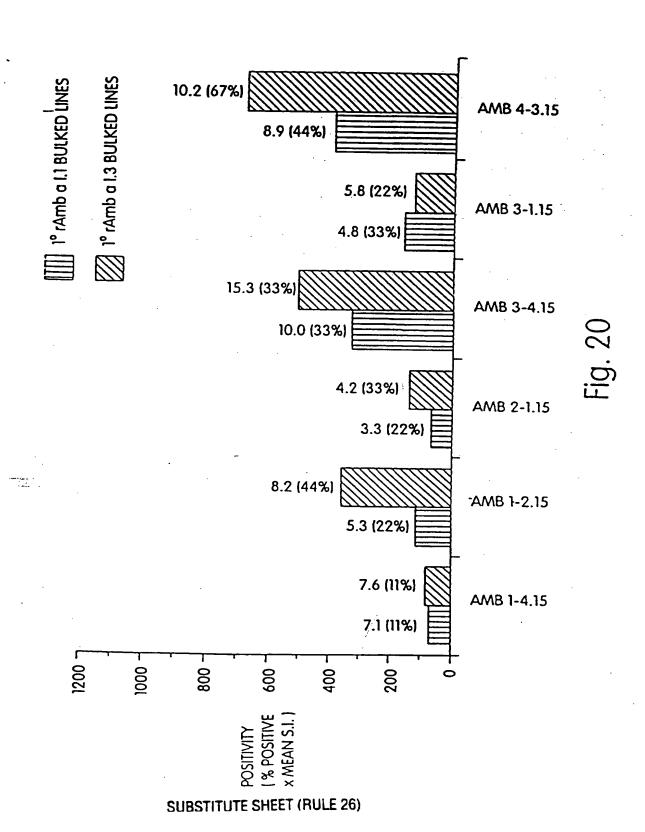




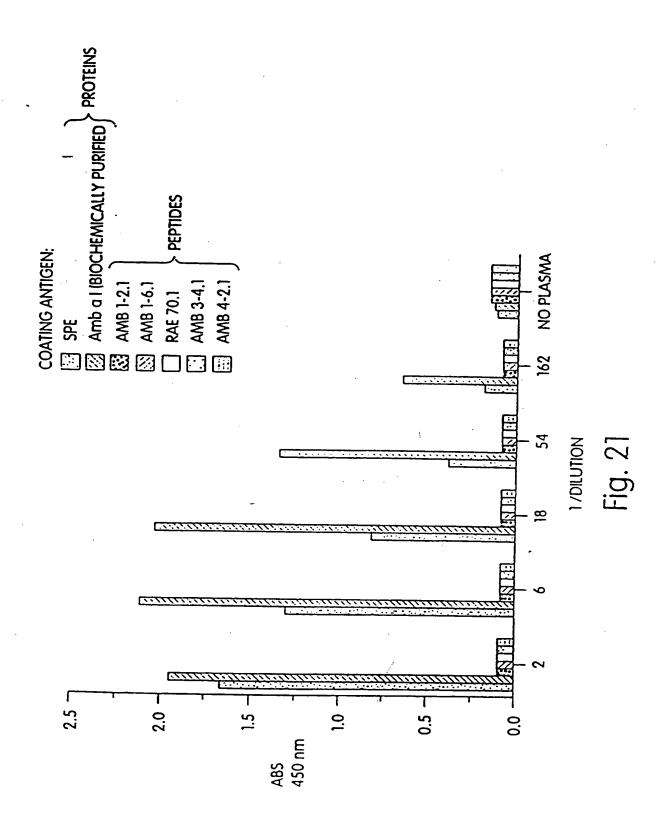




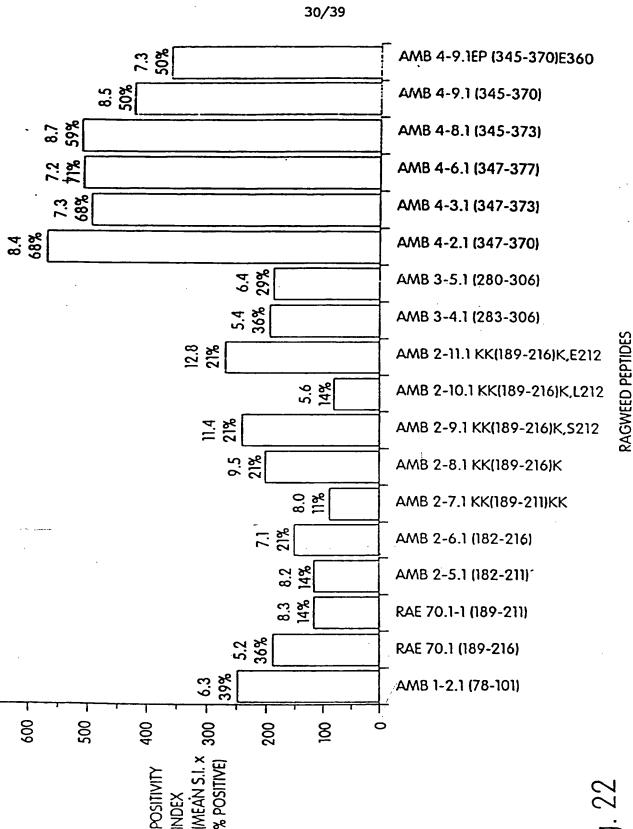
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SUBSTITUTE SHEET (RULE 26)



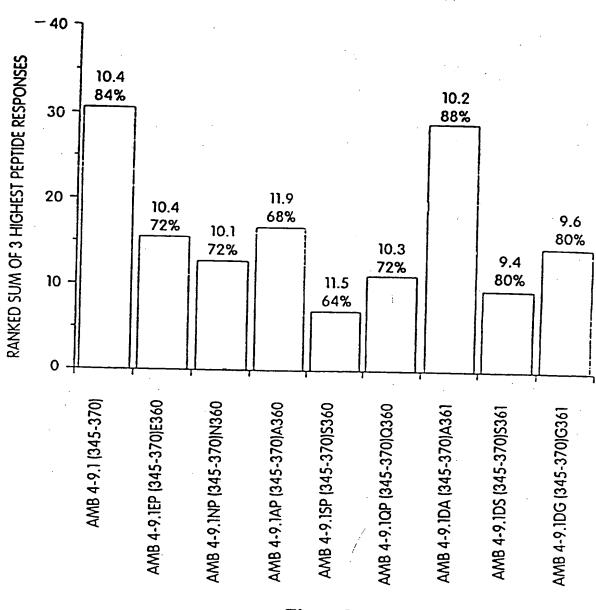
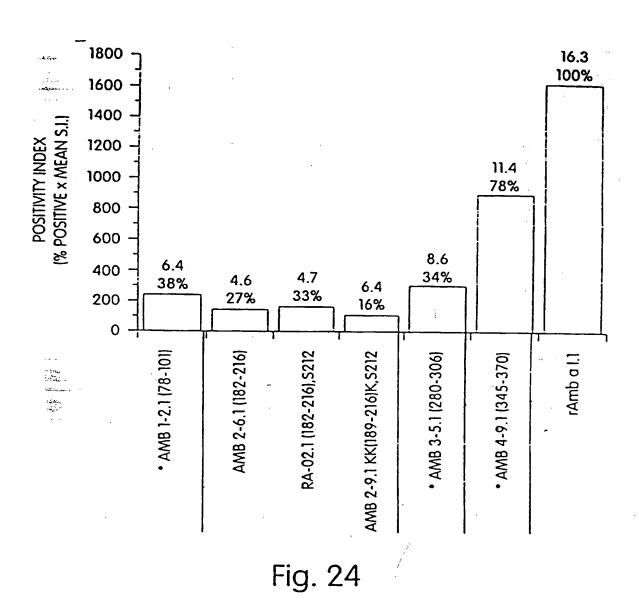


Fig. 23



NAME	SEQUENCE	CRUDE YIELD
ORIGINAL	-SDGDAISISGSSQIWIDHCSLSKS	
Amb2-18.1	SGSSQIWIDHSSLSKS	79%
Amb2-19.1	SISGSSQIWIDHSSLSKS	67%
Amb2-20.1	DAISISGSSQIWIDHSSLSKS	80%
Amb2-21.1	SDGDAISISGSSQIWIDHSSLSKS	87%
Amb2-22.1	SESGSSQIWIDHSSLSKS	84%
Amb2-23.1	SKSGSSQIWIDHSSLSKS	87%
Amb2-26.1	DKSGSSQIWIDHSSLSKE	
Amb2-28.1	DKSISGSSQIWIDHSSLSKE	98%
Amb2-30.1	DKESGSSQIWIDHSSLSKE	
Amb2-32.1	DKSGSSQIWIDHSSLSKK	85%
Amb2-33.1	DKESGSSQIWIDHSSLSKEK	107%
Amb2-34.1	SGSSQIAIDHSSLSKS	
Amb2-35.1	DKESGSSQIAIDHSSLSKSE	
Amb2-36.1	DKESGSSQIWIDHSSLSKEKD	
Amb2-37.1	SQIWIDHSSLSKS	
Amb2-38.1	DKESISGSSQIWIDHSSLSKEKD	

Fig. 25

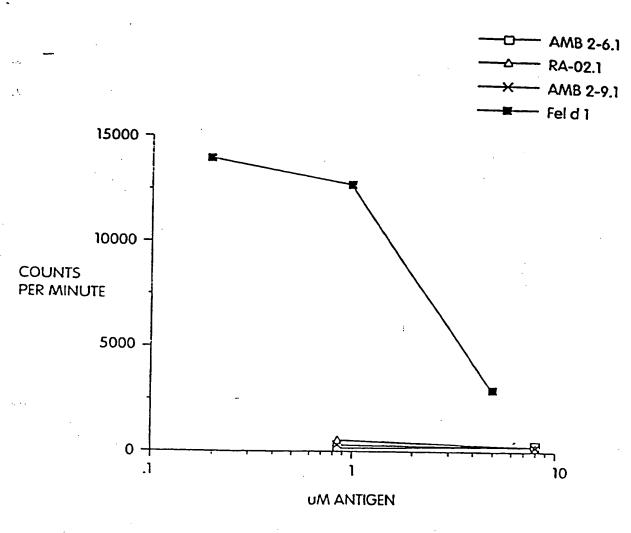
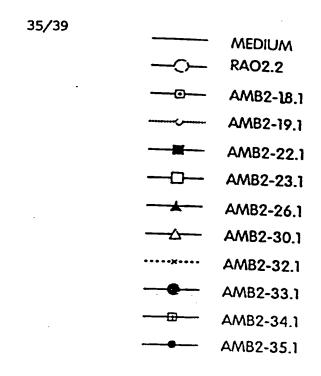


Fig. 26



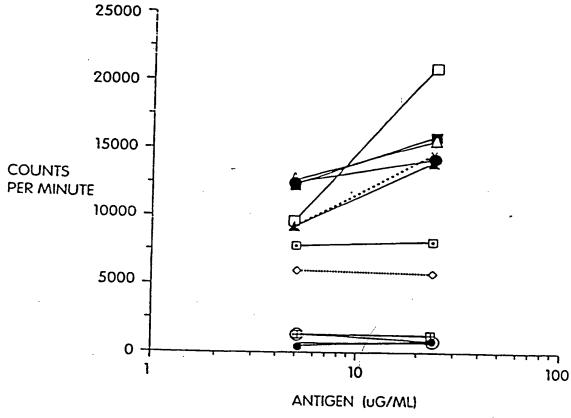


Fig. 27

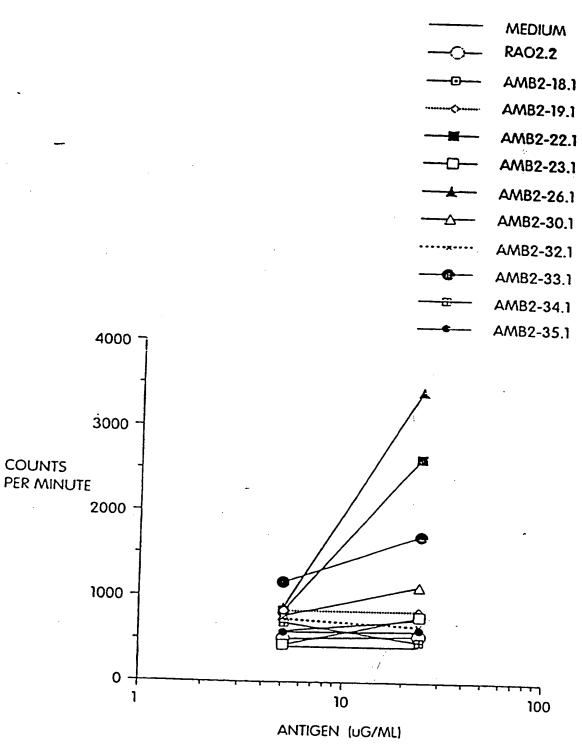


Fig. 28

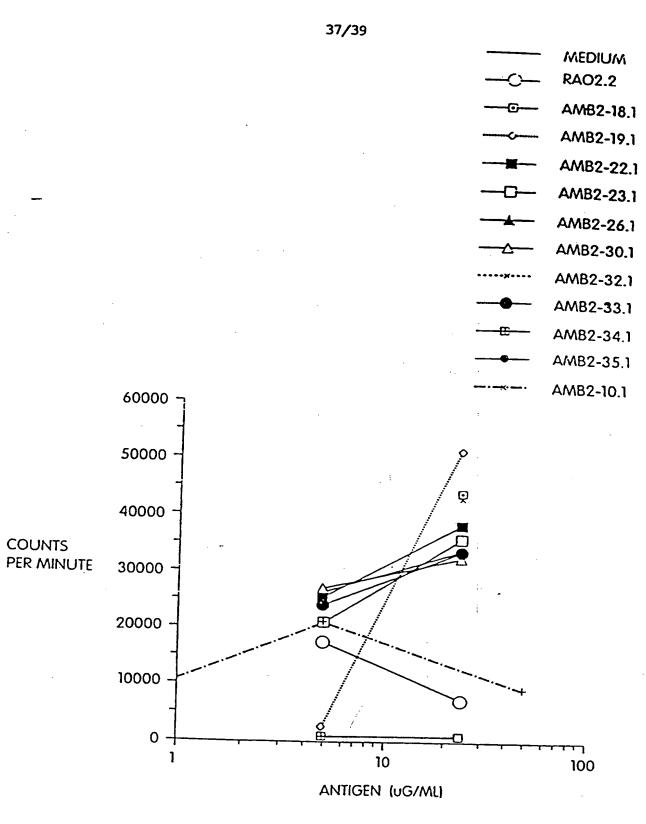
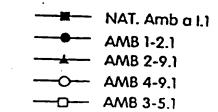


Fig. 29



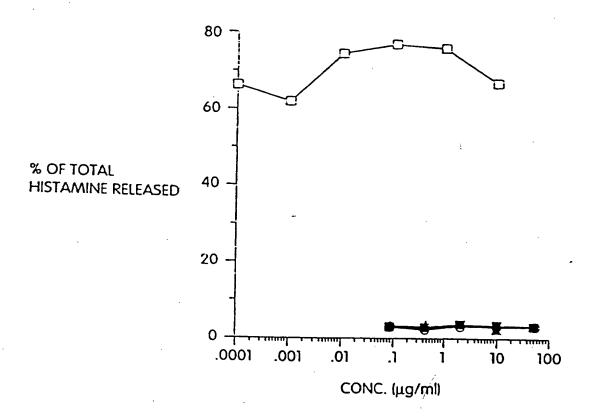
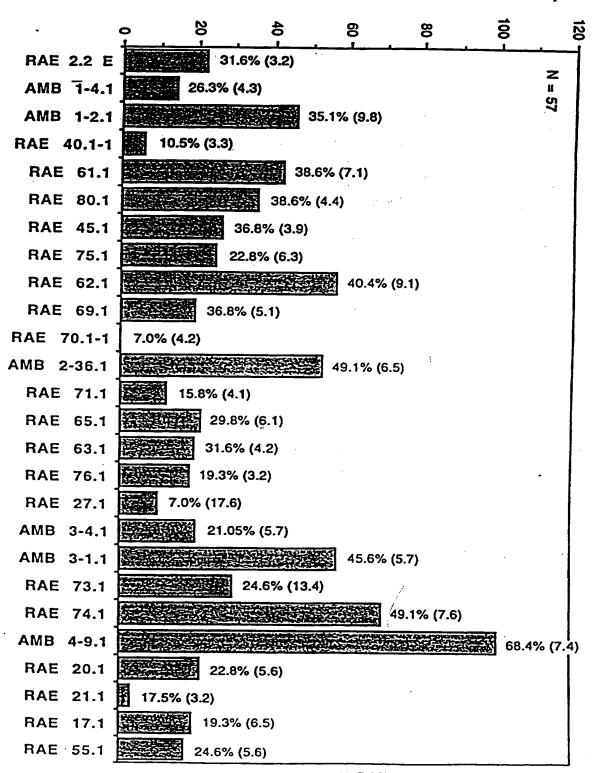


Fig. 30

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CUMULATIVE RANK SUM (TOP 5 RESPONSES) -



INTERNATIONAL SEARCH REPORT

Inter • Application No PCT/US 95/14362

		PCI	/03 95/14362
IPC 6	SIFICATION OF SUBJECT MATTER C12N15/29 C07K14/415 A61K	(39/36 G01N33/68	
According	to International Patent Classification (IPC) or to both national	al classification and IPC	
	S SEARCHED		
Minimum IPC 6	documentation searched (classification system followed by cla C12N C07K A61K	assafication symbols)	
Document	stion searched other than minimum documentation to the exten	nt that such documents are included in	the fields searched
Electronic	data base consulted during the international search (name of d	ata base and, where practical, search te	rms used)
	·		
C. DOCUM	MENTS CONSIDERED TO BE RELEVANT	· · · · · · · · · · · · · · · · · · ·	
Category *	Citation of document, with indication, where appropriate, o	f the relevant passages	Relevant to claim No.
X	WO,A,93 21321 (IMMULOGIC PHAR CORPORATION) 28 October 1993	MACEUTICAL	1-71
	see the whole document	•	
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Furth	er documents are listed in the continuation of box C.	X Patent family members a	re listed in annex.
	egories of cited documents :	"T" later document published after	r the international filing date onflict with the application but
conside E° carlier d	int defining the general state of the art which is not red to be of particular relevance locument but published on or after the international	cited to understand the princi invention	iple or theory underlying the
filing di L° documen	ate nt which may throw doubts on priority claim(s) or	"X" document of particular releva cannot be considered novel o involve an inventive step who	nce; the claimed invention or cannot be considered to the document is taken alone
citation documents	s cited to establish the publication date of another or other special reason (as specified) nt referring to an oral disclosure, use, exhibition or	"Y" document of particular releva cannot be considered to invo- document is combined with o	ince; the claimed invention lve an inventive step when the one or more other such docu-
otner in P° documer	seans In the published prior to the international filing date but	ments, such combination being in the art.	ng obvious to a person skilled
	an the priority date claimed	*& document member of the sam Date of mailing of the interna	
14	March 1996	20	5. 03.9 6
ame and ma	ailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2	Authorized officer	
	NL - 2280 HV Ripwik Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+ 31-70) 340-3016	Cupido, M	

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ional application No.

PCT/US 95/14362

INTERNATIONAL SEARCH REPORT

MARKET TO THE SECOND

B x I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)	
This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following re-	asons:
IIIB discussion see at 1-p and a see	
1. X Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claims 27,28,32-36,39,40,46,47,54,69 and 71 are dire to a method of treatment or diagnostic method practised on the human/ body, the search has been carried out and based on the alleged effect the compositions.	
 Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to s because they relate to parts of the international application that do not comply with the prescribed requirements to s an extent that no meaningful international search can be carried out, specifically: 	uch
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4	(1).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)	
This International Searching Authority found multiple inventions in this international application, as follows:	
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.	
2. As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite pa of any additional fee.	yment
As only some of the required additional search fees were timely paid by the applicant, this international search repo	rt
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report restricted to the invention first mentioned in the claims; it is covered by claims Nos.:	. is
Remark on Protest The additional search fees were accompanied by the applicant's	protest
No protest accompanied the payment of additional search fees	
·	

INTERNATIONAL SEARCH REPORT

Inte Application No PCT/US 195/14362

Patent document cited in search report W0-A-9321321	Publication date	Patent family member(s)		Publication date
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		FI-A-	944697	07-12-94
	•	JP-T-	7507924	07-09-95
		NO-A-	943811	06-12-94
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